Screening for High-Risk Human Papillomavirus by Real-Time PCR and Its Association with Cytological Abnormalities of Pap Smears in Baghdad

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ABSTRACT

Human Papillomaviruses (HPVs) are known to be etiological agents of cervical cancer and have been found in 99.7% of women with high-grade (HG) cervical intraepithelial neoplasia (CIN) pre-cancer. Testing of high-risk HPV (HR-HPV) has been proposed as a way of improving cervical screening, especially for women with low-grade (LG) Papanicolaou (Pap) smears. Therefore, the present study aimed to investigate the diagnostic correlation of real-time PCR technique with Pap smear results for 13 genotypes of HR-HPV in 415 of suspected women. Pap smear results showed the suspected women with abnormal and normal cytological changes were 79.3% (329/415) and 20.7% (86/415), respectively. In contrast, real-time PCR results showed that women with or without abnormal changes had HR-HPV DNA positive were 50.5% (166/329) and 10.5% (9/86), respectively. Thus, Pap smear results had no sensitive to predicate risky for LG-CIN women were observed. According to the real-time PCR results, the incident of HG or cancer was more associated with HR-HPV. Our findings indicated the real-time PCR test with the cytological test as a routine practice for women with LG-CIN reduces the incidence of HG or cancer. After that, the prospective studies should be appropriated to determine HR-HPV genotypes and viral load (copies/cell) with real-time PCR test as predicate risk-factor.

Key words: Human Papillomavirus (HPV), Real-Time PCR, Papanicolaou (Pap) smear.

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INTRODUCTION

Human Papillomaviruses (HPVs) are known to be etiological agents of cervical cancer and have been found in 99.7% of women with high-grade (HG) cervical intraepithelial neoplasia (CIN) pre-cancer (such as CIN II/III and cancer) [1]. Testing of high-risk HPV (HR-HPV) has been proposed as a way of improving cervical screening, especially for women with low-grade (LG) (such as CIN I) Papanicolaou (Pap) smears [2]. The Pap smear test, as the most widely used cancer screening test in the world, is cost effective and organized screening provided a decline in cervical cancer mortality [3]. Testing of HPV DNA was recently introduced into clinical practice with the aim of identifying women at risk of cervical cancer, and encouraging results when are used in conjunction with the traditional cytology screening method. Many molecular methods for HPV DNA testing are currently available. Polymerase chain reaction (PCR) based methods amplifying nucleic acids of HPV are commonly used because of the limited sensitivity of Pap smear test leading to sample preparation and interpretation problems, insufficient serological tests and impracticality of in vitro cultures [4]. Detection of HPV DNA, especially in latent infections, can be helpful in detecting cancer and precursor lesions [2]. Real-time PCR techniques have been developed to quantify HPV DNA in clinical samples. Also, real-time PCR is extremely reproducible, rapid and pertinent in a clinical setting [5]. Our observation suggests the HPV infections more frequency among Iraqi women. Therefore, the present study aimed to investigate the diagnostic correlation of real-time PCR technique with Pap smear results for 13 genotypes of HR-HPV in HG or LG pre-cancer. This could be considered very important in the early diagnosis and or prevention of cervical cancer.

MATERIALS AND METHODS

Studying groups:
In current study, 415 of suspected women with different ages from 15-58 old years were selected. Nearly all women had not been taken at least one Pap smear previously, who suffering from chronic inflammation and abnormal appearance of their cervix by gynecologist examination. Meanwhile, 25 health women as a control group were mainly depending on the absence of clinical symptoms and cervical cytological changes. All cervical samples were attending Women Health Center- Al-Alwia hospital and Al-kadhemia teaching hospital in Baghdad during the 2011.

**Pap smear:**
It is a simple and quick test. A special stick or brush was used to take a few cells from inside and around the cervix of women suspected. The sample, which contains cells and mucus, is then smeared onto a glass slide and send to a lab for a pathologist examination after Pap stain procedure [6].

**Cervical scrape samples:**
Cervical scrape samples were collected in liquid transport medium such as PBS solution; cells were eluted from the spatula in 3ml of PBS [7]. The specimens could be transported at 4-30 °C and stored at -20°C for HPV DNA isolation.

**Real-time PCR HPV test:**
Real-time fluorescent 5’ exonuclease (Taq Man probe) assay was employed for HPV detection from the range of HR-HPV DNA (13 genotypes) most frequently found in different grades of cervical intraepithelial neoplasia, cervical tumors and health controls. HybriBio real-time PCR HPV detection kit (HybriBio co., LTD, Hong kong) was used for *in vitro* detection of HR-HPV DNA (16,18,31,33,35,39,45,51,52,56,58,59 and 68) from scraped cervical cells according to supplied company instructions (www.hybribio.com) for DNA extraction and PCR amplification.

**DNA extraction**
The DNA was extracted from 1ml of the cervical sample by centrifuging at 13000 rpm for 1 min. The pellet was washed in 500 µl PBS solution, centrifuging at 13000 rpm for 1 min, and then discarded the supernatant clearly. The pellet was resuspended in 50 µl lysis solution (HybriBio kit), mixes thoroughly, and then boiled for 10 min in water bath. Finally, the mixture was centrifuged at 13000 rpm at 10 min, pipettes the supernatant to a new tube and stored at -20 °C for amplification.

**PCR amplification**
The PCR amplification was performed in 20 µl total volume containing 17.5 µl PCR mix, 0.5 µl (5U/ µl) Taq polymerase and 2 µl DNA template. Taq Man probes are labeled hydrolysis probes. They consisted of an 18-22 bp oligonucleotides, which are labeled with a reporter dyes as 6-carboxyfluorescein (FAM) at the 5’ end. Amplification and detection were performed with an ABI prism 7500 Sequence Detection System (Applied Biosystem). The amplification ramp included initial hold step of 10 min at 95 °C to release the activity of the DNA polymerase. The hold step was followed by a two-step cycle consisting of 10 s at 95 °C and 50 s at 58 °C for a total of 45 cycles. Tubes that contained all PCR components but without template DNA (denoted no-template control [NTC reactions] were used to ensure that the reagents were free of contamination [8].

**Statistical analysis**
All values were calculated according to the positive results as percentages (%). Differences between multiple groups and control group were analyzed by chi-square (X²) test. A value of P<0.5 was considered statistically significant.

**RESULTS**

**Pap smear**
Results showed the suspected women with abnormal and normal cytological changes were 79.3% (329/415) and 20.7% (86/415), respectively. Women with abnormal changes were comprised of 84.5% (278/329) as LG-CIN and 15.5% (51/329) of them in HG lesions (Table 1). Thus, Screen test is important to reduce LG-CIN or it progression.

From the cytological test, 11.8% (6/51) of women with HG-CIN were constituted as squamous cell carcinoma that produced by HR-HPV, and 22.3% (39/175) of women with LG and HG-CIN was associated with koilocytosis criteria as the most important pathognomonic sign of HPV presence [9, 10].

**Real-time PCR**
High-risk HPV DNA prevalence in all suspected women was 42.17% (175/415) which appeared in pre-cancer and cancer lesions as 94.85% (166/175) and 3.43% (6/175), respectively. Our finding showed 50.5% (166/175) of women with abnormal changes and 10.5% (9/86) of women with normal cytological in Pap smear were positive for HR-HPV DNA (Fig. 1), (Table 1). The chi-square test used to
check for correlations between HR-HPV DNA present and cytological changes was not considered significant (P > 0.05).

According to real-time PCR results, the incident of HG-CIN or cancer were more associated with HR-HPV which constituted 100% (6/6) of women with squamous cell carcinoma and 62.22% (28/45) of women were suffered from moderate to sever dysplasia had HR-HPV DNA positive. Highest frequency of HR-HPV DNA positive was 47.5% (132/278) of women with LG (Fig. 2). On the other hand, the HR-HPV DNA observation of the total risky group in women with LG, HG-CIN and healthy women in cytology were 75.43% (132/175), 19.43% (34/175) and 5.14% (9/175), respectively.

From the results above, the sensitivity and specificity of cytopathology method for HR-HPV diagnosis were 50.5% and 89.5%, respectively. Moreover, real-time PCR results showed more compatibility with Pap smear results for HG-CIN than LG-CIN diagnosis. Detecting of HR-HPV DNA in conjunction with Pap smear may improve the sensitivity of identifying LG pre-cancer that are associated with high-risk of progression.

Fig. 1: Distribution of HR-HPV DNA positive among infected women according to cytological changes appearance with Pap smear test.

Fig. 2: Percentage of HR-HPV DNA positive among infected women according to risk groups of each cytological level.
Table 1. Distribution of Studying groups according to Pap smear results and Real-Time PCR HR-HPV DNA occurrence

<table>
<thead>
<tr>
<th>Changes type</th>
<th>No. of cases</th>
<th>Pap smear results</th>
<th>Positive (%) in Real-Time PCR HR-HPV DNA test</th>
</tr>
</thead>
<tbody>
<tr>
<td>CIN I (LG-CIN)</td>
<td>36</td>
<td>Binucleation, koilocytosis and hyperkeratosis associated with mild dysplasia</td>
<td>18 (4.3%)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Cervicitis and mild dysplasia</td>
<td>108 (2.5%)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Cervicitis and hyperplasia</td>
<td>25 (5.1%)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Monilliasis and mild dysplasia</td>
<td>61 (1.7%)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Trichomonas and mild dysplasia</td>
<td>48 (1.1%)</td>
</tr>
<tr>
<td>CIN II</td>
<td>36</td>
<td>Cervicitis and moderate dysplasia</td>
<td>23 (5.5%)</td>
</tr>
<tr>
<td>CIN III</td>
<td>9</td>
<td>Cervicitis, Sever dysplasia and koilocytosis in three of them</td>
<td>5 (1.2%)</td>
</tr>
<tr>
<td>SCC</td>
<td>6</td>
<td>Malignant cells (Squamous cell carcinoma)</td>
<td>6 (1.4%)</td>
</tr>
<tr>
<td>Normal cytology</td>
<td>86</td>
<td>Cervicitis and no cellular changes</td>
<td>9 (2.2%)</td>
</tr>
<tr>
<td>Control</td>
<td>25</td>
<td>No Changes and asymptoms</td>
<td>0</td>
</tr>
<tr>
<td>Total</td>
<td>440</td>
<td></td>
<td>42.17% (175/415)</td>
</tr>
</tbody>
</table>

**DISCUSSION**

Real-time PCR the most advanced and sensitive of the molecular methods was used in the present study. Our finding showed HR-HPV DNA presence in women with or without abnormal changes was more efficiency than cytological testing alone for clinical evidence. In this study, we gave a hint that consecutive detection of HPV DNA is due to either ongoing viral replication, reactivation of latent infections or new infections. The present study observed that HPV DNA percentages in studied Iraqi women were similar to those reported previously in many studies like 28.8% - 61.3% of cases have been reported in women with abnormal cytology [11, 12, 13, 14]. On the other hand, the HPV prevalence in women with normal cytology was estimated in 5.2% - 20% of cases [14, 15, 16, 17]. These variations among studies are most likely reflecting the differences in the population studied with respect to risk factors for exposure to HPV and methods of evaluations. Although, women were had abnormal cytological changes, real-time PCR was showed those 77.9% (74/95) of LG and 20% (3/15) of HG-CIN as negative result for HR-HPV DNA. This observation suggested the presence of either other microorganism like herpes virus, parasites and fungi which induce these cellular changes or women negative to real-time PCR probes for HR-HPV, may have missed which are positive to other HPV types not included in the probes or samples with very low number of virus DNA copies. Discordance between cytology and molecular detection found in women could be explained as a false-positive diagnosis of LSIL [18]. Meanwhile, real-time PCR results showed HR-HPV DNA was positive in women with normal cytology that similar to Arora et al. [12], suggested that the cases who are HPV positive for any of the high risk types but have concurrently negative Pap smear or no visible lesions should be followed up more frequently than the HPV negative cases as they have a substantially greater risk of developing an abnormal smear. Since morphological changes on cytology such as koilocytosis are not specific for oncogenic HPV [19], our finding showed that eight women had all criteria for HPV infection (such as koilocytosis) was HR-HPV DNA negative. Prevalence of HR-HPV DNA was more frequency in HG, thus a distinction between the pathological diagnoses of HG-CIN (CIN II/III) may be important in future clinical studies of real-time PCR HR-HPV screening and the prevention of cervical cancer.

**CONCLUSION**

Our findings indicated the real-time PCR test with the cytological test as a routine practice for women with LG-CIN reduces the incidence of HG or cancer. After that, the prospective studies should be appropriated to determine HR-HPV genotypes and viral load (copies/cell) with real-time PCR test as predicate risk-factor.
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REFERENCES