



Studies on the Calcium Oxalate Crystals of Some Selected Aroids (Araceae) in Eastern India

Sk. Md. Abu Imam Saadi and Amal Kumar Mondal*

Plant Taxonomy, Biosystematics and Molecular Taxonomy Laboratory,
 Department of Botany and Forestry
 Vidyasagar University, Midnapore- 721 102, West Bengal, India
 Email:saadivu@gmail.com
 *Email: amalcaebotvu@gmail.com

ABSTRACT

The presence or absence of micro-characters in plant system like calcium oxalate crystals has been used for understanding the evolutionary relationships of plant species. The size and appearance of calcium oxalate crystals (COC) can differ within families, genus, and species and these characteristics might be genetically controlled. We have studied the calcium oxalate crystals in the different plant parts (leaves, stems, petiole, corm, and root) of some selected species belonging to the family Araceae. The selected plants belonged to different habitats like marshy, semi aquatic, terrestrial and were mostly herbs, shrubs, and climbers. Among the selected species, two species are edible and economically important. Edibility of petiole, leaves, stems and corms depends upon the frequency and intensity of the calcium oxalate crystals. Two types of crystals were observed which were mostly species specific. The frequency of crystals is probably related with the habit, habitat and also the environmental conditions.

KEYWORDS: Calcium Oxalate Crystals (COC), Araceae, edibility, evolutionary relationships

INTRODUCTION

Needle shaped calcium oxalate crystals called raphides have been upon as an important indicator of aroids. The aroids are a sub-group of Araceae which is a family of mainly herbaceous plants that are known to produce raphides in abundance. When eaten raw they cause swelling of the lips, mouth & throat. Detoxification via cooking, pounding or leaching neutralizes the chemical, hence making the aroids edible but does not destroy or degrade the raphides [1,2,3,4].

Calcium oxalate crystals occur in more than 215 higher plant families, as well as the algae, lichen and fungi, in two hydration states in plants, as the monohydrate [Whewellite ($\text{CaC}_2\text{O}_4 \cdot \text{H}_2\text{O}$)] or dehydrate [weddellite ($\text{CaC}_2\text{O}_4 \cdot 2\text{H}_2\text{O}$)] [5,6]. They can form in any organ or tissue within the plants, including in stems, petiole, leaves, roots & tubers, and have a variety of function including calcium storage, defense and providing structural strength [7,8]. According to Horner and Wagner 1995 [6], unlike phytoliths, which vary considerably in size and shape across families, calcium oxalate crystals are generally restricted to five basic morphological types – Needle shaped raphides, rectangular or pencil shaped styloids, mace head shaped aggregates called druses, block shaped aggregates called crystal sand, variously shaped prisms.

Given that the size and appearance of these crystals can differ within and between families, and that these morphological characteristics are probably under genetic control [5]. Calcium oxalate crystals may have taxonomic potential for both botanist and taxonomist [6].

Types of raphides

Anatomical study indicates that raphide morphology and formation in specific locations within a plant are genetically controlled [6]. Key attributes for differentiating raphides include size, cross-section and termination morphology, all of which appear to vary to differing degrees depending on taxa of origin.

Type I Raphides

It is the most common raphide form and consists of four-sided single crystals that have two symmetrical pointed ends. (Fig.17)

Type II Raphides:

Which are also four-sided, have one pointed and one bidentate or forked end [9] This type of raphide has so far only been recorded in a few families such as the Vitaceae[10,11]. The bidentate end is formed by crystal twinning [12](Fig.17)

Type III Raphides

The third form is crystals with six to eight sides and symmetrical pointed ends. This raphide type is known to occur in the Agavaceae[13]. Typhaceae [14] and Dioscoreaceae. (Fig.17)

Type IV Raphides

The fourth raphide form comprises twinned crystals with H-shaped cross-sections and asymmetrical ends (one wedge-shaped and the other sharply pointed) [1, 15]. (Fig.17)

MATERIAL AND METHODS

Plant material

The plant specimens belonging the family Araceae. The species identification of the selected materials was determined according to standard literature. It was done in the month February to May.

Crystal Isolation and Purification

Crystals were isolated from both fresh and dry plant specimens. However, dry material was preferred to increase crystal recovery. With the purpose of avoiding potential contamination of crystalline samples by soil particles, plant stems, leaf, petiole, root, corm or storage organ were carefully washed with abundant distilled water. After removal of needles epidermis, thin sections of plant stems, leaf, petiole, root, corm or storage organ were excised and washed several times. The raphides could be easily separated manually. Clearing technique is used to specifically locate the calcium oxalate crystals in the plant tissue. Tissue sections were macerated in water and crystals were mechanically freed with the help of dissection knives, segments were fixed in glycerine and water. After that we prepared a slide for observation. The slides were observed under light microscope (10X x 40X) [Olimpus] as well as Phase Contrast Microscope (Leica DM- 1000) for detailed analysis and obtaining better picture as well as measuring the length & breadth of raphide crystal.

RESULT

At first, we observed the shape, size and position of raphides of the selected plant materials under light microscope (10X x 40X) as well as Trinocolor microscope Leica (DM-1000). Considering measurement of the size, shape and the result of the experiment. Experiment reveals that the raphides varies between species to species like druses [Fig-12], needle shaped, H-shaped. We are observed in the leaves, stem, petiole, storage organ or corm and root. Simultaneously few are small to large. The number of raphides also varied because occurring in different environmental conditions. In case of *Alocasia indica*, we are also observed the idioblast arranged in a ring [Fig-16].

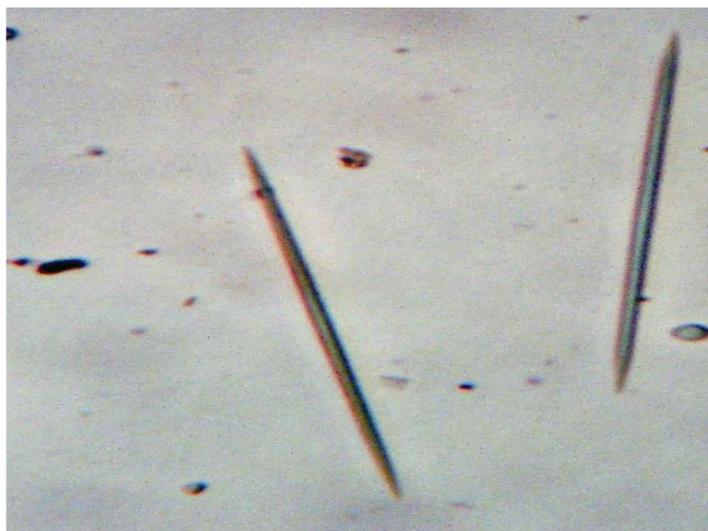


Fig.1 - Calcium oxalate crystal (COC) of *Lasia heterophylla* from petiole.

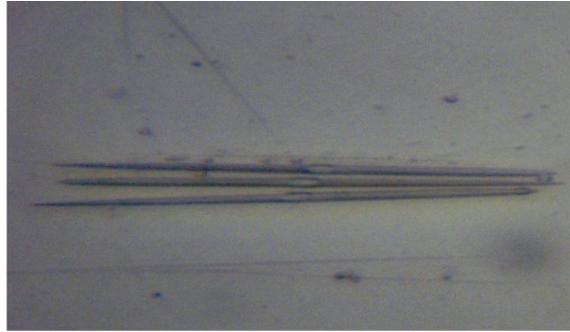


Fig.2 - calcium oxalate crystal of *Colocasia* (black) from storage organ.

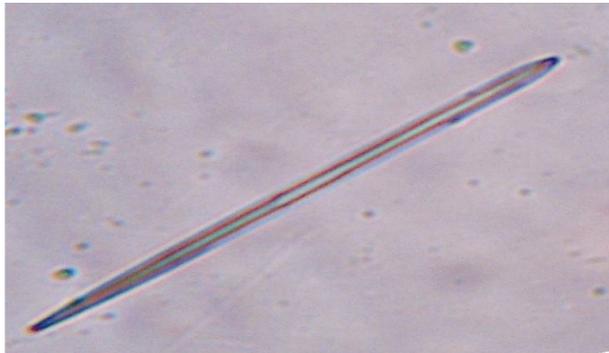


Fig.3 - calcium oxalate crystal of *Caladium schomburgkii* from root.

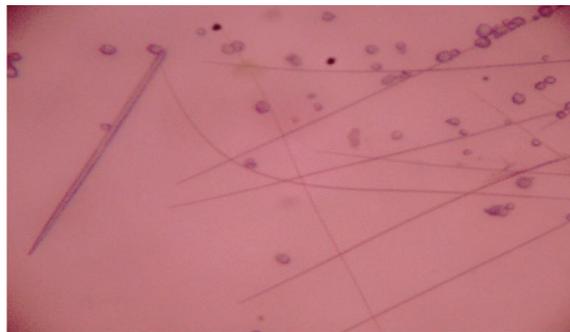


Fig.4 - calcium oxalate crystal of *Colocasia* (black spotted) from leaf.

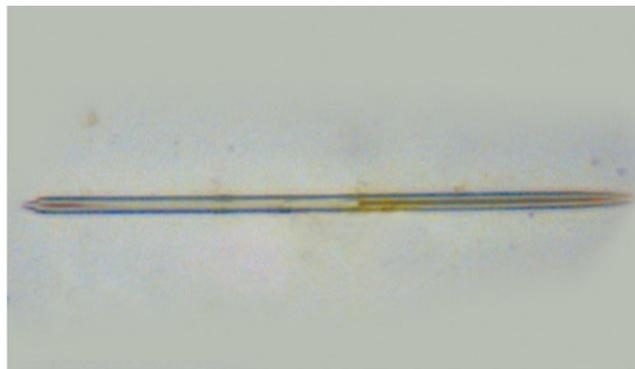


Fig.5 - calcium oxalate crystal of *Caladium schomburgkii* from root.

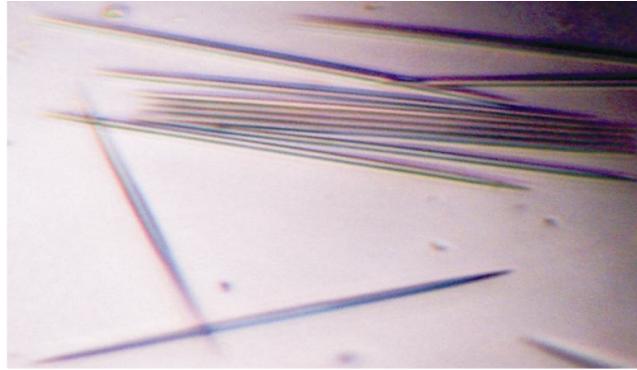


Fig.6- calcium oxalate crystal of *Lasia heterophylla* from root.

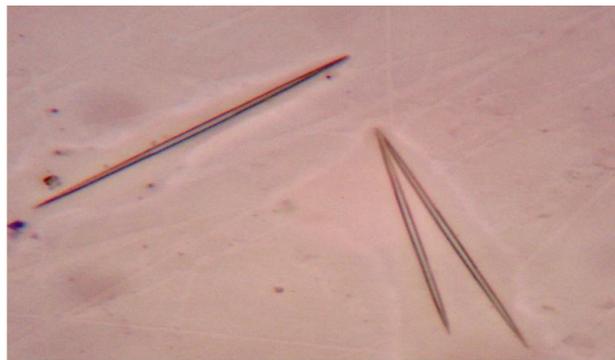


Fig.7 - calcium oxalate crystal of *Epipremnum pinnatum* from petiole.

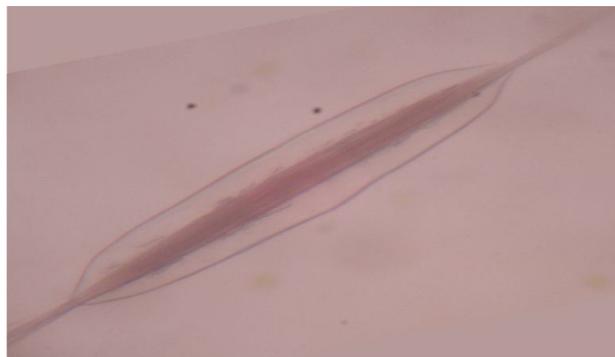


Fig.8 – Idioblast of *Colocasia* (black spotted) from leaf. COC are partially ejected through the papillae at this end.



Fig.9 – Different types of idioblast of *Alocasia indica* from petiole

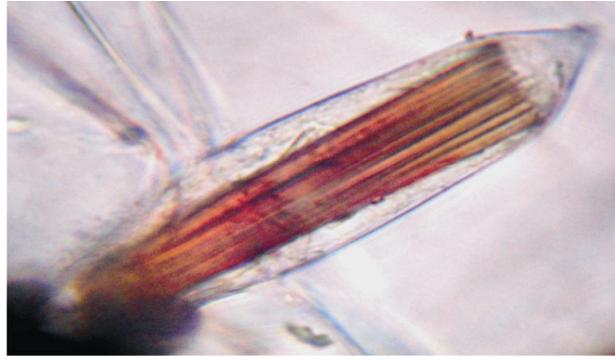


Fig.10- Different types of idioblast of *Alocasia indica* from petiole



Fig.11- Idioblast of *Colocasia* (black) from petiole.



Fig.12- Druses of *Amorphophallus sylvaticus* from leaf.



Fig.13- Idioblast of *Colocasia gigantea* from leaf.

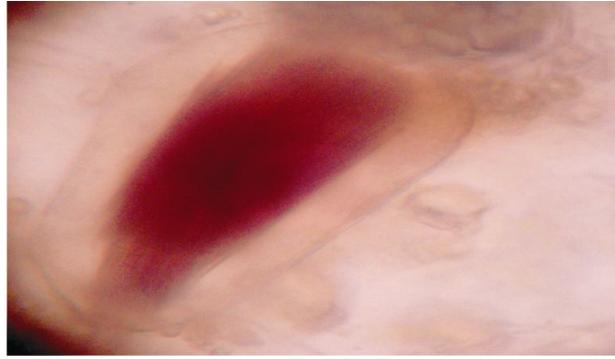


Fig.14- Idioblast of *Colocasia* (black) from leaf.

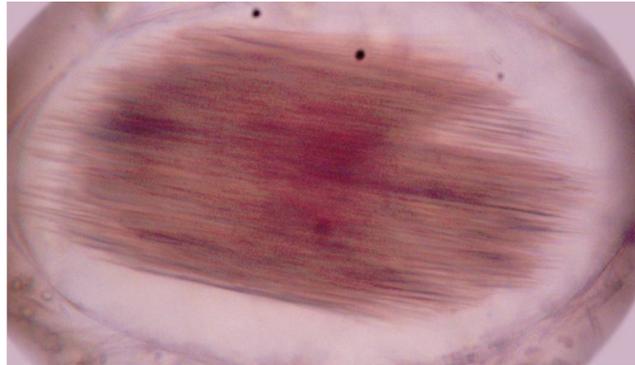


Fig.15- Idioblast of *Colocasia* (black spotted) from petiole.

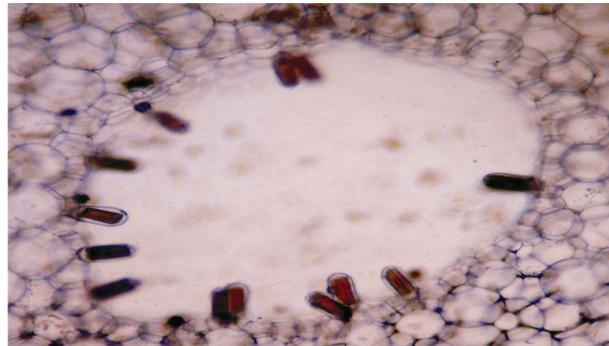


Fig.16- Idioblast of *Alocasia indica* arrangement in a ring from petiole

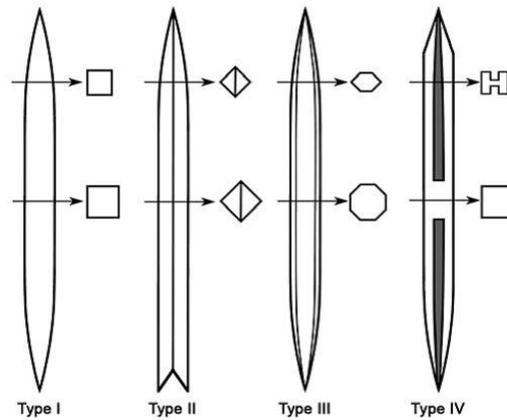


Fig.17-Diagram showing the four basic types of raphide cross sections. End forms are described in text (redrawn from Horner and Wagner 1995:58)

DISCUSSION

In case of *Colocasia esculenta* storage organ shows oblong shaped, petiole of *Colocasia gigantea* [Fig-13] *Colocasia esculenta*(black) [Fig-11], *Amorphophallus sylvaticus* shows slightly rectangular to oval shaped; petiole of *Colocasia esculenta* (black spotted) [Fig-15] shows oval shaped; petiole of *Alocasia odora* (Fig.9 – 10) leaf of *Colocasia esculenta* (black) [Fig-14] shows elliptical shaped; leaf of *Colocasia esculenta* (black spotted) [Fig-8] shows lanceolate idioblast. Our study shows, that some also possess barbs near their tips and along their edges, which occur in some other Araceae.

Type I is the most common raphides form and consist of four sided single crystals that have two symmetrical pointed ends. In our work we observed mainly that *Lasia heterophylla* [Fig-1,6], *Epipremnum pinnatum* [Fig-7] possess Type I raphides. And also petiole, root of *Diffenbachia picta*; leaf, petiole, root of *Dracontium nivosum*, storage of *Colocasia gigantea* possess Type I raphides.

Leaf, length of raphides is longer in *Lasia heterophylla* (154.332 μm), shorter in *Dracontium nivosum* (13.799 μm). Breadth of raphides is greater in *Lasia heterophylla* (4.499 μm), smaller in *Dracontium nivosum* (0.617 μm).

Petiole, length of raphides is longer in *Lasia heterophylla* (284.975 μm), shorter in *Diffenbachia picta* (21.927 μm). Breadth of raphides is greater in *Lasia heterophylla* (8.727 μm), smaller in *Dracontium nivosum* (0.981 μm)

Storage organ or corm, length of raphides is longer in *Lasia heterophylla* (89.959 μm), shorter in *Colocasia gigantea* (43.474 μm). Breadth of raphides is greater in *Lasia heterophylla* (3.464 μm), smaller in *Colocasia gigantea* (1.509 μm).

Root, length of raphides is longer in *Diffenbachia picta* (84.629 μm), shorter in *Lasia heterophylla* (16.350 μm). Breadth of raphides is greater in *Lasia heterophylla* (4.493 μm), smaller in *Diffenbachia picta* (1.573 μm).

Our observation revealed that leaf, petiole, storage organ or corm, root of *Colocasia esculenta*, *Colocasia nymphaeifolia*, *Caladium schomburgkii*[Fig-3,5], *Alocasia indica*, *Alocasia odora*, *Amorphophallus campanulatus*, *Amorphophallus sylvaticus*, *Colocasia esculenta* (black spotted) [Fig-4], *Colocasia esculenta* (black) [Fig-2,], shows H- shaped raphides. And also the leaf, petiole, root of, leaf of *Diffenbachia picta*, storage organ of *Dracontium nivosum* shows H- shaped raphides

Leaf, length of raphides is longer in *Amorphophallus campanulatus* (250.957 μm), shorter in *Diffenbachia picta* (17.992 μm). Breadth of raphides is greater in *Amorphophallus campanulatus* (6.116 μm), smaller in *Colocasia esculenta* (black spotted) (0.401 μm).

Petiole, length of raphides is longer in *Amorphophallus campanulatus* (327.892 μm), shorter in *Caladium schomburgkii* (21.08 μm). Breadth of raphides is greater in *Amorphophallus sylvaticus* (8.596 μm), smaller in *Colocasia esculenta* (black spotted) (0.309 μm).

Storage organ or corm, length of raphides is longer in *Amorphophallus campanulatus* (210.554 μm), shorter in *Colocasia nymphaeifolia* (38.445 μm). Breadth of raphides is greater in *Amorphophallus campanulatus* (5.533 μm), smaller in *Colocasia esculenta* black spotted (0.436 μm).

Root, length of raphides is longer in *Colocasia esculenta* (black spotted) (209.677 μm), shorter in *Colocasia esculenta* (14.466 μm). Breadth of raphides is greater in *Alocasia indica* (3.491 μm), smaller in *Colocasia* (black spotted) (0.310 μm).

From the above discussion we can draw the following inference that Type I and Type IV (H -shaped), both types of raphides present in the family Araceae and therefore it offer great potential for taxonomic marker. Shape, size of idioblast will be taken in consideration for taxonomic marker.

CONCLUSION

The formation of crystals idioblast is a complex process involving changes in development of cells and the formation of specific raphides structure. The presence or absence of crystals is an important character of understanding the evolutionary relationships of plant species. The specific distribution and shape of the raphides of this angiosperms are consistent the character of calcium crystals may be genus and species specific. The observation of Calcium oxalate crystals occur in 14 species of Araceae, revealed that the occurrence and shape of crystals can be useful in the intraspecific level of phylogenetic relationship.

Table: 1 Measurement of Different Types of Calcium Oxalate Crystals of Some Selected Plant Taxa (14) Including Different Plant Parts

SL. NO.	SCIENTIFIC NAME OF THE PLANT	LEAF		PETIOLE		STORAGE ORGAN OR CORM		ROOT	
		L (µm).	B(µm)	L(µm)	B(µm)	L(µm)	B(µm)	L(µm)	B(µm)
1	<i>Colocasia esculanta</i> (Linn.) Schott.	32.785	2.182	40.683	3.147	89.459	2.182	14.466	0.614
		31.732	1.753	44.965	2.182	89.900	2.794	24.877	0.617
		30.809	2.350	43.200	2.618	82.130	2.544	29.418	1.851
2	<i>Alocasia indica</i> (Roxb.) Schott.	80.419	1.301	60.093	1.799	103.264	4.023	26.874	1.745
		90.704	1.109	103.82	1.573	101.290	3.728	31.178	3.491
		91.092	1.002	56.778	2.760	101.050	4.556	27.259	2.380
3	<i>Alocasia odora</i> (Roxb.) K.Koch	88.470	1.380	57.938	1.573	86.880	2.618	37.725	1.234
		90.744	1.309	57.045	1.254	46.089	2.544	32.913	2.618
		121.090	1.809	55.544	0.967	55.081	2.045	41.234	1.799
4	<i>Caladium schomburgkii</i> Schott.	106.130	2.225	37.661	0.873	72.861	1.951	28.162	0.783
		34.580	3.491	26.036	0.783	72.625	1.234	29.279	0.873
		33.747	3.480	21.081	0.801	65.169	1.951	16.198	0.436
5	<i>Colocasia gigantea</i> Hook.f.	32.220	2.102	38.630	1.799	54.504	1.509	23.921	0.463
		39.032	1.703	36.878	1.851	60.618	2.225	21.927	0.436
		40.009	2.596	33.450	1.309	43.474	1.573	22.463	0.489
6	<i>Colocasia nymphaeifolia</i> Kunth.	81.470	1.307	72.226	0.436	128.398	2.205	39.333	1.309
		77.744	0.989	57.602	0.436	68.881	1.005	48.444	1.309
		89.090	1.333	74.179	0.617	38.445	0.873	21.818	0.873
7	<i>Epipremnum pinnatum</i> (L.)Engl.	65.571	2.350	58.456	1.380	N.S.O.F.	N.S.O.F.	24.534	2.182
		68.746	2.760	65.549	1.799			30.483	1.309
		83.609	4.140	34.602	1.309			64.800	1.851

8	<i>Diffenbachia picta</i> Schott.	17.992 28.919 20.676	1.796 1.380 1.436	21.927 76.978 72.646	2.182 3.491 2.760	N.S.O.F.	N.S.O.F.	73.637 59.193 84.629	1.573 1.573 1.851
9	<i>Lasia heterophylla</i> (Roxb.) Schott	150.393 154.332 132.369	4.499 3.995 3.111	269.909 284.975 250.957	8.727 6.293 4.329	89.654 89.959 83.919	2.654 2.670 3.464	56.460 26.650 16.350	4.493 3.001 3.007
10	<i>Dracontium nivosum</i> (Lem.) GHZhu.	21.422 17.068 13.799	0.617 1.234 0.617	35.492 42.954 28.299	1.056 1.987 0.981	53.260 41.397 42.549	2.618 2.350 1.380	21.059 29.551 21.659	2.225 1.745 1.989
11	<i>Colocasia esculenta</i> (black spotted) (Linn.) Schott	208.191 212.311 146.811	0.436 0.401 2.621	170.812 156.941 173.811	0.309 0.399 1.451	173.733 183.331 135.855	0.436 0.556 0.656	209.677 147.081 94.277	0.309 0.331 0.301
12	<i>Amorphophallus sylvaticus</i> (Roxb.) Kunth	216.466 215.654 213.877	5.433 5.443 5.212	326.764 322.111 326.909	6.502 7.702 8.596	91.415 87.012 91.977	0.617 0.412 0.654	41.717 43.252 41.601	1.245 0.481 0.412
13	<i>Amorphophallus campanulatus</i> Blume.ex.Decne	226.171 216.466 250.957	5.656 5.343 6.116	327.892 327.515 326.223	8.391 7.108 7.002	208.621 210.554 197.616	5.533 5.022 4.652	51.671 57.301 54.112	1.231 1.212 1.454
14	<i>Colocasia esculenta</i> (black) (Linn.) Schott	112.811 109.292 135.294	4.171 3.833 4.022	141.599 142.499 139.884	1.201 0.651 1.841	136.599 148.701 153.632	4.982 4.696 4.401	43.584 44.277 39.011	0.921 1.051 0.720

[L= Length; B=Breath]; [N.S.O.F.- No Storage Organ Found]

REFERENCES

- [1] Bradbury, J. H. and Nixon, R. W. (1998). The acidity of raphides from the edible aroids. *Journal of the Science of Food and Agriculture* 76:608–616.
- [2] Johns, T. and Kubo, I. (1988). A survey of traditional methods employed for the detoxification of plant foods. *Journal of Ethnobiology* 8(1):81–129.
- [3] Paull, R. E., Tang, C.-S., Gross, K. and Uru, G. (1999). The nature of the taro acidity factor. *Postharvest Biology and Technology* 16:71–78.
- [4] Sakai, W. S. (1979). Aroid root crops, acidity and raphides. *Tropical Foods* 1:265–278.
- [5] Bouropoulos, N., Weiner, S. and Addadi, L. (2001). Calcium oxalate crystals in tomato and tobacco plants: Morphology and in vitro interactions of crystal-associated macromolecules. *Chemistry* 7:1881–1888.
- [6] Horner, H. T., Jr. and Wagner, B. L. (1995). Calcium oxalate formation in higher plants. In S. R. Khan (ed.), *Calcium Oxalate in Biological Systems*, pp 53–72. Boca Raton: CRC Press.
- [7] Franceschi, V. R. and Horner, H. T., Jr. (1980). Calcium oxalate crystals in plants. *The Botanical Review* 46:361–427.
- [8] Nakata, P. A. (2003). Advances in our understanding of calcium oxalate crystal formation and function in plants. *Plant Science* 164:901–909.
- [9] Prychid, C. J. and Rudall, P. A. (1999). Calcium oxalate crystals in monocotyledons: A review of their structure and systematics. *Annals of Botany* 84:725–739.
- [10] Cody, A. M. and Horner, H. T., Jr. (1983). Twin raphides in the Vitaceae and Araceae and a model for their growth. *Botanical Gazette* 144:318–330.
- [11] Webb, M. A. (1999). Cell-mediated crystallization of calcium oxalate in plants. *Plant Cell* 11:751–761.
- [12] Arnott, H. J. and Webb, M. A. (2000). Twinned raphides of calcium oxalate in grape (vitis): Implications for crystal stability and function. *International Journal of Plant Science* 161:133–142.
- [13] Wattendorff, J. (1976). A third type of raphide crystal in the plant kingdom: Six-sided raphides with laminated sheaths in *Agave americana* L. *Planta* 130:303–311.
- [14] Horner, H. T., Jr, Kausch, A. P. and Wagner, B. L. (1981). Growth and change in shape of raphide and druse calcium oxalate crystals as a function of intracellular development in *Typha angustifolia* L. (Typhaceae) and *Capsicum annuum* L. (Solanaceae). *Scanning Electron Microscopy* 1981:251–62.
- [15] Kostman, T. A. and Franceschi, V. R. (2000). Cell and calcium oxalate crystal growth is coordinated to achieve high-capacity calcium regulation in plants. *Protoplasma* 214:166–179.