

ORIGINAL ARTICLE

Evaluation of Anti-Oxidant Property of *Acoras calamus* l. and *Commiphora Mukul* (Hook. Ex Stocks) Engl. for Medicinal Use

^{1,2*}Subodh Kumar, ^{1#}Nilanjana Prasad, ³Sweta Thakur, ⁴Abhilekha Sharma

¹Department of Biotechnology & Microbiology, School of Sciences, Noida International University, Sector-17 A, GautamBudh Nagar-201308 (UP) India

²Multi-Disciplinary Research Unit, Department of Health Research, Dr. S. N. Medical College, Jodhpur-342001 (Raj.) India

³Department of Agriculture & Environmental Science, School of Sciences, Noida International University, Sector-17 A, GautamBudh Nagar-201308 (UP) India

⁴Department of Chemistry, School of Sciences, Noida International University, Sector-17 A, GautamBudh Nagar-201308 (UP) India

*#Corresponding Authors [Equal Authorship]

ABSTRACT

Increasing burden of allopathic medicine on the society leads to anti-microbial resistance and cytotoxicity due to side effects. Therefore, in the present study we have evaluated the phytochemical screening and presence of phenolic and flavonoid compounds in methanolic extracts of rhizome of *Acora scalamus* and gum of *Commiphora mukul* for medicinal use. Rhizome of *A. calamus* and gum of *C.mukul* were extracted in methanol by maceration followed by their phytochemical analysis. Determination of total phenolic and flavonoid content was performed. ABTS and H₂O₂ antioxidant assays were deployed to evaluate total antioxidant capacity using ascorbic acid as reference standards. Analytical thin layer chromatography was performed to detect the presence of phenols and flavonoids. Total phenolic and flavonoid content of methanolic extract of *A. calamus* and *C.mukul* were 20.901±0.0693 and 5.692±0.0595, mg/g of gallic acid equivalent per gram of extract, and 13.37043±0.1072 and 24.55±0.263, mg/g of quercetin equivalent/gram of extract, respectively. ABTS and H₂O₂ % scavenging activity of *A.calamus* and *C.mukul* were found to be 99.52±0.225, 56.50±0.243, 99.83±0.066 and 99.23±0.0884 respectively. *A.calamus* yielded three phenol bands (R_f = 0.890, 0.727 and 0.581) and two flavonoid bands (R_f = 0.951, 0.548) whereas *C.mukul* yielded three phenol bands, (R_f = 0.726, 0.579, 0.290) and three flavonoid bands, (R_f = 0.967, 0.822 and 0.5). The study shows that rhizome of *A.calamus* and gum of *C.mukul* possessed considerable amounts of phenol and flavonoid content and have high antioxidant activity in methanol.

Keywords: *Acoras calamus*, *Commiphora mukul*, Phytochemical, Antioxidant, Thin layer Chromatography

Received 25.01.2022

Revised 09.02.2022

Accepted 18.03.2022

How to cite this article:

S Kumar, N Prasad, S Thakur, A Sharma. Evaluation of Anti-Oxidant Property of *Acoras calamus* l. and *Commiphora Mukul* (Hook. Ex Stocks) Engl. for Medicinal Use. Adv. Biores. Vol 13 [2] March 2022: 200-207.

INTRODUCTION

India is the largest producer of phytotherapeutics and recognized as the botanical garden of the world[1]. An antioxidant can be broadly defined as any substance that defers or inhibits free radical induced oxidative damage to a target molecule[2]. Phenolic compounds are stable radical intermediates, an important source of natural antioxidants which are known to be produced in large amounts by different plant parts such as seeds, bark of stems and roots and leaves in harsh environments with strong light intensities which mainly protects the plants against pathogens[3]. Molecular structures of flavonoids have been known to be responsible for its strong antioxidant activity at very low concentration[4]. *A. calamus* commonly known as "sweet flag" belongs to the family acoraceae and is a perennial and semiaquatic traditional indigenous medicinal plant distributed in areas of Jammu and Kashmir, Himachal Pradesh, Manipur, Tripura states, etc. in India[5]. The roots and rhizome of *A.calamus* have been reported to exhibit various pharmacological activities including cardiovascular, anti-inflammatory, CNS etc.[6]. *C.mukul*[syn.

Commiphora wightii] (Family- Burseraceae) commonly known in India as “guggul” and in English as “Indian bdellium” is a therapeutically important ethnomedicinal plant since the ancient India Ayurveda system⁸. In India, it is abundantly found at arid regions of Gujarat (Kutch region), Rajasthan, Assam, Madhya Pradesh, and Karnataka[7, 8]. Oleo -gum resin (pale yellow or brown coloured mass) secreted by *C. mukul* is a complex mixture of flavonoids, terpenoids, steroids, minerals and oils[9]. Therefore, the present objectives of the study are to evaluate the phytochemical, total phenol content, total flavonoid content, in- vitro antioxidant activity and detect the presence phenolic and flavonoid compounds in methanolic extract of rhizomes of *A. calamus* and gum of *C. mukul* through thin layer chromatography.

MATERIALS AND METHODS

Collection and Extraction of Plant materials

A. calamus rhizome was collected from forests of Himachal Pradesh and *C. mukul* gum (Neeraj Traders, India) was purchased. Then, they were washed thoroughly with distilled water to remove the dust particles present in it, shade dried and grinded into a coarse powder. Maceration was carried out in methanol for 7 days with constant shaking in every 24 hours. After maceration, the mixture was filtered using Whatmann filter paper and the filtrate was evaporated using rotavapor at low temperature and pressure by a rotary evaporator to obtain the crude extract. All the extracts were preserved at 4°C for further analysis[10].

Preliminary phytochemical screening Of Crude extracts

Methanolic extracts of *A. calamus* rhizome and *C. mukul* gum were subjected to different phytochemical tests for the detection of bioactive chemical constituents such as alkaloids, phenolics, tannins, flavonoid, carbohydrates, saponins and proteins using standard procedures[11, 12].

Determination of total phenols by Folin-Ciocalteu reagent method

Total phenolic content was determined by using Folin-Ciocalteu (FC) method through some modifications[13, 14]. FC reagent was freshly prepared (diluted 1:1 with de-ionized water) before use. Plant extracts (0.025 mL) were mixed with FC reagent (2.5 mL) and then allowed to stand for 5 min. at room temperature in order to allow complete reaction with the reagent. The reaction mixture was further neutralized with sodium carbonate solution (2 mL, 7.5%, w/v). It was kept in dark at room temperature for 30 min. with intermittent shaking for blue color development. After 30 min, the absorbance was read at 765 nm using UV-visible spectrophotometer (Thermo Spectronic) against a blank solution containing all the reagents except FC reagent which is replaced by an equal volume of distilled water. All the measurements were carried in triplicates and averaged. Methanolic solution of gallic acid served as standard (1 mg/mL) phenolic compound (0.2, 1, 2, 3, 4, and 5 µg/mL) for plotting calibration curve. Total phenol content in the extracts were expressed as Gallic acid equivalent (GAE) (mg of Gallic acid equivalent/g of dry weight sample) and was calculated by the formula[15]:

$$T = \frac{C \times V}{M} \dots \dots \dots [\text{Eq. 1}]$$

Where, T=Total content of phenolic compound/g of plant extract, in GAE;

C=Concentration of Gallic acid established from the Calibration curve, microgram/ml;

V=Volume of methanolic extract; M=Weight of methanolic plant extract, g.

Determination of total flavonoid content by aluminium chloride colorimetric method

Total flavonoid content was determined by method described by Madaan et al. through some modifications[16]. Plants extracts (0.5 mL) were separately mixed with methanol (1.5 mL), aluminium chloride (0.1 mL, 10% w/v), potassium acetate (0.1 mL, 1M) and distilled water (2.8 mL). The reaction mixture was then incubated for 30 min at room temperature. After 30 min., using UV-visible spectrophotometer (Thermo Spectronic) the absorbance was measured at 415 nm against a blank solution containing all the reagents except aluminium chloride which is replaced by an equal volume of distilled water. All the measurements were carried in triplicates and averaged. Methanolic solution of quercetin from 20 to 100 µg/mL served as standard (1 mg/mL) flavonoid compound to obtain the calibration curve. The total flavonoid content was expressed as quercetin equivalent (QCE) (mg of quercetin equivalent/g of dry weight sample) and was calculated by the formula mentioned earlier for TPC[Eq.1]

Where, T=Total content of flavonoid compound/g of plant extract, in QE;

C=Concentration of quercetin established from the Calibration curve, microgram/ml;

V=Volume of methanolic extract; M=Weight of methanolic plant extract, g.

ABTS (2,2-azino-bis 3 ethyl benzothiazoline-6- sulfonic acid) radical scavenging Assay

The radical scavenging activity of rhizome extract of *A. calamus* and gum extract of *C. mukul* were assessed by ABTS radical scavenging assay described by Re et al.[17] through some modifications. ABTS aqueous stock solution (7 mM) and potassium persulfate stock solution (2.45 mM) were freshly prepared. Equal

volumes of the stock solutions were mixed and allowed to react in the dark at room temperature for 12-16 h to obtain ABTS^{•+} free radical. After 12-16h, ABTS^{•+} working solution was diluted with methanol to obtain an absorbance of 0.70±0.02 at 734nm. Briefly, each methanolic extracts (10µl) were mixed with working solution of ABTS^{•+}(2ml) and absorbance was taken 1 min. after initial mixing using UV-visible spectrophotometer (Thermo Spectronic). All the measurements were carried in triplicates and averaged. Ascorbic acid was used as standard. 1mg/ml stock solution of ascorbic acid was prepared and the calibration line between % inhibition and ascorbic acid was established using the following concentrations of ascorbic acid: 1,1.5,2,2.5,3,3.5,4,4.5, and 5 µg/ml. The total antioxidant capacity was calculated as percent inhibition of ABTS radical using the following equation,

$$\text{Percentage inhibition of ABTS(\%)} = \left(\frac{\text{Cabs} - \text{Sabs}}{\text{Cabs}} \right) * 100 \dots \dots \dots [\text{Eq.2}]$$

where Cabs is the control absorbance (ABTS radical + methanol); Sabs is the sample absorbance (ABTS radical + sample extract).

Hydrogen peroxide (H₂O₂) scavenging assay

H₂O₂ scavenging capacity of the extracts was evaluated using method described by Owaisi et al[18] through some modifications. Ascorbic acid was used as a positive control. A solution of H₂O₂ (40 mM) was prepared in phosphate buffer (0.05 mM, pH 7.4). Methanolic extracts (20 µl) and ascorbic acid (200 µl) in PBS was mixed with H₂O₂ solution (0.6mL, 40 mM) to make a final volume of 2ml. The absorbance was read at 230nm UV-visible spectrophotometer (Thermo Spectronic) after 10 min. against subsequent blank solutions containing extracts (20µl) in PBS without H₂O₂. The amount of H₂O₂ radical scavenged by each extract was calculated as percent inhibition of H₂O₂ radical using the following equation,

$$\text{Percentage inhibition of H}_2\text{O}_2(\%) = \left(\frac{\text{Cabs} - \text{Sabs}}{\text{Cabs}} \right) * 100 \dots \dots \dots [\text{Eq.3}]$$

where Cabs is the control absorbance (Ascorbic acid + H₂O₂ radical + PBS);

Sabs is the sample absorbance (H₂O₂ radical + sample extract).

Thin Layer Chromatography for detection of phenols and flavonoids

Phenolic and flavonoid compounds in the methanolic crude extracts were separated by thin layer chromatography (TLC) using TLC plates coated with silica [19 – 21]. The plates were air dried and activated at 110° for 1 hr in *heat incubator* (Bioline Technologies, India). Samples (2mg/ml) were dissolved in methanol and concentrated in heat incubator for 10-15 min. Gallic acid and quercetin served as standard phenolic and flavonoid compounds (1mg/1ml). Ethyl acetate: formic acid: acetic acid: water (10:1.1:1.1:2.6, v/v/v/v) solvent system was used for separation of different phenolic compounds whereas for flavonoid separation methanol: chloroform: hexane: water (3:1:3:3) was used. Glass capillaries were used to spot samples and standard separately on the activated TLC plates up to 1.5cm from one edge of the TLC plate marked by pencil immediately after cooling at room temperature. The plates, with dried spotted samples, were gradually placed into the presaturated development tank, closed and left to develop. The mobile phase was allowed to travel until about 1 cm from the top end. After development, TLC plates were removed gently and solvent front was marked using a soft pencil line. For detection of phenols and flavonoids, TLC plates were air dried and then sprayed with a fine spray of 2% FeCl₃ solution and 1% ethanolic aluminium chloride solution, left to dry and then visualized under UV light at 254/365nm. The chromatograms were marked and retention factors were calculated using the formula:

$$\text{Rf value} = \frac{\text{distance travelled by solute}}{\text{distance travelled by solvent}} \dots \dots \dots [\text{Eq.4}]$$

RESULTS

Percentage yield of crude extracts

Approximately, 6-7g (70%) viscous mass was obtained from 10 g powdered rhizome and gum parts of *A.calamus* and *C.mukul* after maceration (Table 1A).

Preliminary phytochemical screening

Preliminary phytochemical studies revealed the presence of major bio-actives in rhizomes of *A.calamus* and gum of *C.mukul* (Table 1B).

Total phenolic content

Total phenolic content was calculated using the following linear regression equation obtained from the standard plot $y = 0.4152x + 0.446$, $r^2 = 0.9906$, where y is absorbance and x is the amount of gallic acid in µg (Fig 1A). The total phenolic content of the methanolic extract of *A.calamus* and *C.mukul* were 20.901±0.0693 and 5.692±0.0595, mg/g of gallic acid equivalent per gram of extract respectively (Fig 1B).

Total Flavonoid Content

Total flavonoid content was calculated using the following linear regression equation obtained from the standard plot $y=0.0184x+0.118$, $r^2=0.9944$, where y is absorbance and x is the amount of quercetin in μg (Fig 2A). The total flavonoid content of *A.calamus* and *C.mukul* were 13.37043 ± 0.1072 and 24.55 ± 0.263 , mg/g of quercetin equivalent/gram of extract (Fig 2B).

ABTS radical scavenging Assay

Radical scavenging activity of rhizome extracts of *A.calamus* and gum extract of *C.mukul* was also determined using ABTS method. ABTS scavenging activity (%) of standard ascorbic acid was calculated using the following linear regression equation obtained from the standard plot, $y=16.077x+24.413$, $r^2=0.981$, where y is absorbance and x is the amount of quercetin in μg (Fig 3A). The ABTS % scavenging activity of *A.calamus* and *C.mukul* were found to be 99.52 ± 0.225 and 56.50 ± 0.243 , respectively (Fig 3B).

Hydrogen Peroxide Radical Scavenging activity

Antioxidant activity in the two methanolic extracts were studied using the well-known H_2O_2 method and expressed in terms of %scavenging activity. The % scavenging activity of *A.calamus* and *C.mukul* were found to be 99.83 ± 0.066 and 99.23 ± 0.0884 , respectively (Fig 3C). Total phenolics, flavonoid and antioxidant content of crude extracts of *A.calamus* and *C.mukul* are summarised in Table 2.

Thin layer chromatography for detection of phenolic and flavonoid compounds

Thin layer chromatography results showed the presence of different phenolic and flavonoid compounds in *A.calamus* and *C.mukul* (Fig 4) and (Table 3).

DISCUSSION

Preliminary phytochemical studies revealed the presence of bioactive constituents of our interest like phenol and flavonoids in methanolic rhizome extract of *A.calamus* and gum extract of *C.mukul*. In our study, the total phenolic content in *A.calamus* and *C.mukul* were found to be 20.901 ± 0.0693 and 5.692 ± 0.0595 , mg/g of gallic acid equivalent per gram of extract, respectively. Previous studies reported that total phenolic content in methanolic rhizome extract of *A.calamus* ranged from 1.67 ± 0.288 to 10.42 ± 0.180 mg/g gallic acid, whereas 12.45 mg GAE/g dw to 23.4 mg GAE/ 100 g dw phenolic content have also been reported [22, 23] which indicated that our result is quite consistent with results reported by other studies. The differences in the total phenolic content in same species of methanolic extract indicated that the total phenol content differed depending on the plant species, environmental factors, developmental stage and genetic make-up [24]. In our study, the total flavonoid content of *A.calamus* and *C.mukul* were found to be 13.37043 ± 0.1072 and 24.55 ± 0.263 , mg/g of quercetin equivalent/gram of extract, respectively. Total flavonoid content in methanolic extract of *A.calamus* rhizome reported by a previous study ranged from 0.50 ± 0.053 to 24.50 ± 0.322 mg/g catechin²³ whereas another study reported 54.2 ± 0.15 mg quercetin /grams in ethyl acetate extract⁵. Our results hypothesized that extraction solvents might be a major factor influencing total flavonoid content amongst same species extracted by different solvents.

Our study showed that rhizomes of *A.calamus* and gum of *C.mukul* possessed high antioxidant activity. Previous studies reported guggulsterones as the chief bioactive compound present in *C.mukul* gum resin responsible for its *in vitro* antioxidant activity [25]. Thin layer chromatography results revealed the presence of different phenolic and flavonoid compounds in *A.calamus* and *C.mukul*. The color and R_f value of *C.mukul* (CM3') matched with that of standard (quercetin) which indicated the presence of quercetin as the major flavonoid compound in *C.mukul* gum extract. This data might be supported by an earlier study which reported based on our quantitative flavonoid analysis and thin layer chromatography results; we can conclude that *C.mukul* gum resin has a high flavonoid content, "quercetin" being the chief flavonoid compound. Our findings are supported by a study which reported quercetin as the major flavonoid compound found in the flowers of *C.mukul* [26].

The present work revealed presence of bioactive compounds and the strong antioxidant activity of methanolic rhizome extract of *A.calamus* and gum extract of *C.mukul*. Both the plants showed the presence of phenols and flavonoids but *A.calamus* had a higher phenolic content whereas *C.mukul* showed a higher flavonoid content in methanol. Furthermore, on the basis of our results, it can be concluded that a pronounced free radical scavenging potential of *A.calamus* rhizome and *C.mukul* gum methanolic extract was observed which suggested a possible therapeutic application of *A.calamus* rhizome and *C.mukul* gum as potential sources of natural antioxidants for development of bioactive compounds, which might help in prevention of oxidative stress. It has been reported that high phenolic content were associated with strong antioxidant activity [27]. Thus, we can conclude from our study that antioxidant potential of *A.calamus* and *C.mukul* might be possibly influenced by high phenolic and flavonoid compounds present in them. Since, *C.mukul* is IUCN listed, so proper usage of this phytotherapeutic drug should be done to

prevent depletion of this wonder drug. More research on *A. calamus* rhizome and *C. mukul gum* resin needs to be done to explore its immense phytotherapeutic potential.

Table 1A: % yield of Methanol extracts from *A. calamus* and *C. mukkul*

Scientific Name	Family	Common names	Parts analysed	Medicinal uses	Yield % (w/w)
<i>Acorus calamus</i>	Acoraceae	Sweet flag, vaccha	Rhizome	Detoxifying agent, treatment of nervous disorders, reducing obesity, aromatic stimulant, febrifuge vermifuge and hypotensive	60%
<i>Commiphora mukul</i>	Bursuraceae	Guggul, Indian myrrh	Gum resin	Cardiac tonic, expectorant, skin complexion enhancer, carminative, hypercholesteremic and antibacterial	70%

Table 1B: Results of preliminary phytochemical screening of methanolic extracts of *A. calamus* and *C. mukkul* (+: present in low concentration, ++: present in moderate concentration, +++: present in higher concentration, -: absent)

Phytochemical	Name of Test	<i>A. calamus</i>	<i>C. mukul</i>
Alkaloid	Dragendorff's test	+++	+++
	Wagner's Test	++	++
Phenol	Ferric chloride test	++	+
Tannin	Ferric chloride test	+	-
Flavonoid	Alkaline reagent test	+	+++
Carbohydrate	Fehling's test	+++	+
	Benedict's test	-	-
	Barfoed's test	-	-
Saponin	Foam test	+	-
Protein	Xanthoproteic test	+++	+
	Ninhydrin test	-	-
	Biuret test	+	-

Table 2: Total phenolics, flavonoid and antioxidant content of crude extracts of *A. calamus* and *C. mukul*

Crude Extracts	<i>A. calamus</i>	<i>C. mukul</i>	P
TPC ^a	20.901±0.0693	5.692±0.0595	0.00001*
TFC ^b	13.37043±0.1072	24.55±0.263	0.00001*
ABTS ^c	99.52±0.225	56.50±0.243	0.00001*
H ₂ O ₂ ^d	99.83±0.066	99.23±0.0884	0.000361*

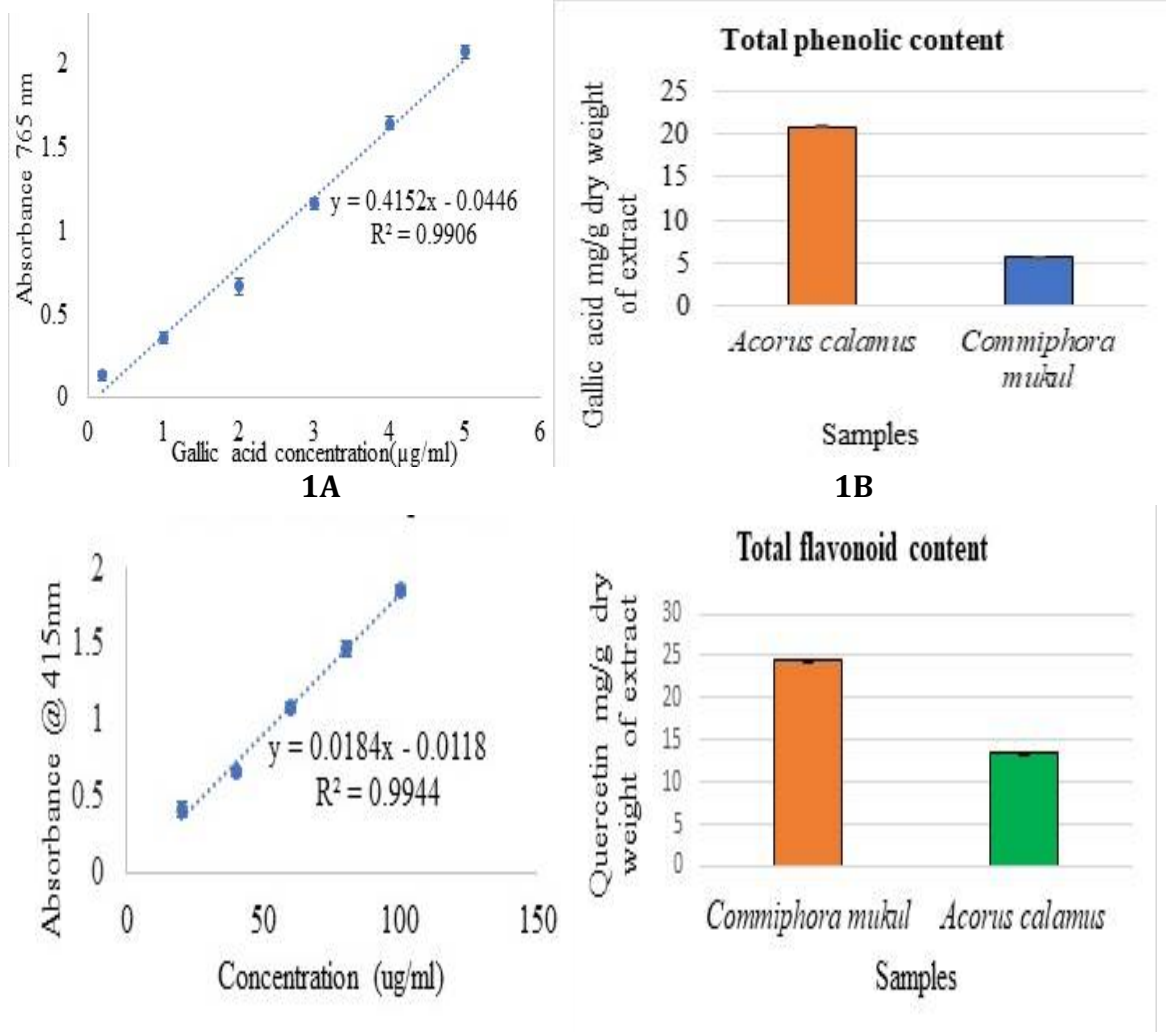
^aTotal phenolic content expressed as mg of gallic acid equivalent (GAE)/g of dry weight (dw).^bTotal flavonoid content expressed as mg of quercetin equivalent (QE)/g of dry weight (dw).^cABTS radical scavenging activity expressed as % inhibition.^dH₂O₂ radical scavenging activity expressed as %inhibition. Values are mean±SD; *:P<0.05

Table 3A: Phenol analysis using 2% FeCl₃ solution

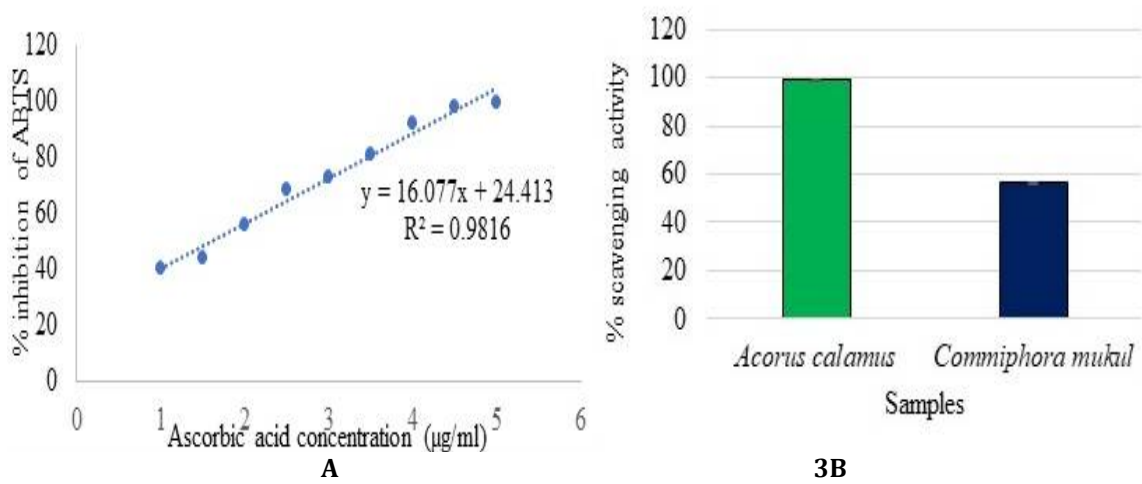
Crude extracts	No of spots	Rf value
Gallic Acid (Std)	1	0.909
<i>A. calamus</i>	3	0.890,0.727,0.581
<i>C. mukul</i>	3	0.726,0.579,0.290

Table 3B:Flavonoid analysis using 1% ethanolic aluminium chloride solution

Crude Extracts	No of spots	Rf value
Quercetin (Std)	1	0.967
<i>A. calamus</i>	2	0.951,0.548
<i>C. mukul</i>	3	0.967,0.822,0.5



1(A):Standard curve of Gallic acid; (B) Total phenolic content in crude extracts
Figure 2(A): Standard curve of Quercetin; (B) Total flavonoid content in crude extracts



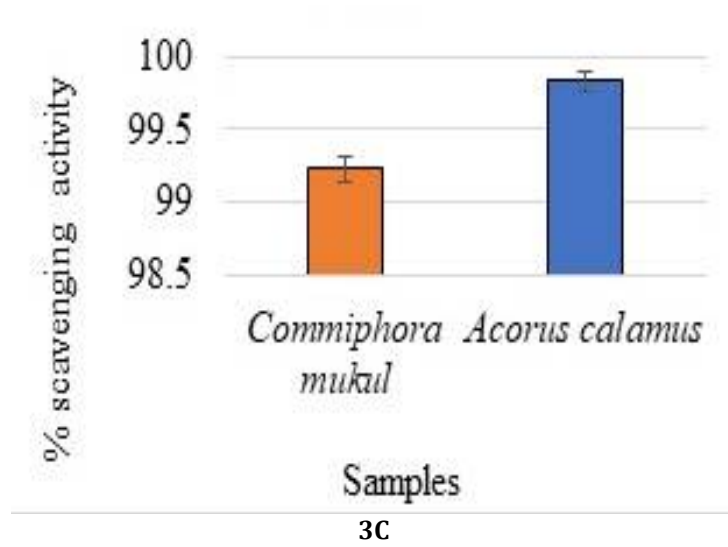


Figure 3: (A) Standard curve of Ascorbic acid; (B) ABTS scavenging activity (%) of crude extracts; (C) Hydrogen peroxide scavenging activity (%) of crude extracts

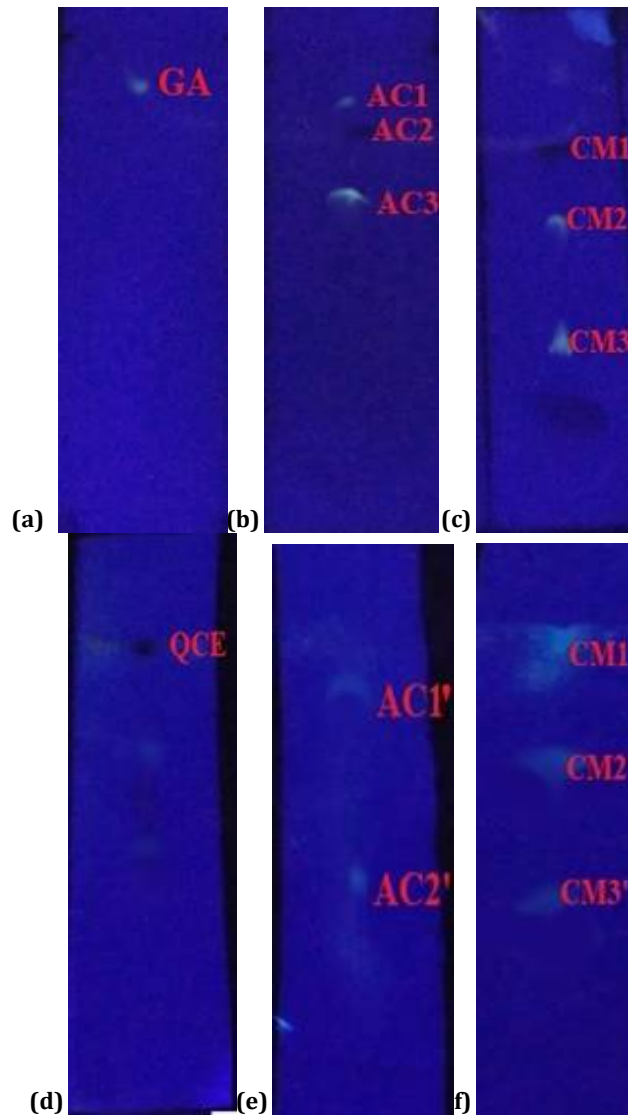


Figure 4. Chromatograms for phenols (a-c) and flavonoids (d-f) under UV: (a) Gallic acid standard (b) *A. calamus* (c) *C. mukul* (d) quercetin standard (e) *A. calamus* (f) *C. mukul*

REFERENCES

- Muthukrishnan, S., Sivakkumar, T. (2018). Physicochemical Evaluation, Preliminary Phytochemical Investigation, Fluorescence and TLC Analysis of Leaves of *Schleicheraoleosa* (Lour.) Oken. Indian J Pharm Sci., 80(30):525-532.
- Mahdi-Pour, B., Jothy, S.L., Latha, L.Y., Chen, Y., Sasidharan, S. (2012). Antioxidant activity of methanol extracts of different parts of *Lantana camara*. Asian Pac J Trop Biomed., 2(12):960-965.
- Nićiforović, N., Mihailović, V., Masković, P., Solujić, S., Stojković, A., Pavlović, Muratspahić, D. (2010). Antioxidant activity of selected plant species; potential new sources of natural antioxidants. Food Chem Toxicol., 48(11):3125-3130.
- Mishra, S.L., Sinhamahapatra, P.K., Nayak, A., Das, R., Sannigrahi, S. (2010). In vitro Antioxidant Potential of Different Parts of *Oroxylum indicum*: A Comparative Study. Indian J Pharm Sci., 72(2):267-269.
- Nath, P., Yadav, A.K. (2016). Anti-helminthic activity of a standardized extract from the rhizomes of *A. calamus* Linn. (Acoraceae) against experimentally induced cestodiasis in rats. J Intercult Ethnopharmacol., 5(4):390-395.
- Pulok, K.M., Venkatesan, K., Mainak, M., Houghton, P.J. (2007). *A. calamus*: Scientific validation of ayurvedic tradition from natural resources. Pharm Biol., 45:651-666.
- Soni, V. (2010). Efficacy of in vitro tissue culture versus stem cutting for propagation of *Commiphora wightii* in Rajasthan, India. Conservation Evidence., 7:91-93.
- Sarup, P., Bala, S., Kamboj, S. (2015). Pharmacology and Phytochemistry of Oleo-Gum Resin of *Commiphora wightii* (Guggulu). Scientifica (Cairo), 2015:138039.
- Dave, P.N., Chopda, L.V. (2013). Review on biological activity and determination of E&Z-Guggulsterones concentration by HPLC & HPTLC Methods. Int J Chem Stud., 1(3):166-175.
- Hussain, A., Murtaza, G., Mehmood, A., Qureshi, R.A. (2017). Conservation of indigenous knowledge of medicinal plants of Western Himalayan region Rawalakot, Azad Kashmir, Pakistan. Pak J Pharm Sci., 30:773-782.
- Sahoo, S., Ghosh, G., Das, D., Nayak, S. (2013). Phytochemical investigation and in vitro antioxidant activity of an indigenous medicinal plant *Alpinianigra* B.L. Burt. Asian Pac J Trop Biomed., 3(11):871-876.
- Dixit, A., Rohilla, A., Dixit, J., Singh, V. (2014). Qualitative analysis of various plant extracts of *Alpinia officinarum*. Int. J. Pharm. Chem. Biol. Sci., 4(3):505-508.
- Alhakmani, F., Kumar, S., Khan, S.A. (2013). Estimation of total phenolic content, in-vitro antioxidant and anti-inflammatory activity of flowers of *Moringaoleifera*. Asian Pac J Trop Biomed., 3(8):623-627.
- Stanly, C., Bhatt, A., Ali, H.M.D., Keng, C.L., Lim, B.P. (2011). Evaluation of free radical scavenging activity and total phenolic content in the petiole-derived callus cultures of *Zingiber zerumbet* Smith. J Med Plant Res., 5(11): 2210-2217.
- Barku, V.Y.A., Opoku, Y., Owusu, A.E, Mensh, E.F. (2013). Antioxidant activity and the estimation of total phenolic and flavonoid contents of the root extract *Amaranthesspinosus*. Asian J Plant Sci Res., 3:69-74.
- Madaan, R., Bansal, G., Kumar, S., Sharma, A. (2011). Estimation of total phenols and flavonoids in extracts of *Actaeaspicata* roots and antioxidant activity studies. Indian J Pharm Sci., 73(6): 666-669.
- Re, R., Pellegrini, N., Proteggente, A., Pannala, A., Yang, M., Rice-Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. Free Radic Biol Med., 26(9-10):1231-1237.
- Owaisi, M.A., Hadiwi, N.A., Khan, S.A. (2014). GC-MS analysis, determination of total phenolics, flavonoid content and free radical scavenging activities of various crude extracts of *Moringa peregrina* (Forssk.) Fiori leaves. Asia Pacific J Tropical Biomedicine., 4:964-970.
- Burman, V., Kanaujia, H., Lehari, K., Aastha, Singh, N.P., Vaishali. (2019). Characterization of phenolic compounds of turmeric using TLC. J Pharmacogn Phytochem., 8(2S):994-998.
- Harborne, B.J. Phytochemical Methods, A Guide to Modern Techniques of Plant Analysis. 3rd ed. New Delhi: Springer Pvt Ltd; 1998.
- Gwatidzo, L., Dzomba, P., Mangena, M. (2018). TLC separation and antioxidant activity of flavonoids from *Carissa bispinosa*, *Ficus sycomorus*, and *Grewia bicolor* fruits. Nutrire., 43(1):1-7.
- Ahmed, F., Chandra, N.S.J.N., Urooja, A., Rangappa, K.S. (2009). In vitro antioxidant and anticholinesterase activity of *A. calamus* and *Nardostachys jatamansi* rhizomes. J Pharm Res., 2:830-33.
- Wojdylo, A., Oszmianski, J., Czemerys, R. (2007). Antioxidant activity of phenolic compounds in 32 selected herbs. Food Chem., 105(3):940-949.
- Kondakova, V., Tsvetkov, I., Batchvarova, R., Badjakov, I., Dzhabazova, T., Slavov, S. (2009). Phenol compounds-qualitative index in small fruits. Biotechnol Biotechnol Equip., 23(4):1444-1448.
- Chander, R., Khanna, A.K., Pratap, R. (2003). Anti hyperlipidemic activity of guggulsterone, the active principle of guggulipid from *C. mukul*. J Med Arom Plant Sci., 24:370-374.
- Baldwa, V.S., Bhasin, V., Ranka, P.C., Mathur, K.M. (1981). Effects of *C. mukul* (Guggul) in experimentally induced hyperlipemia and atherosclerosis. J Assoc Physicians India., 29:13-17.
- Cao, G., Sofic, E., Prior, R. L. (1997). Antioxidant and prooxidant behaviour of flavonoids: structure activity relationships. Free Radic Biol Med., 22(5):749-760.

Copyright: © 2022 Society of Education. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.