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ORIGINAL ARTICLE

Analysis of Molecular Diversity in *Gerbera* Genotypes Revealed By RAPD Markers

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ABSTRACT

Random amplified polymorphic DNA (RAPD) markers were used to study the genetic diversity among seven genotypes of the genus Gerbera an important ornamental plant. Out of 20 RAPD decamers tested, only 13 amplified genomic DNA across all the 7 genotyes of Gerbera. A total of 52 bands were scored in all the genotypes. Our results suggested that genetic diversity in Gerbera genotypes using RAPD banding data may be useful for plant improvement and an efficient way to conserve genetic resources of Gerbera in addition to their ornamental and medicinal uses. Key words: Gerbera, Molecular diversity, RAPD, PCR based DNA markers.

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INTRODUCTION

The Barberton/Transvaal daisy, or Gerbera is a flower with increasing commercial significance. Gerbera is one of the leading cut flowers and ranks among the top ten cut flowers of the world [1] [3]. It has wide applicability in the floral industry as cut flower and potted plant. The species, a perennial herb, is native to South Africa and Asia. It is an important commercial flower grown through the world in a wide range of climatic conditions. The genus consists of about forty species [2]. Out of the recorded species, only one species Gerbera jamesonii is under cultivation. Few important cultivators of Gerbera are Cream Clementine, Maron Clementine, Delphi, Vesta, Uranus, Terra Queen, Dusty, Valantine, Diablo, mariso and Pascal. Based on the flower heads, they may be grouped into single semi- double and double cultivars [4]. Like any ornamental plant Gerberas are produced exclusively for their aesthetic values. Gerbera is a member of the large sunflower family Asteraceae, as such, Gerbera offers great potential for comparative developmental research within a single genotype. Moreover, different Gerbera varieties show an impressive spectrum of colour patterns, directly displaying responses to developmental stages at all important morphological levels (flower type, flower organ and within organs). Further, Gerbera harbors an arsenal of Asteraceae type secondary metabolites, not present in other model plants. With powerful reverse genetics methods, a large collection of sequences and a new cDNA microarray, Gerbera has become a model plant of the sunflower family. Gerbera is one of the top selling cut flowers and pot plants in the world. Gerbera viridifolia is one of the species is referred to in the eastern cape as 'Iyeza lamazi' or the herb of milk and is probably used to stimulate milk production in cattle or as an ingredient of maas (the locally produced soured milk) [5],

[6]. The smoke of *G.viridifolia* is inhaled by the southern poeple to treat colds. Pounded leaf infusions of *G. ambigua* are used in zulu traditional medicine for tapeworm and stomach ache, whereas root infusions are taken orally for coughs. In Zimbabwe, root infusions are used for heart pain and abdominal pain in babies.

Present research work on analysis of genetic diversity of *Gerbera* genotypes using PCR based DNA markers will pave the way to understand economically important genus *Gerbera* and to elucidate the genetic variation in *Gerbera* genotypes.

MATERIAL AND METHODS

Plant material

The seedlings of 7 Gerbera genotypes i,e Rosalin, Savannah, Winter queen, Goliath, Dana ellen, Dalma, and Sunway were purchased from germlasm collections centre Pune. The collected fresh leaves were powdered in liquid nitrogen and stored in deep freezer

(-70°) till further processing.

DNA isolation

Genomic DNA was extracted from young leaves following the modified CTAB method [7]. Quantification and purity of isolated DNA was checked through uv-spectrophotometer and agarose gel electrophoresis. PCR amplification

A set of 40 random decamer primers (Operon Technologies, Almeda, California, USA) were used to amplify the genomic DNA [8]. PCR reactions were conducted in a 20µl reaction mixture consisting of 100ng. geonomic DNA, 5pM / µl RAPD Primer, 10µM dNTPs, 0.5µl Taq. DNA polymerase (3U/ µl) (Bangalore Genei, Bangalore, India), 10 X PCR buffer and 1.5mM MgCl₂. DNA amplification was carried out using a Gradient palm Cycler (Corbett Research, Australia). Amplifications were carried out in a Gradient Palm Cycler with initial denaturation for 4 minutes at 94°C and each cycle with 15 seconds at 4°C, 15 seconds primer annealing at 35°C, 1.15 minutes for extension at 72°C. The reaction was continued for 40 cycles followed by 7 minutes at 72°C to ensure the completeness of the primer extension. The PCR products were analyzed on 1.5 % (w/v) agarose gels in 1X TBE buffer under constant voltage of 100 v for 3 to 4 hours. Experiments were repeated several times to ensure the reproducibility of results and the best gels of the replicates were used for band scoring. The gels were stained with ethidium bromide solution and documented in a gel documentation system (Fig.1). A λ DNA, 500bp ladder (Banglore Genei, India) was used as molecular size marker.

Data Analysis

Polymorphic RAPD amplified product was considered to be a unit character and the populations were manually scored as binary data with presence of a band as "1" and absence as "0". Only clearly distinguishable DNA bands were used in the genetic analysis. The molecular size of the amplification products were calculated from a standard curve based on the known size of the DNA fragments of the ladder. Estimates of genetic similarity were calculated between all pairs of the species according to [accard's Similarity Index (1908) [9]. The matrix obtained was used to evaluate the genetic diversity among the genotypes of *Gerbera* with cluster analysis using an Unweighed Pair Group Method with Arithmetic Averages (UPGMA) [10]. All the statistical analysis were performed with an aid of NTSYS-PC computer program version 2.0 [11]. The similarity matrix was calculated based on Jaccard's similarity coefficient.

In the present investigation, 7 genotypes of the genus *Gerbera* were used to assess interspecific genetic polymorphism. Out of 20 random decamer primers (operon) initially tested, only 13 primers were finally selected on the basis of robustness of the amplification, clarity and scorability of banding patterns, for genetic diversity analysis. These primers differed greatly in their efficiency for revealing polymorphism (Table.2). The estimates of pairwise genetic distance between the eight species of *Gerbera* is given in Table.3.

In all, 52 reproducible and scorable DNA fragments were amplified, 72 % of which were polymorphic and consistently generated from 13 primers across Gerbera genotypes. Primer OPB-8 yielded highest number of bands (15) while OPB-7 and OPB-13 amplified the lowest number of bands (2) with an average of 3.8 bands per primer (Table 2). The representative RAPD profiles of 7 genotypes of Gerbera are shown in (Fig 1). The approximate size of the largest fragment was 2.5kb, where as the smallest recognizable fragment was 0.5kb in size.

Seven accessions of Gerbera (Table 1) revealed 72% of polymorphism with 13 RAPD primers. A total of 52 bands were scored of which 37 bands are polymorphic. The number of bands ranging from 2 to 15 corresponding to an average of 3.8 bands per primer (Table 2). Jaccard's similarity matrix reveals that, the levels of genetic similarity between accessions ranged from 0.1 to 0.75 (Table 3). The maximum number of polymorphic bands (15) were yielded with primer OPB-8 and minimum (2 bands) from OPB-13 (Table 2). Dendrogram (Figure 2) reveals the formation of 2 major genetic groups among 7 accessions studied based on the genetic similarity index (Table 3), even though morphologically they were similar and inseparable. The first group is again classified into two sub groups, where the first sub group includes one accession *Sunway* while the second subgroup includes two subclusters, first subcluster includes three accessions Savannah, Dana ellen and Rosalin. The second subcluster includes two accessions Winter queen and Goliath. Maximum genetic similarity is between accessions Winter queen and Goliath (0.75). The second major group includes single accession Dalma. The denrogram (Fig 2) based on similarity

index (SI) (Table 2) showed distinct separation of the collected accessions though morphologically they were similar and inseparable. The dendrogram separated the accessions into two major groups, having 72% similarity.

RESULTS AND DISCUSSION

RAPD markers represent an efficient and inexpensive tool to generate molecular data and thus, have been used successfully in various taxonomic and phylogenetic studies, [15], [12], High level of genetic similarity is expected among *Gerbera* genotypes due to similar geographical conditions. But, in contrast, they showed broad genetic base indicating earlier introduction of the species, and subsequently leading to accumulation of variation. The genetic differentiation of accessions of *Gerbera* genotypes could broadly be explained as a result of abiotic (geographical for example, hydrographic connections or climatic differentiation) and biotic (pollination between populations and seed dispersal) factors. The percentage polymorphism (72%) among *Gerbera genotypes* is comparatively much higher than other plants, for example, Cathaya argyrophylla (32%) [16], Paeonia rockii (33.3%) [17], a vulnerable medicinal plant Oroxylum indicum (49.61%)[15], Costus speciosus (35%) [18], this clearly indicates high level of genetic diversity within Gerbera genotypes. High level of genetic diversity in populations of Gerbera genotypes may be due to cross pollination by insects, dispersal of seeds, habitat changes. There is a close genetic similarity of 75% observed between accessions Winter queen and Goliath which clearly depict that, genetically the similarity may be because of the similar environmental conditions. In essence, the RAPD method used in this study displayed appreciable intra-population variation or molecular polymorphism, which is pre-existed in different collections. In spite of their morphological similarity, substantial polymorphism was observed among the accessions under study. The study revealed that, though the decamer primers are small in comparison to the large genome of *Gerbera jamsonii*, they produced appreciable amplicons sufficient to demarcate all accessions. The dendrogram also established genetic relatedness among different accessions and quantum of changes that occurred in the genome in the course of evolution. This study confirms the suitability of RAPD as a reliable, simple, easy to handle and elegant tool in molecular diagnosis of different accessions of Gerbera jamsonii. Currently, it is also proved that, the entries that were found to be similar in taxonomical classification based on morphological characteristics do have divergence at DNA level. Vast genetic variation is an indicative of the evolving nature of the taxa.

Sl.No.	Acc. No.	Genotypes of <i>Gerbera</i>
01	G.1	Rosalin
02	G.2	Savannah,
03	G.3	Winter queen
04	G.4	Goliath,
05	G.5	Dana ellen
06	G.6	Dalma,
07	G.7	Sunway

 Table 1: Accession number and different genotypes of used for RAPD analysis
 Gerbera jamsoni

Primers	Nucleotide Sequence 5' → 3'	No. of amplified bands	No. of polymorphic bands
OPA-02	TGCCGAGCTG	5	3
OPC-06	GAACGGACTC	6	3
OPD-02	GGACCCAACC	6	4
OPA-12	TCGGCGATAG	-	-
OPB-07	GGTGACGCAG	5	2
OPB-08	GTCCACACGC	15	12
OPB-10	CTGCTGGGAC	-	-
OPB-11	GTAGACCCGT	7	6
OPB-13	TTCCCCCGCT	3	2
OPB-16	TTTGCCCGGA	-	-
OPB-17	AGGGAACGAG	-	-
OPB-19	ACCCCCGAAG	-	-
OPB-20	GGACCCTTAC	5	5
	Total:	52	37

	1	2	3	4	5	6	7
1	1.000						
2	0.663	1.000					
3	0.538	0.357	1.000				
4	0.703	0.554	0.752	1.000			
5	0.578	0.653	0.271	0.470	1.000		
6	0.423	0.140	0.333	0.201	0.100	1.000	
7	0.535	0.491	0.423	0.594	0.491	0.167	1.000

Table 3: Similarity matrix based on the Jaccard's Similarity Index



Fig.1. Gerbera genotypes, A. Dana ellen and B. Dalma

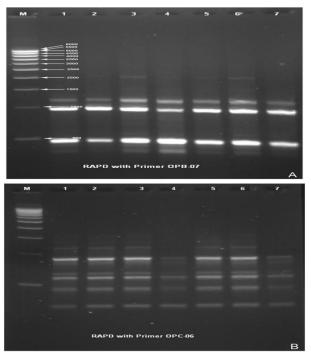


Fig. 2. RAPD profiles of 7 *Gerbera* genotypes generated by primer (A) OPC-6 and (B) OPB-7. M- indicates the DNA molecular size marker (500 bp ladder). Lane numbers corresponds to the species serial number given in Table 1.

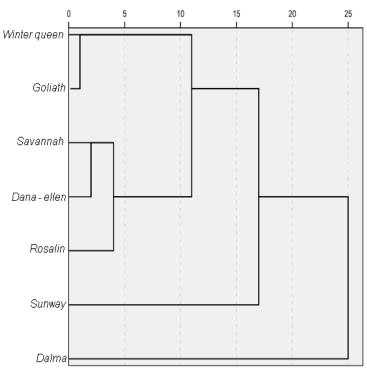


Fig.3. Dendrogram showing the genetic relationship among 7 *Gerbera* genotypes generated by RAPD data using the UPGMA method.

CONCLUSION

The study was undertaken to evaluate the extent and range of genetic diversity available in *Gerbera* genotypes. Accurate estimates of diversity are a prerequisite for optimizing sampling strategies and for conserving the genetic resources. The high diversity revealed by RAPD is in agreement with the conclusion that, out breeding plants retains considerable variability [19],[13],[14].Variation within the species suggests that, this species has large effective population size or large mutation rate due to longer generation.

As most of the *Gerbera* genotypes are used as an ornamental plant, the presence of a unique RAPD marker among the various*Gerbera* genotypes indicate the utility of the approach for DNA fingerprinting purposes. RAPD fingerprinting has a number of potential applications and include the determination of cultivar purity, efficient use and management of genetic resources particularly the establishment of property rights (plant variety protection and patenting). This study represents only the first step in using RAPD markers as a tool to develop species specific SCAR DNA markers for molecular analysis of individual *Gerbera* genotypes.

REFERENCES

- 1. Parthsarathy V.A. and Nagaraju V. [1999]. *In vitro* propagation in *Gerbera jamesonii* Bolus. Indian journal of Horticulture, **56**:82-85.
- 2. Kanwar J.K. and Kumar S. [2008]. In vitro propagation of Gerbera-A Review. Hort. Sci. (Prague), 35(1) 35-44.
- 3. Das P. and Singh P.K.S. [1989]. *Gerbera*. In: Bose T.K. and Yadav L.P. (eds), Commercial Flowers. Calcutta, Naya Prokash: 601-622.
- 4. Loeser H. [1986]. New Gerbera cultivars at Heidelberg. Deutscher Gartenbau, 40: 1461-1464.
- 5. Dold A.P. and Cocks M.L. [2002]. The trade in medicinal plants in the eastern Cape Province, South Africa. South African Journal of Science. 98:589-597.
- 6. JainS.M., Vitti D., TucciM., Grassotti A., Rugini E.and SaccardoF. [1998]. Biotechnology and mutagenesis in *Gerbera* improvement . Advances in Hort. Sci. **12**:1.47-53.
- 7. DoyleJ.J.and Doyle J.L.[1990].Isolation of Plant DNAfrom fresh tissue.Focus, **12**:13-15.
- 8. Williams G.K., Kubelik, A.R., Livak K.J., Rafalsky J.A. and Tingey S.V.[1990]. DNA polymorphisms amplified by arbitary primers are useful as genetic markers. Nucl. Acids. Res. **18**: 6531–6535.
- 9. Jaccard P. [1908]. Nouvelles researches sur la distribution florale.Bull. Vaudoise Sci. Nat. 44 :223-270.
- 10. Sneath P.H.A. and Sokal R.R.1973. Numerical Taxonomy. WH Freeman, San Francisco.
- 11. Rohlf F.J. [1998]. NTSYS-PC, Numerical Taxonomy and Multivariate Analysis System, version 2.0, New York: Applied Biostatistics Inc.

- 12. Iruela M., Rubio J., Cubero J.I., Gil J. and Milan T. [2002]. Phylogenetic analysis in the genus *Cicer* and cultivated chickpea using RAPD and ISSR markers. Theor. Appl. Genet. **104** : 643–651.
- 13. Sharma T.R. and Jana S. [2002]. Species relationships in *Fagopyrum* revealed by PCR-based DNA fingerprinting. Theor. Appl. Genet. **105**: 306–312.
- 14. Nebauer S.G., Del C.A. and Segura J.[2000]. An assessment of genetic relationships within the genus *Digitalis* based on PCR generated RAPD markers. Theor.Appl.Genet. **100**:1209–1216.
- 15. Jayaram K. and Prasad M.N.V. [2008].Genetic diversity in *Oroxylum indicum* (L.) Vent. (Bignoniaceae), a vulnerable medicinal plant by random amplified polymorphic DNA marker. Afr. J. Biotechnol. vol. **7(3)** : 254-262.
- 16. Wang X.Q., Zou Y.P., Zhang D.M. and Hong D.Y. [1996]. RAPD analysis for genetic polymorphism in *Cathaya argyrophylla*. Sci. China ©. **26**:437-441.
- 17. SuY.J.,Wang T.and Huang C.[1999].RAPD analysis of different population of *Dacydium pierrei*. Acta Bot. Sin. 40 : 169-175.
- 18. Asit B.M., Vincy A.T. and Elanchezhian R.[2007]. RAPD pattern of *Costus speciosus*. Koen ex. Retz., an important medicinal plant from the Andaman and Nicobar Islands. Curr. Sci.Vol. **93**: No.3.
- 19. Hamrick J.L.[1990].Isozymes and the analysis of genetic structure in plant populations. In: Soltis DE and Soltis PS (eds).Isozyme in Plant Biol. pp. 87-105, Chapman and Hall Landon.