

## ORIGINAL ARTICLE

# Superoxide Dismutase and Peroxidase activities under Lead and Cadmium Stresses in *Berberis integerrima* and *Cercis siliquasrum*

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### ABSTRACT

Plants have various biochemical mechanisms to tolerate and adapt to environmental stresses. The study was conducted to evaluate superoxide dismutase (SOD) and peroxidase (POD) activities under Pb and Cd stresses in *Berberis integerrima* and *Cercis siliquasrum*. For this purpose, three-year-old seedlings with various concentrations (1000, 2000, 4000 and 6000 ppm) of Cd and Pb for 45 days in 15-days intervals were selected, and superoxide dismutase (SOD) and peroxidase (POD) were investigated as markers of oxidative stress. Results showed the response of antioxidant enzymes to Cd and Pb treatments was different in both species. *Berberis integerrima* and *Cercis siliquasrum* have different response to Cd and Pb treatments that the reason due to different biochemical responses of plants to environmental stresses. The results of this study can be used to choose the most appropriate plant according to the related stress especially under pollution stress.

**Keywords:** *Berberis integerrima*, *Cercis siliquasrum*, SOD, POD.

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## INTRODUCTION

Soil contamination with heavy metals is one of the most eminent environmental issues around the world [4], [13]. Heavy metals can be remained in the environment with no major changes and bioavailable of these elements are different from other soil components [11]. All heavy metals are considered as pollutant in high concentration [7]. Although heavy metals have adverse impacts on plants, they can live in soils contaminated by high concentration of heavy metals [17]. Plants can remove pollutants in various ways. For instance they can mitigate pollution by absorption, stability and transmission methods [14]. Cadmium (Cd) and lead (Pb) are the most poisonous elements [27] which the first one leads to contaminate soil and prevent the root and stem growth as well as affects plant homeostasis. Antioxidant enzymes can't have their normal activities in high concentration of Cd [18] and also Cd changes cell members by lipid peroxidation and chloroplast metabolism by prevention of chlorophyll biosynthesis [21], Second one (Pb) due to its wide distribution and danger is extremely considered for environment. Hence, Soil contamination with Pb can affect both microorganism activities and physiological parameters of plants [18] Heavy metals have impacts on physiological processes that one of them is the production of reactive oxygen species (ROS) which damages proteins, lipids and DNA [24]. The activities of antioxidant enzymes, such as SOD, CAT, APX and GR, will be increased in response to increasing the ROS. In addition, in stress conditions, ROS causes damaging to cell membranes which results in lipid peroxidation and production of malondialdehyde (MDA) [10]. Urban green space can alleviate the pollution. Therefore, using the trees and shrubs tolerating to pollution is one of the most important principles in creating the efficient green space. Hence, the more information about biochemical responses of plants to pollution stress is essential to choose the most appropriate species in urban areas. For this purpose, the present

study aimed to evaluate the Superoxide dismutase and Peroxidase responses of *Berberis integerrima* and *Cercis siliquasrum*, two main ornamental shrubs planting in urban areas of Iran, to Pb and Cd stress.

## MATERIAL AND METHODS

### Experimental treatments

Three-year-old seedlings of *Berberis integerrima* and *Cercis siliquasrum* in Alborz nursery belonging to the Research Institute of Forests and Rangelands, Karaj, were cultivated in plastic pots containing approximately 8 kg of soil. After that, the plants were treated by cadmium chloride and lead nitrate separately in concentrations of 0, 1000, 2000, 4000, 6000 ppm, 100 cc per plant in three times at 15-days intervals. Subsequently, the leaves of both species were sampled in four directions of crown one month after the last treatment.

### Superoxide dismutase (SOD)

The method of Giannopolitis and Ries [15] was used to measure SOD activity. The mixture consisted of phosphate buffer 50 mM (pH = 7.8), methionine 13 mM, Nitro blue tetrazolium (NBT) 75  $\mu$ M, EDTA 1/0 mM, riboflavin 13 mM and 50 ml of enzyme extract. Then the mixture was under fluorescent light for 15 min and in the dark for 15 min and its absorption was read by spectrophotometer at the wavelength of 560 nm according to Unit / mg protein FW.

### Peroxidase (POD)

POD activity was recorded according to the method of [8]. The mixture was contained phosphate buffer 0.1 mM (pH = 7.5), Pyrogallol solution 4 mM, hydrogen peroxide 3 mM and 100 ml of enzyme extract. After that, the samples were read by spectrophotometer at wavelength of 240 nm for 20 min based on Unit / mg protein FW.

### Statistical analyses

Data were submitted to CRD and Duncan multiple range test using SAS 9.1. Differences were considered significant at  $p < 0.05$ .

## RESULTS AND DISCUSSION

### SOD activity

As is shown by figure 1, SOD activity was lower than that in control at all treatments of Cd stress for *Cercis siliquasrum*. There was a significant reduction at low levels of stress (1000 ppm and 2000 ppm) by approximately 42%-48%. In addition, the enzyme activity was increased in concentration of 4000 ppm and dramatically decreased in 6000 ppm (77%) under Cd stress. However, the reduction in SOD activity in 4000 ppm of Pb was not statistically significant and the SOD activity was increased in 6000 ppm. Generally, enzyme activity in *Berberis integerrima* was less than *Cercis siliquasrum*, but plant response to stress levels was different. SOD was significantly increased by increasing Cd stress (64-75%) and there was found a remarkable reduction of SOD at 1000 ppm (47%). SOD exists in all oxygen metabolism cells, and SOD in the cytoplasm, mitochondria and chloroplasts are found to superoxide radicals, hydrogen peroxide converts [2]. Similar results in different plant species about alleviating oxidative stress by increasing antioxidant enzymes have been reported [5], [28]. In some cases increasing stress have resulted in reduction of enzyme activity that this issue can be due to inactivation of the enzyme by enzyme catalysis or non-specific binding of heavy metals to be related enzyme activity center [12].

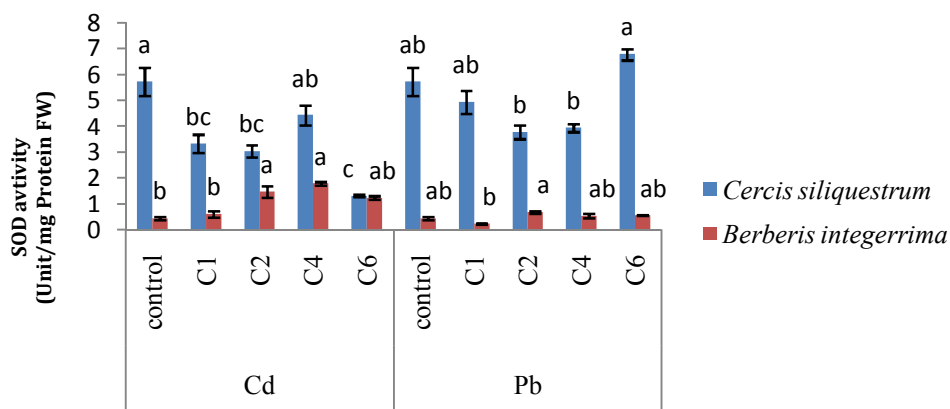


Figure 1: The effects of Cd and Pb on SOD activity of *Berberis integerrima* and *Cercis siliquasrum* leaves. C1, C2, C4 and C6 are 1000, 2000, 4000 and 6000 ppm, respectively. Letters indicate significant differences at  $p < 0.05$ .

## POD activity

Results of Cd and Pb effects on POD activity are presented in figure 2. There was no significant difference in both *Berberis integerrima* and *Cercis siliquasrum* for various levels of Cd. POD activity in *Berberis integerrima* was more than *Cercis siliquasrum*. However, plant responses to Pb stress were different at different concentrations. There was observed a 42% increase of POD activity at 1000 ppm in *Cercis siliquasrum* in comparison with control, while POD activity in *Berberis integerrima* was less than control and the lowest activity was observed in 1000 and 6000 ppm of Pb. POD has an important role in plant response to stress and it will be active under stress condition [25]. Antioxidant enzyme response to heavy metals varies among different species and even among different organs [20]. This issue can be supported by different POD activity between *Berberis integerrima* and *Cercis siliquasrum* even in same treatments in the present study. Many studies have revealed that increasing POD is in association with heavy metals (Shu et al, 2011; [16], while decreasing POD activity by heavy metals have been reported by some authors (Shu et al, 2011; [19], [22]. In the present study, decreasing the enzyme activity was observed in the high concentrations. The ability of plants appears to be limited to increase the activity of antioxidant enzymes to counteract the effects of stress. Studies have shown increasing the concentrations of heavy metals eventually lead to the loss of all antioxidant enzymes. Enzyme activity at low concentrations of heavy metals increases and it will decrease by increasing heavy metal concentration and finally will decrease after passing threshold, depending on plant [6]. It also induces long-term effects of heavy metals increase the enzyme activity, especially the activity of POD [23].

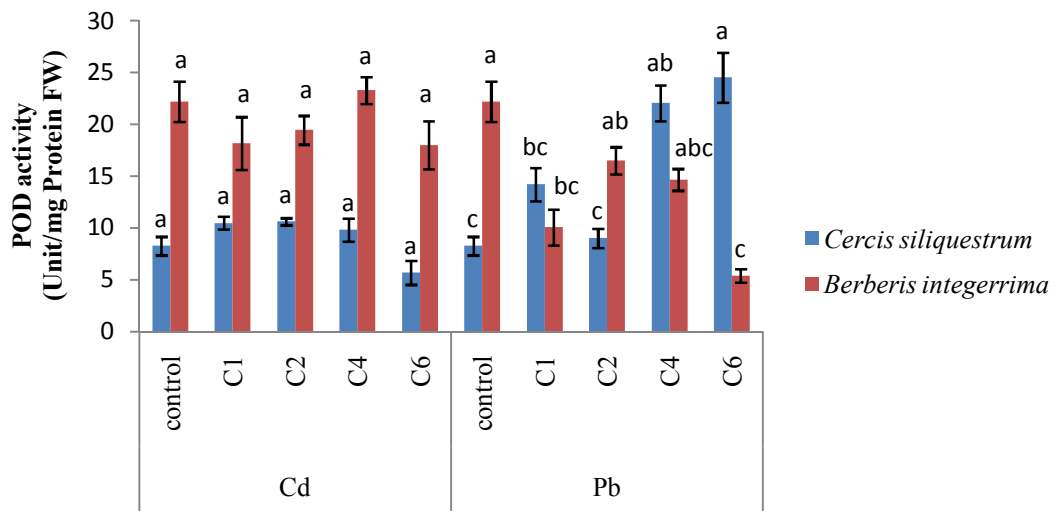


Figure2: The effects of Cd and Pb on POD activity of *Berberis integerrima* and *Cercis siliquasrum* leaves. C1, C2, C4 and C6 are 1000, 2000, 4000 and 6000 ppm, respectively. Letters indicate significant differences at  $p < 0.05$ .

## CONCLUSION

The present study has shown the effects of Cd and Pb concentrations on biochemical properties of *Berberis integerrima* and *Cercis siliquasrum* and their response to these metals. Both species have different biochemical responses to heavy metal stress except to Cd stress, and results showed both species have the same potential for tolerating various concentrations of Cd. In regarding to Pb stress, *Cercis siliquasrum* is more tolerant than *Berberis integerrima*. Therefore, in soils contaminated by Pb, *Cercis siliquasrum* will be recommended.

## REFERENCES

- Ahmad, P., Sharma, S., Srivastava, P.S., (2006). Differential physio-biochemical responses of high yielding varieties of Mulberry (*Morus alba*) under alkalinity ( $\text{Na}_2\text{CO}_3$ ) stress in vitro. *Physiol. Mol. Biol. Plants* 12, 59-66.
- Alscher, R.G., Erturk, N., and Heath, L. S. (2002). Role of superoxide dismutases (SODs) in controlling oxidative stress in plants. *J. Exp. Bot.* 53:1331-1341.
- Arnon, D.J. (1949). Copper enzymes in isolated chloroplasts. *Plant Physiol.* 24: 1-15.
- Bodar, C.W.M., Pronk, M.E.J., and Sijm, D.T.H.M. (2005). The European Union risk assessment on zinc and zinc compounds: The process and the facts. *Integr Environ Assess Manag* 1:301-319.

5. Candan, N., and Tarhan, L. (2003), The correlation between antioxidant enzyme activities and lipid peroxidation levels in *Mentha pulegium* organs grown in Ca<sup>2+</sup>, Mg<sup>2+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup> and Mn<sup>2+</sup> stress conditions, *Plant Science*, 163, 769-779.
6. Cao, X., Ma, L.Q. and Tu, C., (2004). Antioxidative responses to arsenic in the arsenic- hyperaccumulator Chinese brake fern (*Pteris vittata* L.). *Environmental Pollution*, 128: 317-325.
7. Chehregani, A. and Hajisadeghian, S. (2009). New chromosome counts in some species of Asteraceae from Iran. *Nordic J. Bot.* 27: 247-250
8. Chance, B. and Maehly, A. C. (1955). Assay of catalase and peroxidases. *Methods Enzymol.* 2: 764-775.
9. Chaoui, A., Mazhoudi, S., Ghorbal, M.H., and Ferjani, E. (1997). Cadmium and zinc induction of lipid peroxidation and effects on antioxidant enzyme activities in bean (*Phaseolus vulgaris* L.). *Plant Sci* 127:139-147.
10. De Britto, J. A., Roshan Sebastian, S., Sheeba Gracelin, D. H. (2013). Effect of lead on malondialdehyde, superoxide dismutase, proline activity and chlorophyll content in *Capsicum annum*. *Bioresearch Bulletin*. Oct 17 [last modified: 2013 Oct 17]. Edition 1.
11. Doumett, S., Lamperi, L., Checchini, L., Azzarello, E., Mugnai, S., Mancuso, S., Petruzzelli, G., and Del Bubba, M. (2008). Heavy metal distribution between contaminated soil and *Paulownia tomentosa*, in a pilot-scale assisted phytoremediation study: Influence of different complexing agents. *Chemosphere*, 72 (10), 1481-1490 (10 pages).
12. Filek, M., Keskinen, R., Hartikainen, H., Szarejko, I., Janiak, A., Miszalski, Z., and Golda, A. (2008). The protective role of selenium in rape seedlings subjected to cadmium stress. *Journal of Plant Physiology* 165 (8), 833-844.
13. Fotakis, G., and Timbrel J. A. (2006). Role of trace elements in cadmium chloride uptake in hepatoma cell lines. *Toxicol Lett.* 164; 79-103.
14. Ghosh, M. and Singh, S.P. 2005. A review on phytoremediation of heavy metals and utilization of its by products. *Applied Ecology and Environmental Research* 3(1), pp 1-18.
15. Giannopolitis, C. N and Ries, S. K. (1977). Superoxide Dismutases. II. Purification and quantitative relationship with water-soluble protein in seedlings. *Plant Physiol.* 59, 315-318.
16. Hayat, S., Hayat, Q., Alyemeni, M. N., and Ahmad, A. (2013). Proline enhances antioxidative enzyme activity, photosynthesis and yield of *Cicer arietinum* L. exposed to cadmium stress. *Acta Bot. Croat.* 72 (2), 323-335.
17. Ismail, S., Khan, F., and Zafir Iqbal, M. (2013). Phytoremediation: assessing tolerance of tree species against heavy metal (pb and cd) toxicity. *Pak. J. Bot.*, 45(6): 2181-2186.
18. John, R., Ahmad, p., Gadgil, K., and Sharma, S. (2009). Heavy metal toxicity: Effect on plant growth, biochemical parameters and metal accumulation by *Brassica juncea* L. *International Journal of Plant Production* 3 (3), ISSN: 1735-6814 (Print), 1735-8043 (Online).
19. Luna, C.M., Gonzalez, C.A. and Trippi, V.S. (1994). Oxidative damage caused by an excess of copper in oat leaves. *Plant and Cell Physiology*, 35(1): 11-15.
20. Mazhoudi, S., Chaoui, A., Ghorbal, M.H. and El Ferjani, E. (1997). Response of antioxidant enzymes to excess copper in tomato (*Lycopersicon esculentum* Mill.). *Plant Science*, 127(2): 129-137.
21. Nagajyoti, P. C., Lee, K. D., and Srekanth, T. V. M. (2010). Heavy metals, occurrence and toxicity for plants: a review. *Environ Chem Lett* . 8:199-216. DOI 10.1007/s10311-010-0297-8.
22. Palma, J.M., Gómez, M., Yáñez, J. and del Río, L.A. (1987). Increased levels of peroxisomal active oxygen related enzymes in copper tolerant pea plants. *Plant Physiology*, 85(2): 570-574.
23. Qadir, S., Qureshi, M.I., Javed, S. and Abdin, M.Z. (2004). Genotypic variation in phytoremediation potential of *Brassica juncea* cultivars exposed to Cd stress. *Plant Science*, 167(5): 1171-1181.
24. Schützendübel, A. and Polle, A. (2002). Plant responses to abiotic stresses: Heavy metal-induced oxidative stress and protection by mycorrhization. *Journal of experimental botany*, Vol. 52, No.372, Antioxidants and Reactive Oxygen Species In Plants Special Issue, pp. 1351-1365.
25. Shalini, V. and Duey, R. S. (2003). Lead toxicity induced lipid peroxidation and alters the activities of antioxidant enzymes in growing rice plant. *Plant Science* 164: 1645-1655.
26. Sharma, P., and Dubey, R.S.H. (2005). Lead toxicity in Plants. *Plant Physiol.* 17: 35-52.
27. Sharma, R. K., Agrawal, M., and Marshall F. M. (2008). Heavy metal (Cu, Zn, Cd and Pb) contamination of vegetables in urban India: A case study in Varanasi. *Environmental Pollution* 154. 254-263
28. Zembala, M., Filek, M., Walas, S., Mrowiec, H., Korná's, A., Miszalski, Z., and Hartikainen, H. (2010). Effect of selenium on macro- and microelement distribution and physiological parameters of rape and wheat seedlings exposed to cadmium stress. *Plant and Soil* 329 (1-2), 457-468