## **ORIGINAL ARTICLE**

# Does drought stress induce physiological mechanisms in *Celtis* caucasica L. Seedlings?

Fereshteh Kordrostami<sup>1</sup>, Anoushirvan Shirvany<sup>2</sup>, Pedram Attarod<sup>3\*</sup>, and Mostafa Khoshnevis<sup>4</sup>

<sup>1</sup>Department of Forestry and Forest Economics, Faculty of Natural Resources, University of Tehran, Karaj, Iran

<sup>2</sup>Department of Forestry and Forest Economics, Faculty of Natural Resources, University of Tehran, Karaj, Iran

<sup>3</sup>Department of Forestry and Forest Economics, Faculty of Natural Resources, University of Tehran, Karaj, Iran

> <sup>4</sup>Research Institute of Forests and Rangelands, Tehran, Iran <sup>\*</sup> Corresponding E-mail address: attarod@ut.ac.ir

## ABSTRACT

Drought stress is an important environmental factor inhibiting plant growth. The examination of physiological reaction of different species to the drought stress is useful to recognize effective mechanisms in drought tolerance and choosing the suitable species for plantation in dry lands. In order to determine the resistance of Celtis caucasica and understand the physiological mechanisms in response to the drought condition, an experiment was designed in a completely randomized block with one factor (irrigation), at five levels (1, 3, 5, 7, and 9 days) with five replications chlorophyll fluorescence parameters (Fv/Fm, Fm and F0) and pigments content (total chlorophyll, chlorophyll a, and chlorophyll b) of leaves. The results indicated that drought stress induced a significant reduction at five percent level in maximum photochemical efficiency of photosystem II (Fv/Fm) and maximum fluorescence (Fm) while the minimum fluorescence (Fo) was not significantly affected by drought. With increasing the intensity of drought, chlorophyll content in leaves was increased. Maintaining the leaf chlorophyll content under stress conditions, is one of the physiological indicators of stress resistance concluding that C. caucasica would be one of the most suitable species for afforestation plans in arid and semiarid regions.

Key words: drought stress, Celtis caucasica, chlorophyll fluorescence, proline

Received 02/09/2014 Accepted 24/11/2014

©2014 Society of Education, India

How to cite this article:

Fereshteh K, Anoushirvan S, Pedram A, Mostafa K. Does drought stress induce physiological mechanisms in *Celtis caucasica* L. Seedlings?. Adv. Biores., Vol 5 [4] December 2014: 30-34. DOI: 10.15515/abr.0976-4585.5.4.3034

## INTRODUCTION

Drought stress is an important environmental factor that affects plants growth, productivity, photosynthesis and changes plants metabolism [23], [24]. Drought stress is one of the most important environmental inhibitor for photosynthesis via stomatal closure [13].In this condition, decreasing photosynthesis is related to disturbing the biochemical processes of the photosynthetic apparatus. Photosystem II is the most sensitive part of the photosynthetic systems and plays a critical role in plants reaction to environmental stresses [13]. Damage to photosystem II is frequently the first symptoms of stress in a leaf [23].

Plant physiologists have suggested chlorophyll fluorescence as a sensitive indicator of stress condition in plants. The measurement of chlorophyll fluorescence in natural environment is a useful technique to understand the tolerance of photosynthetic apparatus of the environmental stress [23]. This technique is nondestructive and can be just as operative as the gas exchange techniques to disclose differences between drought tolerant and susceptible species [25]. There have been many fluorescence parameters so that it is not possible to give an extensive review of all of these here.  $F_0$  (minimum fluorescence from dark adapted leaf is the level of fluorescence when primary quinine electron acceptors of PS II (Q<sub>A</sub>) are maximally oxidized (PS II centers are open),  $F_m$  (Maximum fluorescence from dark adapted leaf is the

level of fluorescence when  $Q_A$  is maximally reduced (PSII centers are closed),  $F_v$ (variable fluorescence from dark adapted leaf shows the ability of PSII to perform primary photochemistry (photoreduction of  $Q_A$ ) and  $F_v/F_m$ . The values of  $F_v/F_m$  in dark adapted leaves indicates the potential yield of the photochemical reacting in PSII used as a practical alternative to monitor the photosynthetic performance [15]. The optimal values of  $F_v/F_m$  in healthy leaves of most plant species are about 0.832 [6] and the lower values show the exposure of plants to stress. This is the consequence of photo damage to PSII reaction centers, and the progression of quenching process [23], [9]. Literature reviews show that there are differences between physiological responses of plants to drought stress. These differences are related to species type and to some extent, experimental conditions. In the present research, *C.caucasica* was employed to study the photosynthetic responses to the drought stress. It is commonly used as an urban tree because of its ability to withstand drought and tolerate in urban environments. The aims of this study were: (i) to evaluate the effect of drought stress on the chlorophyll fluorescence parameters (ii) to find the relationship between chlorophyll contents and drought tolerance in *C. caucasica* and, (iii) assessing the vitality of *C.caucasica* in different irrigation regimes for planting in semi-arid areas.

### **MATERIAL AND METHODS**

#### Materials and Experimental design

This study was conducted in the Alborz Research Station in southwest slope of Alborz mountain in Iran (latitude 35° 48′N, 50° 54′E and 1300 m a.s.l) with a semi-arid climate where means annual temperature and rainfall are 13.7 ° C and 230 mm, respectively.

The seeds were collected from 7km in the road of Chaloos-Nojan with elevation of 1450m and in April, 2011, they were sawn in plastic pots (15×40 cm) containing an equal mixture (2:1:1) of clay, sand and organic fertilizer. Pots were irrigated every day for about one year. No artificial lighting was used in this study excluding natural light from the sun. Regular management was conducted until the seedlings were about 50 cm height. Twenty five seedlings were selected randomly and classified to five groups of treatments. Drought experiment started in August 2012. Measurements were performed on 1, 3, 5, 7 and 9 days after drought. Soil was fully drenched before the experiment.

#### Chlorophyll fluorescence

Chlorophyll fluorescence was measured using a portable chlorophyll fluorometer (PAM 2500 WalzGermany) on three fully expanded leaves of five pots per treatment. Leaves were adapted to darkness for 30 minutes by attaching light clips to the leaf surface. All measurements carried out every day, from 10 AM to 14 PM.

## Chlorophyll content

To assess chlorophyll content, fresh and mature leaves (0.1g) were extracted with 10 ml of 80% acetone and centrifuged at 4000 rpm for 15 min. The absorbance was read spectrophotometric ally at 654 and 663 nm and calculated for chlorophyll a, chlorophyll b, and total chlorophyll according to [3]. *Statistical Analysis* 

The normality of data was assessed using the Kolmogorov-smirnov test and the homogeneity of variances was determined using the Leven's test. The average of parameters was compared using Duncan or Games Howel test.Data analysis was performed using SPSS 17.0.

## **RESULTS AND DISCUSSION**

### Chlorophyll fluorescence

As can be seen in Table 1, maximum photochemical efficiency of PSII ( $F_v/F_m$ ) and maximum fluorescence ( $F_m$ ) in *C. caucasica* decreased significantly after 5 days drought. During the experiment, the lowest amount of  $F_v/F_m$  reaching 0.254 after 9 days withholding water.

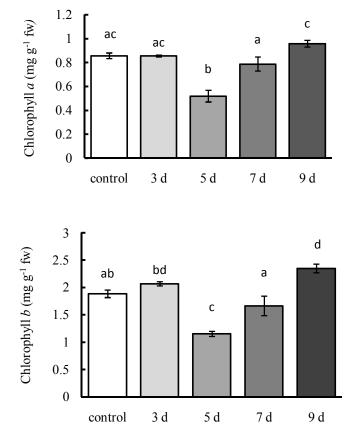
Table 1. Fo, Fm and Fv/Fmvalues in <i>C. caucasica</i> L. under different watering regimes. Data were expressed as mean ± SE			
Treatment	F <sub>0</sub> Mean+SE	F <sub>m</sub> Mean+SE	F <sub>v</sub> /F <sub>m</sub> Mean+SE
3d	1.607±0.109 a	6.272±0.204 a	0.740±0.008 a
5d	1.686±0.110 a	5.872±0.287 a	0.695±0.111 b
7d	1.731±0.127 a	6.023±0.251 a	0.685±0.020a b
9d	1.927±0.166 a	3.236±0.416 b	0.254±0.066 c

Note: Data in the same column followed by different small letters are significantly different at 0.05 level of significance.

The amount of  $F_m$  in 9 days drought has been significant differences with other treatments with the measure of 3.236 ( $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>). The minimal fluorescence (F<sub>0</sub>) was constant throughout the entire course of the experiment. Most studies showed that under drought conditions, the reduction of photosynthesis associated with disorder in biochemical reactions[14]. Drought damages the reaction center of photosystem II [8], [9], [20]. Hence the reduction of the maximum photochemical efficiency of photosystem II  $(F_v/F_m)$  and fluorescence changes in the time interval is used as a criterion of tolerance and stress resistance[12]. We found out that the maximum photochemical efficiency of photosystem II  $(F_v/F_m)$  and maximum fluorescence  $(F_m)$  had a downward trend with increasing the drought stress. This is in agreement with [4] who reported the reduction of  $F_v/F_m$  and  $F_m$  in seedlings of *Pinushalepensis*, Quercuscoccifera and Quercus ilex. We also showed that C. caucasica had significant differences in the ratio of  $F_v/F_m$  since 5 days and in  $F_m$  since 9 days after drought. The high level of  $F_v/F_m$  and  $F_m$  indicated the high amount of photosynthesis. The  $F_v/F_m$  represents the electron transport capacity of photosystem II [10]. That has high correlation with quantum yield of net photosynthetic [2]. According to Alidib et al., [1] decrease of efficiency of photosystem II centers to consumption of photons, exhibited the measure of photo inhibition under drought stress conditions. The decrease in F<sub>m</sub> revealed that the lower oxidation of Q<sub>A</sub> under drought conditions and reduced photochemical reactions.

The rapid changes in fluorescence that occur during the rapid induction to a peak have long been attractive for detecting differences in photosynthetic performance in plants. On immediate exposure to light, fluorescence rises to the minimum level of fluorescence, termed  $F_0$  level, which is the fluorescence level obtained when the PSII reaction centers are in the open state, i.e., capable of photochemistry since  $Q_A$ , the primary quinone acceptor of PSII, is maximally oxidized. The results indicated that the minimum fluorescence in *C.caucasica* was not significantly affected by drought in different treatments. This result is in agreement with findings of [11] and [22]. Since  $F_0$  values are related to chlorophyll fluorescence of PSI receptors [2], [27] and considering that non-significant difference of  $F_0$  between irrigation regimes, it seems the receptors of chlorophylls had almost a similar efficiency between irrigation regimes. *Chlorophyll content* 

With the drought process, Chlorophyll a, Chlorophyll b, and total Chlorophyll in *C. caucasica* had significant upward trend. Chlorophyll a, Chlorophyll b, and total Chlorophyll in 9 days drought were 1.1, 1.2, 1.2 times, respectively, in compare to that of the control.



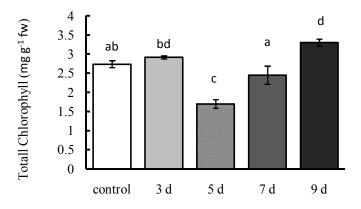


Fig.1. Effects of different watering regimes on chlorophyll a, chlorophyll b, and total chlorophyll of C. caucasica leave. Values are mean  $\pm$  SD and bars indicate standard error.

Chloroplast pigments play an important role for the light absorption and conversion in the photosynthesis process. Generally in most plants the contents of photosynthetic pigments (chlorophyll) are reduced with increasing drought [7]. Research showed that the chlorophyll content in plants with low resistance is decreased with increasing the drought [17], [4]. which is in contradiction with our results. In our experiments, chlorophyll content of *C. caucasica* increased with drought deepening. Guan *et al* [16] found that total chlorophyll content in Moslach chinensis increases with decreasing soil water content. The increase in chlorophyll content may be associated with the decrease of leaf water content during the drought [19]. This may compensate the reducing leaf area under drought stress. Increase in chlorophyll content on one hand and reducing leaf area on the other can be attributed to the maintenance of photosynthetic rate under drought stress [24]. Drought stress damages the reaction centers of photosystem II [8], [9]. In our study, the  $F_v/F_m$  value decreased with increasing the levels of stress. This decrease was very small until 7 days after drought, reflecting *C. caucasica* seedlings has a strong ability to tolerate drought stress.

We concluded that *C. caucasica* seedlings are resistant to drought stress. *C. caucasica* seedlings can increase its chlorophyll content and on the other hand the decline rate of the photochemical efficiency of photosystem II is very small. Since the drought damages the photosynthetic apparatus via reducing the capacity of electron acceptance, it has a negative impact on the photochemical efficiency of photosystem II [22]. This indicates the higher photosynthetic efficiency of *C. caucasica* under drought conditions. The results of chlorophyll fluorescence confirm that a significant increase in chlorophyll content under stress enhances the excitation capacity of photosystem II being one of the physiological indicators of stress resistance. *C. caucasica* would be one of the most suitable species for afforestation plans in arid and semiarid regions.

#### REFERENCES

- 1. Ali-Dib T., Monneveux PH., Acevedo J. and Nachil MM. (1994). Evaluation of praline analysis and chlorophyll fluorescence quenching measurements as drough tolerance indicators in durum wheat (*Triticum turgidum* L. Var. durum). Euphytica.: 79 (1-2): 65-73.
- 2. Anonymous: (1993). An introduction to fluorescence measurements with the plant efficiency analyzer.(PEA) Hansatech Instruments Ltd. England.
- 3. Arnon DI: (1949). Copper enzymes in isolated chloroplast polyphenol oxidase in Beta vulgaris. Plant Physiol. 24:1-15.
- Baquedano FJ., Castillo FJ: (2006). Comparative ecophysiological effects of drought on seedlings of the Mediterranean water-saver Pinushalepensis and water-spenders Quercuscoccifera and Quercus ilex. *Trees.* 20: 689–700.
- 5. Barker DJ., Sullivan CY. and Moser. (1993). LE1 Water deficit effects on osmotic potential, cell wall elasticity, and pralin in five forage grasses. *Agronomy Journal*. 1993: 85: 270-275.
- 6. Bjorkman O., Demming B: (1987). Photon yield of O<sub>2</sub> evolution and chlorophyll fluorescence characteristics at 77 K among vascular plants of diverse origins. Planta.: 170: 489-504.
- 7. Burce JA: (1991). Comparative responses of leaf conductance to humidity in single attached leaves. *Journal of Experimental Botany*. 32: 629-634.
- 8. Comic G., Briatais JM:(1991). Partioning of photosynthetic electron flow between Co<sub>2</sub>& O2 reduction in a C3 leaf (*Phaseolus vulgaris* L.) at different Co<sub>2</sub> concentration & during drought stress. Planta. 183: 178-184.

- 9. Cornuc G., Baker NB and Bower (eds) JR: (1994). Drought stress and high light effect on leaf photosynthesis. In Photoinhibition of photosynthesis from molecular mechanisms to the field. Oxford, UK. Bios Scientific Publishers. 297-313.
- 10. Fischer RA., Rees D., Sayre KD., Lu ZM., Candon AG. and Saavedra AL: (1998). Wheat yield progress associated with higher stomatal conductance and photosynthetic rate, and cooler canopies. Crop Science. 38: 1467–75.
- 11. Flagella Z., Pastore D., Campanile RG. and DiFonzo N: (1994). Photochemical quenching of chlorophyll fluorescence and drought tolerance in different durum wheat (*Triticum durum*) cultivars. Agriculture Science, Cambridge. 122: 183-192.
- 12. Fracheboud Y: (2006). Using chlorophyll fluorescence to study photosynthesis. Institute of Plant Sciences ETH, Universitatstrass. CH-8092 Zurich.
- 13. Giardi MT., Colia A., Geikeo D., Kucera T., Masojidek J. and Matto AK: (1996). Long term drough stress induces structural and functional reorganization of photosystem II. Planta. 1996: 199: 118-125.
- 14. Graan T., Boyer JS: (1990). Very high CO<sub>2</sub> partially restores photosynthesis in sunflower at low water potentials. Planta. 1990:181: 378-384.
- 15. Greaves JA., Wilson JM: (1987). Chlorophyll fluorescence analysis an aid to plant breeders. Biologist. 1987: 34: 209-14.
- 16. Guan BH.,Ge YF. and Fan MY *et al*: (2003). Phenotypic plasticity of growth and morphology in Moslachchinensis responds to diverse relative soil water content. Journal of Actaecologica Sinica. 23(2): 259-263
- 17. HascEL.SunYM, and Li J. (2006). The influence of draught stress on the physiological indexes of seedlings of four tree species in Maowusu. Journal of Forest Research. 19(3): 358-364.
- 18. Havaux M., Emez M. and Lannoye R: (1998). Selection de varietes de bledur (*Triticum durum* Desf.) etdeble tender (*TriticumaestivumL.*) adapted a la secberesse par I mesure de I extinction fluorescence de la chlorophylle in vivo. Agronomie. 1998: 8(3): 193-199.
- 19. Jun-ya Y., Aiko O., Yuko H. Eimiko M. and Yasumaro K: (2003). Effect of high light and low temperature during winter on needle photo damage of Abiesmarieseii growing at the forgets limit on Mt.Norikura in Centural Japan [J]. Plant Science. 165:257-264.
- 20. Kuchaki A., Soltani A. and Azizi M: (1997). Plant ecophysiology. Mashhad's Jahad Publisher.Vol:1 P 271 (In Farsi).
- 21. Lu Q., Lu C., Zhang J. and Kuang T: (2002). Photosynthesis and chlorophyll a fluorescence during flag leaf senescence of field grown wheat plants. *Journal of Plant Physiol*. 159: 1173-1178.
- 22. Maxwell K., Johnson GN (2000). Chlorophyll fluorescence a practical guide. *Journal of Expt*. Botany. 51(345): 659-668.
- 23. Rigoberto RS., Josue AAG, and Calros TL et al:(2004). Biomass distribution, maturity acceleration and yield in drought stressed common bean cultivars. Journal of Field Crop Research. : 85: 203-211.
- 24. Smorenburg K., Courreges-Lacoste GB., Berger M., Bushmann C., Court A., Del Bello U., Langsdorf G., Lichtenthaler HK., Sioris C., Stoll MP., Visser H., Deall J R. and Toivonen PMA eds: (2003). Practical Applications of Chlorophyll Fluorescence In Plant Biology. Kluwer Academic Publishers Boston, Dordrecht, London. 4542: 178–190.
- 25. Tezara W., Mitchell VJ., Driscoll SD. and Lawlor DW: Water stress inhibits plant photosynthesis by decreasing coupling factor and ATP, Nature. 1999: 401: 914-917.
- 26. Wilson JM., Greaves JA: (1993). Development of fluorescence based screening programs for temperature and water stress in crop plants. Adaptation of Food Crop to Temperature and Water Stress. A VRDC, Shanhua, Taiwan. p: 389-398.
- 27. Yin CY., Duan BL., Wang X. and Li CY: (2004). Morphological and physiological responses of two contrasting Poplar species to drought stress and exogenous abscisic acid application. Plant Science. 2004: 167: 1091–1097.
- 28. Zhang, X., Wu, N., Li, C. (2005). Physiological and growth responses of Populusdavidiana ecotypes to different soil water contents. *Journal of. Arid Environment*. 2005: 60: 567-579,