Advances in Bioresearch Adv. Biores., Vol 9 (2) March 2018: 67-72 ©2018 Society of Education, India Print ISSN 0976-4585; Online ISSN 2277-1573 Journal's URL:http://www.soeagra.com/abr.html CODEN: ABRDC3 DOI: 10.15515/abr.0976-4585.9.2.6772

Advances in Bioresearch

# **ORIGINAL ARTICLE**

# Comparative analysis of Antibiotic Susceptibility pattern in Transgenic Cotton varieties

P. Srikanth, M. Devasahayam, R. Singh and S. A. Masih\*

Centre for Transgenic Studies, Sam Higginbottom University of Agriculture, Technology and Sciences, Allahabad (UP), India, 211007

\*Email: sam.masih@shiats.edu.in

## ABSTRACT

Transgenic Bt cotton is one of the most adopted transgenic crop throughout the world. Transgenic Bt cotton is considerably effective in controlling lepidopteran pests owing to the presence of Cry genes such as Cry 1Ac (Bollgard I), Cry 1Ac + Cry 2Ab (Bollgard II). In the present study a field trail of Bollgard I and bollgard II of Bt cotton were conducted cyclohexamide and cyclohexamide + kanamycin resistance soil bacteria were isolated from Bt cotton and nBt cotton of Bollgard I and Bollgard II of Bt cotton. NptIIgene was isolated from Bollgard I and Bollgard II of Bt cotton. The results were found as cyclohexamide in Bollgard II were found 23.29% higher than Bollgard I of Bt cotton. cyclohexamide + kanamycin in Bollgard II were found 8.9% higher than Bollgard II of Bt cotton, resistance against cyclohexamide in Bollgard I were found 6.5% higher than Bollgard II of Bt cotton. NptII gene in Bollgard I and Bollgard II of Bt cotton resistance against cyclohexamide I were found 6.5% higher than Bollgard II of Bt cotton. NptII gene in Bollgard I and Bollgard II of Bt cotton seeds were similarly found.

Key words : Transgenic Bt cottonCyclohexamide, Kanamycin, NptII and soil bacteria CFU.

Received 21.10.2017

Revised 10.12.2017

Accepted 22.02.2018

### How to cite this article:

P. Srikanth, M. Devasahayam, R. Singh and S. A. Masih. Comparative analysis of Antibiotic Susceptibility pattern in Transgenic Cotton varieties. Adv. Biores., Vol 9 [2] March 2018.67-72.

## INTRODUCTION

Transgenic *Bt* cotton is one of the most adopted transgenic crop throughout the world [1].Transgenic *Bt* cotton is considerably effective in controlling lepidopteran pests owing to the presence of Cry genes such as Cry 1Ac (Bollgard I), Cry 1Ac + Cry 2Ab (Bollgard II). Moreover, they are beneficial to the grower and the environment as they reduce chemical insecticides. However, poor performance of the transgenic traits during boll period and variable performance between different regions has been reported [2].

These former bacterial genes could be transferred more easily than other plant genes to soil bacteria because of a high degree of homology facilitating recombination in potential bacterial recipients[3]. *Bt*-toxin from *Bt* cotton plants introduced into the soil through two pathways, i.e., biomass bin corporation and root exudates [4].

Some studies indicate that *Bt* cotton has no negative effects on soil flora and fauna and may even have beneficial effects[4].While some studies have reported that *Bt*cotton creates adverse effects [5].Similarly *Bt*cotton contain antibiotic resistance markers such as the neomycin phosphor transferase II (*nptII*) gene which confers resistance to kanamycin and neomycin. The presence of antibiotic resistance genes in transgenic crops has raised concerns over the possible transfer of antibiotic resistance genes from *Bt* cotton to soil bacteria [6].

The study has been conducted to find out the CFU of (Cyclohexamide and Cyclohexamide + kanamycine) resistance soil bacteria from bollgard I and Bollgard II of soil samples were analyzed and the presence of *nptII* gene were identify in the seed samples of the Bollgard I and Bollgard II of *Bt* cotton in this experiment.

# **MATERIAL AND METHODS**

*Bt* cotton seeds such as Bollgard I (cotton 1007- 9810 BG1) and Bollgard II (KCH14K59) Jaddu seeds were obtained from local market,nptII primers [7] were obtained from Merck and Chromus, India. The sequence of the primers are as follows *nptII* forward primer 5' CTCACCTTGCTCCTGCCGAGA3'; nptII reverse primer 5' CGCCTTGA GCCTGGCGAACAG 3'; Taq DNA polymerase and dNTPs were from Merck. *BL21 DE3*.

## **Field Trials**

The field trails were conducted at the horticulture field and central field of Sam Higginbottom University of Agriculture Technology and Sciences, Allahabad, Uttar Pradesh. Two plots were

maintained at a spacing of 0.65m x 0.65m and 0.35 x 0.35m with alternatively sown *Bt* and *nBt* of bollgard I seeds. The selected experimental field was harrowed and ploughed with a cultivator and prepared for sowing. Before sowing the field received one full dose of an organic source of field yard manure (FYM) 1.25 gm/m2 were applied in the field. *Bt*cotton and non *Bt*-cotton was be sowed in June 2016. Three plots containing seven rows and seven columns each were maintained at a spacing of 0.65m x 0.65m with 10%, 25% and 50% of refuge n*Bt*cotton for Bollgard II, one plot containing eleven rows and seven columns. The recommended agronomic and crop management practices was been followed thereafter. While no pesticide was used during the trial, irrigation was at 10 day intervals initially and before flowering irrigation was done, hand weeding were done one month inter well.

Isolation of Cyclohexamide and Kanamycin Resistant soil bacteria from Bt Cotton soil Samples

The soil sample of Bollgard I were collected at 4 and 7 moth after sowing of *Bt* cotton, for bollgard II soil were collected for every two months intervals for three times the crop growing period. Soil samples were collected at 5-7cm depth from the rhizosphere of Bt Cotton and n*Bt*cotton separately. 2gm of soil was mixed with 15ml PBS and left at room temperature with shaking for 30 minutes in a sterile falcon tube [8]. The mud was removed by centrifugation at 600g for 10 minutes and the supernatant containing bacteria was collected for kanamycin and cyclohexamide screening. 1:10 dilution was plated on cycloheximide + Kanamycin (c+k) and 1:100 cycloheximide (c) LB plates respectively with both cycloheximide and kanamycin at 50 µg/ml. Plates were incubated at 25°C and colonies counted after 1 day for *c* and 2 days for c+kplates and recorded as colony forming units/ 2 mg of soil. When the cLB plates contained a lawn of colonies these plates were not counted while their corresponding c+kLB plates were counted. The plates containing colonies were stored for possible further analysis.

# NptII Detection in DNA from Soil Bacteria

*NptII* detection was done by PCR using specific primers [7]. The PCR reaction was using Taq DNA polymerase with 2.0µl primers, 0.2 µl dNTPs, Taq DNA polymerase 1.0 µl. The reaction condition was 98°C for 10 minutes followed by 40 cycles at 95°C – 30 seconds; 50°C-30 seconds and 72°C – 30 seconds. This was followed by 1 cycle at 72°C for 10 minutes. The positive control for *nptII*gene was *BL21 DE3* containing incorporated cry1ACgene. The PCR product was analyzed on a 2% agarose gel.

## **Correlation Coefficient Calculation**

Phenotypic correlations were estimated using the standard procedure [9] from the corresponding variance and covariance components using the following equation

$$pxy = \frac{\sigma pxy}{\sqrt{\sigma px} x \sigma py}$$

Where, r pxy = phenotypic correlation coefficient between characters X and Y ; $\sigma$ pxy the covariance for X and Y and,  $\sigma$ px and  $\sigma$ py the variance for the two characters X and Y.

CFU/ ml =Number of conilies × Dilution factor /volume of the culture plate

# **RESULTS AND DISCUSSION**

The colony forming unit(CFU) of soil bacteria against cylohexamide (C) and Cyclohexamide + kanamycin (C+K) of bollgard at 0.65m spacing of Bollgard I *Bt* cotton (Table.1) were found as mean 77.41 × 10<sup>3</sup> CFU. g<sup>-2</sup>(C), 19.58 × 10<sup>2</sup> (C+K). In the spacing of 0.30m the cyclohexamide were found as  $70.28 \times 10^3$  CFU. g<sup>-1</sup>(C), were as in cyclohexamide + kanamycin (C+K) were found as  $13.3 \times 10^2$  CFU. g<sup>-2</sup>, for the n*Bt* cotton were found as mean 40 × 10<sup>3</sup> CFU. g<sup>-1</sup>(C), 3.4 × 10<sup>2</sup> CFU. g<sup>-2</sup> (C+K). In the spacing of 0.30m the cyclohexamide were found as  $52.7 \times 10^3$  CFU. g<sup>-2</sup>(C), were as in cyclohexamide + kanamycin (C+K) were found as  $2 \times 10^2$  CFU. g<sup>-2</sup>.

Similarly for the second time colony forming unit(CFU) of soil bacteria of Bollgard I of *Bt* cotton(Table.2) against cylohexamide (C) and Cyclohexamide + kanamycin (C+K) of bollgard at 0.65m spacing of *Bt* cotton were found as mean  $36 \times 10^3$  CFU. g<sup>-2</sup>(C),  $10.4 \times 10^2$  CFU. g<sup>-2</sup>(C+K). In the spacing of 0.30m the cyclohexamide were found asmean  $45 \times 10^3$  CFU. g<sup>-2</sup>(C), were as in cyclohexamide + kanamycin (C+K) were found as 3, for the n*Bt* cotton were found as 77 × 10<sup>3</sup> CFU. g<sup>-2</sup>(C),  $10 \times 10^2$  CFU. g<sup>-2</sup> (C+K). In the

spacing of 0.30m the cyclohexamide were found as mean 86.4 ×  $10^3$  CFU. g<sup>-2</sup>(C), were as in cyclohexamide + kanamycin (C+K) were found as  $4.88 \times 10^2$  CFU. g<sup>-2</sup>.

These results were similarly found in  $Bt \operatorname{corn} 47 \ge 10^6 \operatorname{CFU}$ . g<sup>-1</sup>.Icoz *et al.* [10]who reported that after 4 consecutive years of corn cultivation. Muchaonyerwa*et al.* [11]reported that antibiotic resistance soil bacteria in Bt maize could persist in tropical soils as a result of adsorption on soil clays, but that there were no observable effects on the soil microbial biomass carbon or counts of culturable bacteria. Rui *et al.* [12] found increased numbers of culturable functional groups of bacteria in rhizosphere soil of Bt cotton in the early and middle stages of growth of cotton. But there was no significant difference on the numbers of these groups.

The colony forming unit(CFU) of *Bt* plants soil bacteria against cylohexamide (C) of bollgardII (Table.3) at 0.65m spacing of Bollgard II *Bt* cotton at the month of July were found as mean  $80 \times 10^3$  CFU. g<sup>-2</sup> in 50% *Bt* with 50% n*Bt*, 75% *Bt* with 25% n*Bt* and 90% *Bt* with 10% n*Bt* similarly and in *Bt* with border refuge of both sides were found as 31.666 × 10<sup>3</sup> CFU. g<sup>-2</sup> in 50% *Bt*, 287 × 10<sup>2</sup> CFU. g<sup>-2</sup> in 75% *Bt* with 25% n*Bt*, 48.333 × 10<sup>2</sup> CFU. g<sup>-2</sup> and final treatment were found as 49 × 10<sup>2</sup> CFU. g<sup>-2</sup> in *Bt* with border refuge of both sides.

Where as in n*Bt*cotton the results were found as mean  $80 \times 10^3$  CFU (Table .3). g<sup>-2</sup> in 50% *Bt* with 50% n*Bt*, 75%*Bt* with 25%*nBt* and 90%*Bt* with 10%*nBt* similarly and in *Bt* with border refuge of both sides were found as  $185 \times 10^3$  CFU. g<sup>-2</sup>, were as in C+K the CFU colonies were found as mean 4.666 ×  $10^2$  CFU. g<sup>-2</sup> in 50% *Bt* with 50%*nBt*, 204 ×  $10^2$  CFU. g<sup>-2</sup> in 75%*Bt* with 25%*nBt*, 276 ×  $10^2$  CFU. g<sup>-2</sup> and final treatment were found as  $364 \times 10^2$  CFU. g<sup>-2</sup> in *Bt* with border refuge of both sides.

Table. 1 The *Bt* cotton and *nBt* cotton of plant soil samples from the 0.65m and 0.3m on c and c+k plates had a mean colon forming units (CFU) of Bollgard I

plates	nau a i	incan c		ming u	into (C	10,0	Dunga	lui		
		Bt c	otton	n <i>Bt</i> cotton						
Sapcing	0.	65	0.1	3	0.6	5	0.3	3		
	С	C+K	С	C+K	С	C+K	С	C+K		
n	75	79	21	21	9	10	5	5		
mean	77.41	19.58	70.28	13.3	40	3.4	52.7	2		
SD	47.5	31.53	43.4	16.41	17.4	1.82	15.35	1.58		
CV (%)	61.4	161	61.75	123.1	42.75	53.5	26.8	79		
range	2-208	0-177	35-197	0-45	19-74	1-6	31-70	0-4		
_	_					-		_		

Table. 2 The *Bt* cotton and *nBt* cotton of plant soil samples from the 0.65m and 0.3m on cand c+k plates had a mean colon forming units (CFU) of Bollgard I

plates had a mean color for ming ands (or of or Dongara i														
		Bt co	tton		n <i>Bt</i> cotton									
Sapcing	0.65		0.3		0.65		0.3							
	С	C+K	С	C+K	С	C+K	С	C+K						
n	141	139	36	37	28	28	9	9						
mean	36	10.4	45	3	77	10	86.4	4.88						
SD	27.8	17.4	59	5.47	42	24.74	27.7	5						
CV (%)	77.3	167.7	131	183	54.5	247.4	32	100.8						
range	0-164	0-109	3-234	0-30	2-180	0-52	43-128	0-14						

The colony forming unit(CFU) of *Bt* plants soil bacteria against cylohexamide (C) of bollgard at 0.65m spacing of Bollgard II *Bt* cotton at the month of September were found as mean 80 × 10<sup>3</sup>(Table.4) CFU. g<sup>-2</sup> in 50% *Bt* with 50% n*Bt*, 75% *Bt* with 25% n*Bt* and 90% *Bt* with 10% n*Bt* were found s 18× 10<sup>3</sup> CFU. g<sup>-2</sup> and in *Bt* with border refuge of both sides were found as 50 × 10<sup>3</sup> CFU. g<sup>-2</sup>, were as in C+K the CFU colonies were found as mean 3 × 10<sup>2</sup> CFU. g<sup>-2</sup> in 50% *Bt* with 50% n*Bt*, 5 × 10<sup>2</sup> CFU. g<sup>-2</sup> in 75% *Bt* with 25% n*Bt*, 0.333 × 10<sup>2</sup> CFU. g<sup>-2</sup> and final treatment were found as 0.666 × 10<sup>2</sup> CFU. g<sup>-2</sup> in *Bt* with border refuge of both sides.

Where as in nBtcotton the results were found as mean  $80 \times 10^3$  (Table.4)CFU. g<sup>-2</sup> in 50% Bt with 50% nBt, 75%Bt with 25%nBt and 90%Bt with 10%nBt were found as 49.666× 10<sup>3</sup> CFU. g-2 and in Bt with border refuge of both sides were found as 21 × 10<sup>3</sup> CFU. g<sup>-2</sup>, were as in C+K the CFU colonies were found as mean 26 × 10<sup>2</sup> CFU. g<sup>-2</sup> in 50% Bt with 50%nBt, 58.666 × 10<sup>2</sup> CFU. g-2 in 75%Bt with 25%nBt, 0.666 × 10<sup>2</sup> CFU. g-2 and final treatment were found as 0.333 × 10<sup>2</sup> CFU. g<sup>-2</sup> in Bt with border refuge of both sides.

The colony forming unit(CFU) of *Bt* plants soil bacteria against cylohexamide (C) of bollgard at 0.65m spacing of Bollgard II *Bt* cotton at the month of December were found as mean 53.333 × 10<sup>3</sup> CFU. g<sup>-2</sup> in 50% *Bt* with 50% n*Bt*, 75% *Bt* with 25% n*Bt* were found as 82.666 × 10<sup>3</sup> (Table.5) CFU. g-2, 90% *Bt* with 10% n*Bt* were found s 85.333 × 10<sup>3</sup> CFU. g-2 and in *Bt* with border refuge of both sides were found as 29.333 × 10<sup>3</sup> CFU. g<sup>-2</sup>, were as in C+K the CFU colonies were found as mean 10.333 × 10<sup>2</sup> (Table.5) CFU. g-2 in 50% *Bt* with 50% n*Bt*, 3 × 10<sup>2</sup> CFU. g<sup>-2</sup> in 75% *Bt* with 25% n*Bt*, 4.333 × 10<sup>2</sup> CFU. g<sup>-2</sup> and final treatment were found as 7.666 × 10<sup>2</sup> CFU. g-2 in *Bt* with border refuge of both sides.

									8			,		0		<u>()</u> ,	,			
tails	Bt	Plants									nBt	Plants								
Treatment details	С					C+K					С					C+K				
Treatn	u	mean	SD	CV(%)	range	u	mean	SD	CV(%)	range	n	mean	SD	CV(%)	range	u	mean	SD	CV(%)	range
50% Bt with 50% Bt	3	80	0.00	0.00	0	3	3.666	3.518	95.776	7	3	80	0.00	0.00	0	3	4.666	4.728	101.264	9
75%Bt with 250%nBt	3	80	0.00	0.00	0	3	287.333	210.732	73.347	366	3	80	0.00	0.00	0	3	204.000	204.470	100.234	360
90%Bt with 10%*Bt	3	80	0.00	0.00	0	3	48.333	42.525	87.987	80	3	80	0.00	0.00	0	3	276.000	63.490	23.005	120
Bt with border refuge of both cidor	3	31.666	41.868	132.192	73	3	67	78.889	160.998	140	3	185.000	67.352	36.403	125	3	364.000	229.188	62.962	412

Table 3. The *Bt* cotton and n*Bt* cotton of plant soil samples from the 0.65m and 0.3m on c and c+kplates had a mean colon forming units (CFU) of Bollgard II ( July)

 Table. 4 The *Bt* cotton and n*Bt* cotton of plant soil samples from the 0.65m and 0.3m on c and c+k plates had a mean colon forming units (CFU) of Bollgard II ( September)

									0	inteo (						pren		/		
slis	Bt	Plants									nBt	<b>FIAILS</b>								
nt deta	С					X+J					С					X+J				
Treatment details	n	mean	SD	CV(%)	range	n	mean	SD	CV(%)	range	u	mean	SD	CV(%)	range	u	mean	SD	CV(%)	range
50% Bt with 50%nBt	3	80	0.00	0.00	0	3	3.000	2.647	88.197	5	3	80	0.00	0.00	0	3	26	25.154	96.762	45
75%Bt with 25%nBt	3	80	0.00	0.00	0	3	5.000	2.000	40.000	4	3	80	0.00	0.00	0	3	58.666	36.954	62.986	64
90%Bt with 10%nBt	3	18.000	4.585	25.457	6	3	0.333	0.573	173.200	1	3	49.666	34.580	69.643	68	3	0.666	1.157	173.200	2
Bt with border refuge of both sides.	3	50	27.079	79.623	50	3	0.666	1.157	173.200	2	3	21.000	8.881	42.327	17	3	0.333	0.573	173.200	1

plates had a mean colon forming units (CFU) of Bollgard II ( December)	Table. 5 The <i>Bt</i> cotton and <i>nBt</i> cotton of plant soil samples from the 0.65m and 0.3m on cand c+k
	plates had a mean colon forming units (CFU) of Bollgard II (December)

		piac	00 110	uun	lean	001			-6 uii	105 (	01 0	<u>, , , , , , , , , , , , , , , , , , , </u>	ongai	un	( 200	cemp	crj			
S	Bt	Flants									$\mathrm{n}Bt$	Plants								
nt detail	С					C+K					С					C+K				
Treatment details	u	mean	SD	CV(%)	range	n	mean	SD	CV(%)	range	u	mean	SD	CV(%)	range	u	mean	SD	CV(%)	range
50% Bt with 50%nBt	3	53.333	46.180	86.605	80	3	10.333	9.077	87.816	18	3	53.333	46.180	86.605	80	3	10.333	11.842	114.460	21
75%Bt with 25%nBt	3	82.666	4.618	5.582	8	3	3.000	1.730	57.730	3	3	86.000	10.393	12.080	18	3	10.666	6.111	57.281	12
90%Bt with 10%nBt	3	85.333	9.236	10.823	16	3	4.333	2.887	66.613	5		80.000	0.000	0.000	00	3	6.000	3.461	57.730	9
Bt with border refuge of both sides.	3	29.333	2.516	8.573	5	3	7.666	7.231	94.358	13	3	80.000	0.000	0.000	00	3	10.000	4.000	40.000	8

Where as in nBt cotton the results were found as mean  $53.333 \times 10^3$  (Table.5) CFU.g<sup>-2</sup> in 50% Bt with 50% nBt were found as 86 × 10<sup>3</sup> CFU. g<sup>-2</sup>, 75% Bt with 25% nBt, 90% Bt with 10% nBt were found as 80 × 10<sup>3</sup> CFU. g<sup>-2</sup> and in Bt with border refuge of both sides were found as 80 × 10<sup>3</sup> CFU. gv, were as in C+K the CFU colonies were found as mean 10.333 × 10<sup>2</sup> (Table.5) CFU. g-2 in 50% Bt with 50% nBt, 10.666 × 10<sup>2</sup> CFU. g<sup>-2</sup> in 75% Bt with 25% nBt, 6 × 10<sup>2</sup> CFU. g<sup>-2</sup> and final treatment were found as 10 × 10<sup>2</sup> CFU. g-2 in Bt with border refuge of both sides.

These results were similarly found by Pindi and Sultana [13] research results revealed that non Bt cotton plant growth is more than Bt cotton plant growth and rhizosphere soil sample of non Bt cotton has shown increased number of antibiotic. In the studies of Tesfaye *et al.* [14] the CFU were found as 6.3 x 10<sup>3</sup> CFU. g-1 dry soil in non Bt cotton at maturity stage of cotton growth in the field.

Differences in the composition of crop residues as the result of the introduction of transgenic traits have been observed in transgenic Bt crops [15]. Other studies have shown that the effects of GM plants on microbial communities depend more on seasonal variations or to other environmental factors, such as soil type and agricultural practices than to expression of Cry or other proteins in plants [16]. The Bollgard I of *Bt* cotton *NptII* results were similarly were found as Singh *et al.* [17].

# CONCLUSIONS

The proportion of indigenous soil bacteria resistance against cyclohexamide in Bollgard II were found 23.29% higher than Bollgard I of *Bt* cotton and in the n*Bt* cotton, resistance against cyclohexamide in Bollgard II were found 21.5% higher than Bollgard I of *Bt* cotton. The proportion of indigenous soil bacteria resistance against cyclohexamide +kanamycin in Bollgard I were found 8.9% higher than Bollgard II of *Bt* cotton and in the n*Bt* cotton, resistance against cyclohexamide +kanamycin in Bollgard I were found 8.9% higher than Bollgard II of *Bt* cotton and in the n*Bt* cotton, resistance against cyclohexamide + kanamycin in Bollgard I

were found 6.5 % higher than Bollgard II of *Bt* cotton.*NptII* gene in Bollgard I and Bollgard II of *Bt* cotton seeds were similarly found.

## ACKNOWLEDGEMENT

The authors acknowledge Prof. (Dr.) R.B. Lal, Vice Chancellor and Prof. (Dr.) S.B. Lal, Dean Faculty of Agriculture and Dr. Sunil Zacharia for usage of the DBT referral laboratory and UGC-Rajiv Gandhi National Fellowship (grant no.F1-17.1/2014-15) for financial supporting Doctoral program of PerumallaSrikanth.

### REFERENCES

- 1. Ismael, Y., Bennett, R. & Morse, S. (2002). Benefits from Bt cotton use by small holder farmers in South Africa. Agric. Bio. Forum. J. Agrobio. Mana.Eco., 5(1): 1-5.
- 2. Olsen, K. M. & Daly, J. C. (2000). Plant-toxin interactions in transgenic Bt cotton and their effects on mortality of *Helicoverpaarmigera*. Entomol. Soc. Am., (93): 1293-1299.
- 3. Goldstein, D.A. (2005). Human safety and genetically modified plants: A review of antibiotic resistance markers and future transformation selection technologies. J ApplMicrobiol(99):7–23.
- 4. Saxena, D. &Stotzky, G. (2001.) *Bacillus thuringiensis*(Bt) toxin released from rootexudates and biomass of Bt corn has no apparent effect on earthworms, nematodes, protozoa, bacteria, and fungiin soil. Soil BiolBiochem., (33):1225–30.
- 5. Cui, J. &Xia, J. (2000). Effects of transgenic Bt cotton R93-6 on the insect community. ActaEntomol Sin., (43):43–51.
- 6. Dröge, M., Pühler, A., &Selbitschka, W. (1998). Horizontal gene transfer as a biosafety issue: a natural phenomenon of public concern. J. Biotechnol. (64): 75-90.
- 7. Kamle, S., Kumar, A. &Bhatnagar, R. K. (2011). Development of multiplex and construct specific PCR assay for detection of cry2Ab transgene in genetically modified crops and product. GM Crops., (2): 74-81.
- 8. Ma, B. L., Blackshaw, R. E., Roy, J. & He, T. (2011). Investigation on gene transfer from genetically modified corn (Zea mays L.) plants to soil bacteria. Journal of Environmental Science and Health Part B., (46): 590-599.
- 9. Al-Tabbal, J. A. & Al-Fraihat, A. H. (2012). Genetic variation, heritability, phenotypic and genotypic correlation studies for yield and yield components in promising barley genotypes. J Agric Sci., (4):193-210.
- 10. Icoz, I., Saxena, D., Andow, D., Zwahlen, C. &Stotzky, G. (2007). Microbial populations and enzyme activities in soil *in situ* under transgenic corn expressing Cry proteins from *Bacillus thuringiensis*. J. Environ. Quality., (37):647-662.
- 11. Muchaonyerwa, P., Waladde, S., Nyamugafata, P., Mpepereki, S. &Ristori, G.G. (2005).Persistence and impact on microorganisms of *Bacillus thuringiensis*proteins in some Zimbabwean soils.Plant Soil, (266):41-46.
- 12. Rui, Y.K., Yi, G.X., Zhao, J., Wang, B.M., Li, Z.H., Zhai, Z.X., He, Z.P. & Li, Q.X. (2005). Changes of Bt toxin in the rhizosphere of transgenic Bt cotton and its influence on soil Functional bacteria. World J. Microbiol.Biotechnol., (21):1279-1284.
- 13. Pindi, P.K. & Sultana, T. (2013). Bacterial and fungal diversity in rhizosphere soils of Bt and non-Bt cotton in natural systems. Bulg. J. Agric. Sci., (19): 1306-1310.
- 14. Tesfaye, M., Temple, S.J., Allan, D.L., Vance, C.P. &Samac, D.A. (2001). Over-expression of malate dehydrogenase in transgenic alfalfa enhances organic acid synthesis and confers tolerance to aluminum. Plant Physiol., (127):1836-1844.
- 15. Poerschmann, J., A. Gathmann, Augustin, J., Langer, U. &Gorecki, T. (2005). Molecular composition of leaves and stems of genetically modified Bt and near isogenic non Bt maize characterization of lignin patterns. J. Environ. Qual., (34):1508-1518.
- 16. Fang, M., R.J. Kremer, P.P., Motavalli& Davis, G. (2005). Bacterial diversity in rhizospheres of nontransgenic and transgenic corn. Appl. Environ. Microbiol., (71):4132-4136.
- 17. Singh, C. K., Oiha, A, &Kachru, D. N. (2007). Detection and characterization of cry1Ac transgene construct in Bt cotton: multiple polymerase chain reaction approach.J AOAC Int., (90):1517-25.

**Copyright:** © **2018 Society of Education**. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.