REVIEW ARTICLE

Vital Role of Nanoparticles as Nanofertilizer in Crop Yield Improvement

¹Gunmala Gugalia, ²Anjali Sharma

¹ SDM PG Girls College, Bhilwara, Rajasthan ²Department of Botany, Sangam University, Bhilwara Corresponding author: Dr. Gunmala Gugalia Email: gunmala24@gmail.com

ABSTRACT

Nanofertilizers are nutrient transporters with nanoscale dimensions ranging from 30 to 40 nm that can retain a large number of nutrient ions extremely large surface area and dispense them slowly and surely in response to crop demand. Controlling the release of nutrients from fertilizer granules with nanofertilizers and composite materials can improve uptake of nutrients while limiting nutrient ions from being fixed or lost to the environment. Nanofertilizers have a high application efficiency, therefore nutrients may be transported to a rhizospheric target quickly. Keywords: Nanofertiliser, nanoparticles, rhizosperic target.

Received 24.05.2023

Revised 01.06.2023

Accepted 11.11.2023

How to cite this article:

Gunmala G, Anjali S. Vital Role of Nanoparticles as Nanofertilizer In Crop Yield Improvement. Adv. Biores., Vol 14 (6) November 2023: 586-595.

INTRODUCTION

Crop yield plateau, poor nutrient use efficiency, declining organic matter, lack of nutrients, arable farming, water resources and availability of labor due to agricultural migration are among the challenges affecting today's agricultural zones [1,2]. According to the Food and Agriculture Organization of the United Nations, depletion of land and water poses serious challenges to producing enough food and other agricultural commodities to sustain life and meet the needs of the world's ever-growing population [3]. Nanoscience, which generates ultra-small molecules with a remarkably high surface area to volume ratio and improved semiconductor-based and physico-chemical properties compared to bulk derivatives [4], is now considered as a potential mechanism for enhancing plant growth and development [5-8]. This notion is critical to mainstream agricultural science, in which farmers use technology to ensure efficient irrigation, fertilization, and other resources. Species-appropriate management increases agricultural sustainability by reducing waste and energy consumption. Nanoparticles and their compounds are being studied for a range of uses in agriculture, including increasing crop yields and reducing pesticide use. Agricultural strategies are used that are effective in increasing crop yield while polluting the soil or water. It can also help prevent nitrogen losses due to erosion and runoff along with soil microorganisms. Nanoscale sensors are used to monitor nutrient presence, pesticide use and contamination in agricultural products. Through the post-harvest process, extend the lifespan of agricultural products, nanopesticides eliminate pathogens, increase biomolecules and minerals in food, water treatment, use smart fertilizer, nano-fertilizer [9-12]. In this article, we discuss nano-fertilizers and their important role in improving crops.

ROLE OF FERTILIZER IN PLANT GROWTH

Fertilizers provide the nutrients that plants need to grow to thrive. Fertilizers are frequently widely used in agricultural via surface applications, beneath application, or in combination with irrigation water. However, a large portion of the fertiliser delivered by these methods is discharged into the environment or surface freshwater resources, causing harm to the biosphere [13,14]. Excess nitrogen, for example, is

Gugalia and Sharma

lost as NH3 by vaporization, released as N₂O or NO, or leached or drained into bodies of water. Excess phosphate, is from the other hand, becomes "trapped" in soil, where it creates synthesized connections with other elements including Ca-P, Mg-P, Al-P, Fe-P, and Zn-P, making it unavailable to plant absorption. Finally, rain bleaches the N and P elements into natural freshwater bodies, rivers and streams, and the sea, where they can cause pollution problems. Fertilizer use is increasing in lockstep with global population growth. Due to the fact that plants can only absorb about 42% of the phosphorus given to them, farmers use over 85% of the world's total mining phosphorus as nutrients [15]. If this scenario is correct, the world's phosphorus supply may run out in the next eighty years, limiting agricultural output. trends in urbanised states in the USA suggest a 77% reduction in cropland between 1978 and 1987 Farmland [16, 17]. Nowadays, farmers use more chemical fertilizers to meet food demand, which pollutes the soil and environmental health as well as depletes natural resources. It is possible to grow some crops with hydroponics, but it costs (in terms of energy and money) more than conventional farming, but these methods are not cost-effective nor sustainable in the long run. [16]. Therefore, it is essential to develop long-term strategies to yield higher agricultural and nutrient yields while consuming fewer resources and fertilizers. about the use of small quantities. The term "nano-fertilizers" is used in the literature on applications of nanotechnology in agriculture to describe both materials with a physical size of 1-100 nm in at least one dimension, and materials that exist. at the volumetric scale but modified with nanomaterials (eg, fertilizers coated with nanoparticles). Therefore, the term "nano-fertilizer" is used in this article to refer to both the actual nanoparticles and the nano-activated raw materials used as fertilizers. Nanoparticles have the potential to improve the mobility of minerals and nutrients such as phosphorus in the biosphere and may have an effect on the metabolic activities of plants compared with other products [18],19]. Nanotechnology-based agricultural inputs are shown in Figure 1. In the following sections, we will discuss recent papers on nanoparticles/nanomaterials used as nanofertilizers for growth of plants. The discussion is therefore structured so that farmers and researchers can better understand the fundamental aspects and applications of nano-fertilizers for precision and sustainable agriculture

NANOFERTILISERS

Nanofertilizers include all or part of nanostructured formulations that can be applied to plants and allow for efficient absorption or gradual release of the active ingredients. Because traditional bulk fertilizers are applied in large quantities, their absorption is very slow. Two main problems are related to chemical fertilizers that absorb less phosphorus and nitrogen. They changed their chemical form so that plants could not easily absorb and wash away rainwater and degrade soil fertility. As a result, hazardous carbon emissions (such as some nitrogen oxides) and eutrophication, have serious consequences for agricultural safety and the environment. Therefore, it is important to develop smart fertilizers that are easily absorbed by plants. Scientists are actively exploring the various uses of metal nanoparticles in plant science and agriculture as possible ways to achieve this goal in a sustainable and concise way. Therefore, nanotechnology is a possible approach to achieving this goal in a sustainable and appropriate way. In addition, the environmental, health and safety aspects of nanotechnology need to be considered to determine the effectiveness of nanofertilizers [20-22]. The plant receives the nanocomposite, which gives different results and benefits over traditional or ionic salts [23-26]. A kind of inorganic, organic, and hybrid nanomaterials were examined on lots of vegetation to peer how they have an effect on crop development, productivity, and yield. The desk blanketed data approximately nanoparticle characteristics, nanoscale properties, mechanism of nanoparticle delivery, examined seedlings, and studied responses due to the fact the consequences of nanomaterials and the corresponding plant responses are stimulated through a selection of things associated with nanoscale assets, soil, and eco system. The following subsections cross over particular examples of nanoparticles which have been used as nanofertilizers or nanonutrients.

ANALYTICAL METHODOLOGY

The intrinsic properties and external relations of nanoparticles have a significant impact on their effect on plants. This is likely among the many purposes why the research has discovered conflicting results from the same particle category. For example, TiO2 nanoparticles retarded corn seed germination [85-86], but had no effect on paddy seed germination and enhanced wheat seed germination. In the following section, we'll go over some of the factors to consider when assessing a nanoparticle type and its affect on plants. **Nanoparticle/nanofertilizer synthesis**

Nanomaterials are commonly synthesized by both "wet" methods such as solgel, hydrothermy, homogenous precipitation, biosynthesis using enzyme and protein models and reverse micelle methods

Gugalia and Sharma

[87-88], and a "dry" synthesis strategy " as an aerosol-based process [8990], varying from single-element nanoparticles, semiconductor oxides, other metal oxides, metal alloys, polymers, doped nanoparticles, and composites. Nanoparticles for use as fertilizers require a synthesis method capable of producing large-scale particles with controlled physicochemical properties at low cost [91].

Nanoparticles delivery, uptake, translocation, and biodistribution

Agrochemicals can be applied to plants by three methods: seed treatment, soil application or foliar spray. When the nanoparticles were mixed into the soil, the exposure and localization concentrations were significantly greater than those of repeated exposure during foliar spray or transfer to fruit roots, resulting in very little ingestion and contribution. In addition, high risk concentrations may affect soil microorganisms or the biosphere [92-95], as well as cause caking or clumps due to the physico-chemical characteristics of the soil which may constrain plant particles. [96-97]. A study comparing the delivery of nanoparticles to plants through foliar spray versus soil treatment showed that foliar spraying offers significant benefits for nutrient delivery at scale. nano [98]. Experiments showed that an effective aerosol spray supports the generation of monodisperse particles and avoidance of soft aggregates during foliar application. Nutrients and insecticides have been applied to the leaves for many years. Theoretically, soil treatment or fertilizer treatment is based on soil nutrient deficiencies, while foliar treatment (aerosol treatment) is based on plant nutrient efficiency characteristics. [99]. To reduce nutrient loss, foliar treatments must have a higher leaf area index, a low exposure dose, the potential for multiple applications, and an application duration that varies with the season. Poisoning could occur if people or other animals inhale an aerosol of manufactured nanoparticles. However, this conclusion is influenced by particle concentrations in the environment, daily/time weather patterns, exposure concentration and physico-chemical properties of the particles [100-102]. To ensure safe foliar application of nano-fertilizer, it is essential to use appropriate protective equipment such as masks, gloves and eye protection [103-104]. Particles in aerosols are monodisperse, dispersed and significantly more stable than particles in typical solution spray or soil applications that stack as a result of particle or particle-soil interactions [105-106].

The features of nanofertilizers are especially significant for foliar distribution because they allow shape to be limited upward along the abdominal channel [107]. It has been demonstrated that controlling nanofertilizer particle size in conjunction with the aerosol delivery strategy, as reported in the preceding [108], improves stomatal uptake. Furthermore, foliar dispersion of iron and magnesium nanofertilizer for Urd (Vigna unguiculata) was demonstrated to have a considerable favourable influence on plant growth and development [109]. The results demonstrated that nanoparticles with diameters less than 100 nm created by an aerosol technique enter the foliar part of watermelon plants through the stomach, transit through the phloem, and reach the root. It is important to note that in many situations, plant parts are studied to explore the transit of nanomaterials using the ICP-MS technique, which detects ions rather than particles. Several other research, however, have confirmed their ICP-MS observations with microscopic or X-ray spectroscopy investigations of plant components to explain the presence of actual particles. However, non-representative results when it comes to imaging of a relatively small fraction of the entire plant are a main emphasis of electron microscopy of plant tissue [110-113]. Aerosol-mediated foil coating, cuticle, permeates the nanoparticles, overcoming the primary blockage of the plant cell [114-115]. Convective systems - the phloem transport channel, a bidirectional channel along the photosynthate gradients - transfer nanoparticles from shoot to root. Both the apoplast and symplast routes transport nanoparticles within cells. The apoplast pathway prefers larger molecules (200 nm), whereas the simple pathway prefers smaller particles (50 nm).

FUTURE EXPECTATIONS

Due to the specific physico-chemical properties of nanostructures, their use as agrochemicals (fertilizers or pesticides) for plant development and crop protection is under constant investigation. Currently, investments and future study requests appear to be more focused on the development of eco-friendly nanoscale materials for successful reactions. Research on nanotechnology in agriculture is still in its infancy but is progressing rapidly. However, before nano-fertilizers can be used in the field for typical agricultural practices, a better understanding of their mechanisms of action is required in accordance with the regulatory structure that provides for the appropriate use of such agrochemicals. The United States Food and Drug Administration has previously published guidance on the use of nanometry in animal nutrition [116]. Manufacturers are also incorporating engineered nanoparticles into foods, skin care products, and other consumer products. Silica is an example. Nanoparticles in baby food, titanium dioxide nanoparticles in powdered sugar donuts and other nanomaterials in paints, plastics, paper fibres, medicines and tubes of toothpaste [117-118]. Many characteristics of nanoparticles, such as size, shape,

Gugalia and Sharma

crystalline phase, solubility, type of substance, exposure and dose concentration are considered hazardous to the health of the market and are safe to consume, but this was an area that needed more thorough investigation. These challenges require additional research on the mode of action of nanoparticles [121]. Once the human body has been exposed to nanoparticles through nano foods, researchers need to develop a life cycle assessment of the health and environmental impacts of nanomaterials, as well as methods to assess and manage any risks, and also the sustainability of the nanostructured materials used in agriculture. To come up with solutions. Some of the key issues with nanoscale innovation and its use in agrochemicals need to be addressed.

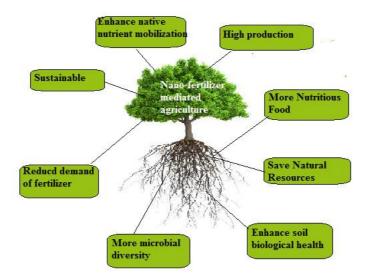


Figure1: Nanotechnology based Agriculture

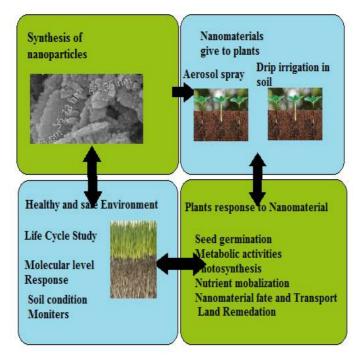


Figure 2: Methodical analysis of nanoparticles in agriculture

| Type of NP | Concentration | planning | Plant | Observation | | | |
|--|------------------------------|--|---|--|--|--|--|
| rypeon | (ppm) | method in | Tiant | observation | | | |
| | | plants | | | | | |
| ESSENTIAL PLANT NUTRIENT | | | | | | | |
| Carbon based NPs | 5-500 | Nutrient media and uptake; Seed treatment | Tomato [27,28,29]; Tobacco [30,31,32] Wheat [33]; Gram [34,35] melon [36] <i>Saltmarsh</i> <i>cordgrass</i> [37] Soybean [38,39,40] Corn, Barley, rice, [41,42] | Progressing upregulation of stress-related genes. Risen root elongation. Enhancing crop yield and seed property. Reduce heavy metal toxicity. | | | |
| Nitrogen Urea HA | 50 Kg/ha | Soil exposure | Rice [43-44] | 1. A gradual release of nitrogen 2. Enriched rice yield | | | |
| phorous CaPO4, CMC –HA, Phosphorite Zn induced P | 10-100 | Soil and foliar applications | Cotton[45]; Pearlmillets [46] Beans [47-49] Wheat, Rye, Pea, Barley, Corn, Buckwheat, Radish, Cucumber [50] | Protect toward oxidative pressure Mobilize native P and improve uptake Improves plant growth and yield | | | |
| Magnesium MgO | 15 | Foliar | Clusterbean [51] | 1.Advancing biomass, chlorophyll content & phenological growth | | | |
| Manganese | 100 -1200 | Foliar & seed treatment | Corn [52], Tomato [27,28,29] | 1.Improved seedling germination, plant biomass, and biochemical activities | | | |
| Zinc ZnO | 10 - 2000 | Foliar application Seed application | Peanut [53] Beans [54-57] Tomato [27,28,29] Cotton [58] Maize [59] | Accretion of yield potential and plant growth. Enhance plant hormone level and plant growth Help to decrease drought intensity and improve agronomic support Enhance shoot length, root length, root and shoot dry mass, leaf area and number of roots, plant biomass, root and shoot growth | | | |
| Iron Iron Oxide | 1.5-4000 | Foliar Spray | Wheat [60]; Watermelon [61,62] Clover [63]; Soybean [64] Rice [44]; Tomato [27,28,29] Peanut [53]; Corn [52] Zea mays [65] Pumpkin [66] | Enhance photosynthesis rate, chlorophyll content, biomass, grain yield & nutritional quality Improving plant growth Enhance nutrient absorption by increase microbial enzyme activity in the rhizosphere chlorophyll content, root length, leaf length, and stem length | | | |
| | NON-ESSENTIAL PLANT NUTRIENT | | | | | | |
| | | | | | | | |

Table 1 Type of nanoparticle used as in crop and effect on plant growth

| Type of NP | Concentration (ppm) | planning method in plants | Plant | Observation |
|--|------------------------|--|--|---|
| Titanium TiO2 | 200-600 | Seed, soil and foliar exposure | Spinach [67- 69] Lemna minor [70] Tomato [27,28,29] ; Wheat [71] Watermelon [61-62]; Mung Bean [72] Moth bean [73] Pearl millet [74] Clusterbean [75,76] | Raised plant biomass and photosynthetic movement. Increased biochemical enzyme activity and light absorption by chloroplast, carbon fixation Enhanced germination rate. Increased nitrogen metabolism |
| Cerium CeO | 0.1-250 | Irrigation; Seed/root | Tomato [77- 78] Arabidopsis thaliana [79- 80] Cilantro [81]; Wheat [60] | Cultivated plant growth and yield Enhanced physiological and Molecular response Increase stress threshold enzyme activity |
| Silver Ag | 1 -10 | Hydroponics, Soil | Poplars [82] Arabidopsis thaliana [79- 80] Clover | 1.Influence Phyto-stimulatory effect 2. Increase nutrient consumption by improving microbial activity in the rhizosphere. |
| Silica Si SiO SiO ₂ | 5-800 | Soil irrigation, Seed & Root exposure | Tomato [27,28,29] Wheat, Lupin [83] <i>Larix olgensis</i> [84] | Help overcome from salinity stress and Improve plant growth. Enhance germination and growth. Enrichment in total protein and chlorophyll content. Raised seedling growth and quality. |
| Cobalt Ferrite | 1-1000 | Root exposure | Tomato [27- 29] | 1. Encourage root growth |
| Indium In ₂ O ₃ | 250 | Seed/Root | Arabidopsis thaliana [79- 80] | Improved physiological and molecular response |

REFERENCES

- 1. FAO (2017). http://www.fao.org/fileadmin/templates/wsfs/docs/Issues_papers/HLEF2050_Global_Agricultu re.pdf
- 2. Godfray, H. C. J., Beddington, J. R., Crute, I. R., Haddad, L., Lawrence, D., Muir, J. F., Pretty, J.Robinson, S., Thomas, S. M. Toulmin, C. (2010). Food security: The challenge of feeding 9 billion people. Science, *327*:812-818.
- 3. Nations, (2017) U. http://www.un.org/en/development/desa/news/population/2015-report.html
- 4. Initiative, N. N. http://www.nano.gov/ (March 1, 2017),
- 5. USDA-(2017).NIFA https://nifa.usda.gov/program/nanotechnology-program
- 6. Tarafdar, J., Sharma, S., Raliya, R.(2013). Nanotechnology: Interdisciplinary science of applications.. African J. Biotechnol. *12*.(3):219-226
- 7. Raliya, R., Tarafdar, J., Gulecha, K., Choudhary, K., Ram, R., Mal, P., Saran, R. (2013). Review article; scope of nanoscience and nanotechnology in agriculture. J. Appl. Biol. Biotechnol., *1*: 041-044.
- 8. Gogos, A.,Knauer, K., Bucheli, T. D. (2012). Nanomaterials in plant protection and fertilization: Current state, foreseen applications, and research priorities. J. Agric. Food Chem., *60*:9781-9792.
- 9. Chhowalla, M. (2017). Slow release nanofertilizers for bumper crops. ACS Central Science,, 3, 156.
- 10. Dimkpa, C. O. and Bindraban, P. S. (2018). Nanofertilizers: New products for the industry? J. Agric. Food Chem., 66 (26): 6462-6473
- 11. Nair, R., Varghese, S. H., Nair, B. G., Maekawa, T.; Yoshida, Y.; Kumar, D. S. (2010). Nanoparticulate material delivery to plants. Plant Sci., *179*:154-163.
- 12. Saharan, V., Khatik, R., Kumari, M., Raliya, R., Nallamuthu, I., Pal, A. (2014). International Conference on Advances in Biotechnology (BioTech). In: Proceedings of Global Science and Technology Forum : p 23.

- 13. Tilman, D.,Cassman, K. G.,Matson, P. A., Naylor, R., Polasky, S. (2002). Agricultural sustainability and intensive production practices. Nature, *, 418:*671-677.
- 14. Carpenter, S. R., Caraco, N. F., Correll, D. L., Howarth, R. W., Sharpley, A. N., Smith, V. H. (1998). Nonpoint pollution of surface waters with phosphorus and nitrogen. *Ecol. Appl.*, *8*:559-568.
- 15. Tilman, D. Cassman, K. G. Matson, P. A.Naylor, R. Polasky, S.(**2002**). Agricultural sustainability and intensive production practices. Nature, *418*:671-677.
- 16. Carpenter, S. R., Caraco, N. F., Correll, D. L., Howarth, R. W., Sharpley, A. N., Smith, V. H.(1998) Nonpoint pollution of surface waters with phosphorus and nitrogen. Ecol. Appl., *8*, 559-568.
- 17. MIT-USA, 2017 http://web.mit.edu/12.000/www/m2016/finalwebsite/solutions/phosphorus.html
- 18. Dawson, C. J., Hilton, J.(2011). Fertiliser availability in a resource-limited world: Production and recycling of nitrogen and phosphorus. Food Policy, *36*:S14-S22.
- 19. USGIS http://www.usgs.gov/climate_landuse/. (March 1, 2017),
- 20. Greene, R. P., Harlin, J. M. (**1995**). Threat to high market value agricultural lands from urban encroachment: A national and regional perspecitve. *Social Sci. J.*, *32*: 137-155.
- 21. Pimentel, D.; Wilson, A. (2004). World population agriculture and malnutrition. World Watch, 22-25.
- 22. Saharan, V. Kumaraswamy, R., Choudhary, R. C., Kumari, S., Pal, A., Raliya, R., Biswas, P. (2016). Cu- chitosan nanoparticle mediated sustainable approach to enhance seedling growth in maize by mobilizing reserved food. J. Agric. Food Chemi., *64*, 6148-6155.
- 23. Zahra, Z.; Arshad, M.; Rafique, R.; Mahmood, A.; Habib, A.; Qazi, I. A.; Khan, S. A. (2015). Metallic nanoparticle (tio₂ and fe₃o₄) application modifies rhizosphere phosphorus availability and uptake by *lactuca sativa*. J. Agric. Food Chemi., *63*:6876-6882.
- 24. Suttiponparnit, K. Jiang, J., Sahu, M., Suvachittanont, S, Charinpanitkul, T., Biswas, P. (2010). Role of surface area, primary particle size, and crystal phase on titanium dioxide nanoparticle dispersion properties. *Nanoscale Res. Lett.*: 6, 1
- 25. Biswas, P., Wu, C.-Y. (2005) .Nanoparticles and the environment. J. Air Waste Manag. Assoc., 55, 708-746.
- 26. Wiesner, M. R., Lowry, G. V., Alvarez, P., Dionysiou, D., Biswas, P. (2006). Assessing the risks of manufactured nanomaterials. Environ. Sci. Technol., *40*, 4336-4345.
- 27. Lahiani, M. H., Chen, J., Irin, F., Puretzky, A. A., Green, M. J., Khodakovskaya, M. V.(2015). Interaction of carbon nanohorns with plants: Uptake and biological effects. Carbon, *81*, 607-619.
- 28. Khodakovskaya, M. V., Kim, B. S.,Kim, J. N.,Alimohammadi, M., Dervishi, E.; Mustafa, T., Cernigla, C. E. (2013). Carbon nanotubes as plant growth regulators: Effects on tomato growth, reproductive system, and soil microbial community. Small, *9*:115-123.
- 29. Lahiani, M. H., Dervishi, E., Ivanov, I., Chen, J., Khodakovskaya, M.(2016). Comparative study of plant responses to carbon-based nanomaterials with different morphologies. Nanotechnol., *27*, 265102.
- 30. Khodakovskaya, M. V.; de Silva, K.; Biris, A. S.; Dervishi, E.; Villagarcia, H.(2012). Carbon nanotubes induce growth enhancement of tobacco cells. *ACS Nano*, *6*:2128-2135.
- 31. Villagarcia, H., Dervishi, E., de Silva, K., Biris, A. S., Khodakovskaya, M. V. (2012). Surface chemistry of carbon nanotubes impacts the growth and expression of water channel protein in tomato plants. Small, *8*:2328-2334.
- Gao J., Wang Y., Folta K.M., Krishna V., Bai W., Indeglia P., Georgieva A., Nakamura H., Koopman B., Moudgil B. (2011). Polyhydroxy fullerenes (fullerols or fullerenols): Beneficial effects on growth and lifespan in diverse biological models. PLoS ONE. 6:19976.
- 33. Wang, X.; Han, H.; Liu, X.; Gu, X.; Chen, K.; Lu, D. (**2012**). Multi-walled carbon nanotubes can enhance root elongation of wheat (*triticum aestivum*) plants. *J.* Nanopart. Res., *14*, 841.
- 34. Tripathi, S.; Sonkar, S. K.; Sarkar, S.(2011). Growth stimulation of gram (*Cicer arietinum*) plant by water soluble carbon nanotubes. Nanoscale, *3*:1176-1181.
- 35. Sonkar, S. K., Roy, M., Babar, D. G.; Sarkar, S. (2012). Water soluble carbon nano-onions from wood wool as growth promoters for gram plants. Nanoscale, *4*:7670-7675.
- 36. Kole, C.,Kole, P., Randunu, K. M., Choudhary, P., Podila, R.,Ke, P. C., Rao, A. M., Marcus, R. K.(2013). Nanobiotechnology can boost crop production and quality: First evidence from increased plant biomass, fruit yield and phytomedicine content in bitter melon (*Momordica charantia*). *BMC Biotechnol.*, *13*: 37.
- 37. Wand, X., Huang, Q., Wang, L., Wang, L. (2012. Effect of single-wall carbon nanotube on soybean (*glycine max*) regeneration from mature cotyledonary node explants. Nano Life, *2:* 1250014.
- 38. Lahiani, M. H., Dervishi, E., Chen, J., Nima, Z.,Gaume, A.; Biris, A. S.; Khodakovskaya, M. V. (2013) Impact of carbon nanotube exposure to seeds of valuable crops. *ACS* Appl. Mater. Inter., *5:* 7965-7973.
- 39. Kottegoda, N., Sandaruwan, C., Priyadarshana, G., Siriwardhana, A., Rathnayake, U., Berugoda Arachchige, D., Kumarasinghe, A., Dahanayake, D., Karunaratne, V., Amaratunga, G. (2017) Urea- hydroxyapatite nanohybrids for slow release of nitrogen. *ACS* Nano, *11*: 1214-1221.
- 40. Liu, R.; Lal, R. (2014). Synthetic apatite nanoparticles as a phosphorus fertilizer for soybean (glycine max). Sci. Rep., *4*: 5686.
- 41. Sharonova, N. L., Yapparov, A. K., Khisamutdinov, N. S., Ezhkova, A. M., Yapparov, I. A., Ezhkov, V. O., Degtyareva, I. A., Babynin, E. V.(2015). Nanostructured water-phosphorite suspension is a new promising fertilizer. Nanotechnol. Russia, *10*:651-661.
- 42. Raliya, R.; Tarafdar, J. C.(2017). Zno nanoparticle biosynthesis and its effect on phosphorous-mobilizing enzyme secretion and gum contents in clusterbean (*Cyamopsis tetragonoloba* l.). Agric. Res., 736, *2:* 48-57.

- 43. Raliya, R., Tarafdar, J. C., Biswas, P.(2016). Enhancing the mobilization of native phosphorus in the mung bean rhizosphere using zno nanoparticles synthesized by soil fungi. *J.* Agric. Food Chemi, *64*: 3111-3118.
- 44. Tarafdar, J., Raliya, R., Mahawar, H., Rathore, I. (2014) .Development of zinc nanofertilizer to enhance crop production in pearl millet (*Pennisetum americanum*). Agric. Res., **14**, 3:257-262.
- 45. Yousefzadeh S. and Sabaghnia N.(2016). Nano-iron fertilizer effects on some plant traits of dragonhead (*Dracocephalum moldavica*) under different sowing densities. *Acta Agric. Slov.*,107:429–437.
- 46. Raliya, R. Tarafdar, J. Singh, S., Gautam, R., Choudhary, K., Maurino, V. G., Saharan, V.(2014). MgO nanoparticles biosynthesis and its effect on chlorophyll contents in the leaves of cluster bean (*Cyamopsis tetragonoloba*.). *Adv. Sci. Eng. Med.*, *6*: 538-545.
- 47. Pradhan, S., Patra, P., Mitra, S., Dey, K. K., Jain, S., Sarkar, S., Roy, S., Palit, P., Goswami, A.(2014). Manganese nanoparticles: Impact on non-nodulated plant as a potent enhancer in nitrogen metabolism and toxicity study both in vivo and in vitro. J. Agric. Food Chemi., 62: 8777-8785.
- 48. Saharan, V., Sharma, G., Yadav, M., Choudhary, M. K., Sharma, S., Pal, A., Raliya, R., Biswas, P.()2015). Synthesis and in vitro antifungal efficacy of cu–chitosan nanoparticles against pathogenic fungi of tomato. Int. J. Biol. Macromol., 75:346-353.
- 49. Prasad, T., Sudhakar, P., Sreenivasulu, Y., Latha, P., Munaswamy, V., Reddy, K. R., Sreeprasad, T., Sajanlal, P., Pradeep, T. (2012). Effect of nanoscale zinc oxide particles on the germination, growth andyield of peanut. J. Plant Nutrition, 35: 905-927.
- 50. Sheykhbaglou, R., Sedghi, M., Shishevan, M. T., Sharifi, R. S.(2010) .Effects of nano-iron oxide particles on agronomic traits of soybean. Notulae Scientia Biologicae, 2010, 2:112.
- 51. Giordani, T., Fabrizi, A, Guidi, L., Natali, L., Giunti, G., Ravasi, F., Cavallini, A., Pardossi, A.(2012). Response of tomato plants exposed to treatment with nanoparticles. EQA-International J. Environ. Quality, 8: 27-38.
- 52. Laurent, S., Forge, D., Port, M., Roch, A., Robic, C., Vander Elst, L., Muller, R. N.(2008). Magnetic iron oxide nanoparticles: Synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications. Chemi. Rev., 108, 2064-2110.
- 53. Li, J.,Hu, J., Ma, C., Wang, Y., Wu, C., Huang, J., Xing, B.(2016). Uptake, translocation and physiological effects of magnetic iron oxide (γ-Fe₂O₃) nanoparticles in corn (*Zea mays*.). Chemosphere, 159, 326-334.
- 54. Zhu, H., Han, J., Xiao, J. Q., Jin, Y. (2008). Uptake, translocation, and accumulation of manufactured iron oxide nanoparticles by pumpkin plants. *J. Environ. Monit.*, 10: 713-717.
- 55. Li, J.; Chang, P. R.; Huang, J.; Wang, Y.; Yuan, H.; Ren, H. Physiological effects of magnetic iron oxide nanoparticles towards watermelon. *J. Nanosci. Nanotechnol.* 2013, 13, 5561-5567.
- 56. Ghafari, H.; Razmjoo, J.(2013). Effect of foliar application of nano-iron oxidase, iron chelate and iron sulphate rates on yield and quality of wheat. *Int. J. Agronom. Plant Product*, 4:2997-3003.
- 57. Feng, Y.,Cui, X., He, S., Dong, G.,Chen, M.,Wang, J., Lin, X. (2013). The role of metal nanoparticles in influencing arbuscular mycorrhizal fungi effects on plant growth. Environ. Sci. Technol., 47: 9496-9504.
- 58. Ghafariyan, M. H., Malakouti, M. J., Dadpour, M. R., Stroeve, P., Mahmoudi,(2013). Effects of magnetite nanoparticles on soybean chlorophyll. Environ. Sci. Technol., 47:10645-10652.
- 59. Alidoust, D. and Isoda, A.(2013). Effect of γ Fe₂O₃ nanoparticles on photosynthetic characteristic of soybean (Glycine max (L.) merr.): Foliar spray versus soil amendment. Acta Physiologiae Plantarum, 35: 3365-3375.
- 60. Alidoust, D. and Isoda, A.(2014). Phytotoxicity assessment of γ-Fe₂O₃ nanoparticles on root elongation and growth of rice plant. Environ. Earth Sci., 71, 5173-5182.
- 61. Rui, M., Ma, C., Hao, Y., Guo, J., Rui, Y., Tang, X., Zhao, Q., Fan, X., Zhang, Z., Hou, T. (2016). Iron oxide nanoparticles as a potential iron fertilizer for peanut (*Arachis hypogaea*). Front. Plant Sci., 7: 815.
- 62. Tarafdar, A., Raliya, R., Wang, W.-N., Biswas, P., Tarafdar, J.(2013). Green synthesis of tio2 nanoparticle using Aspergillus tubingensis. Adv. Sci., Eng. Med., 5: 943-949.
- 63. Raliya, R. (2012). Appliance of nanoparticles on plant system and associated rhizospheric microflora. Ph.D Thesis, J. N. Vyas University Jodhpur, p:199.
- 64. Yang, F., Hong, F., You, W., Liu, C., Gao, F., Wu, C., Yang, P.(2006). Influence of nano-anatase tio2 on the nitrogen metabolism of growing spinach. Biol. Trace Element Res., 110: 179-190.
- 65. Mingyu, S., Xiao, W., Chao, L., Chunxiang, Q., Xiaoqing, L., Liang, C., Hao, H., Fashui, H. (2007). Promotion of energy transfer and oxygen evolution in spinach photosystem ii by nano-anatase TiO₂. *B*iol. Trace Element Res., 119:183-192.
- 66. Gao, F., Hong, F.,Liu, C.; Zheng, L.,Su, M., Wu, X.,Yang, F.,Wu, C.Yang, P. (2012). Mechanism of nano anatase TiO₂ on promoting photosynthetic carbon reaction of spinach. Biol. Trace Element Res., 111: 239-253.
- 67. Hong, F., Yang, F., Liu, C., Gao, Q., Wan, Z., Gu, F., Wu, C., Ma, Z., Zhou, J., Yang, P.(2005). Influences of nano-TiO₂ on the chloroplast aging of spinach under light. *Biol. Trace Element Res.* 104:249-260.
- 68. Hong, F., Zhou, J., Liu, C., Yang, F., Wu, C., Zheng, L., Yang, P. (2005). Effect of nano-TiO₂ on photochemical reaction of chloroplasts of spinach. Biol. Trace Element Res., 105: 269-279.
- 69. Linglan, M., Chao, L., Chunxiang, Q., Sitao, Y., Jie, L., Fengqing, G., Fashui, H.(2008) , Rubisco activase m -RNA expression in spinach: Modulation by nanoanatase treatment. *Biol. Trace Element Res.*, 168-178.
- 70. Feizi H., Moghaddam, P. R., Shahtahmassebi, N., Fotovat, A. (2012).Impact of bulk and nanosized titanium dioxide (TiO₂) on wheat seed germination and seedling growth. *B*iol. Trace Element Res., 146: 101-106.
- 71. Liang, Y.; Nikolic, M.; Bélanger, R.; Gong, H.; Song, (2015). Silicon in Agriculture: From Theory to Practice A. Springer: 123-142.

- 72. Chen, X. and Mao, S. S.(2007). Titanium dioxide nanomaterials: Synthesis, properties, modifications, and applications. Chem. Rev., 107: 2891-2959.
- 73. Raliya, R.,Biswas, P.,Tarafdar, J. (2015).TiO₂ nanoparticle biosynthesis and its physiological effect on mung bean (Vigna radiata L.). Biotechnol. Rep., 5: 22-26.
- 74. Ma, C., Liu, H., Guo, H., Musante, C., Coskun, S. H., Nelson, B. C., White, J. C., Xing, B., Dhankher, O. P.(2016). Defense mechanisms and nutrient displacement in *arabidopsis thaliana* upon exposure to CeO₂ and In₂O₃ nanoparticles. Environ. Sci.: Nano, 3: 1369-1379.
- 75. Wang, Q., Ma, X., Zhang, W., Pei, H., Chen, Y. (2012). The impact of cerium oxide nanoparticles on tomato (Solanum lycopersicum L.) and its implications for food safety. Metallomics, 4:1105-1112.
- 76. Wang, Q. Ebbs, S. D., Chen, Y.; Ma, X.(2013). Trans-generational impact of cerium oxide nanoparticles on tomato plants. Metallomics, *,* 5: 753-759.
- 77. Morales, M. I.,Rico, C. M., Hernandez-Viezcas, J. A., Nunez, J. E., Barrios, A. C., Tafoya, A., Flores-Marges, J. P., Peralta-Videa, J. R., Gardea-Torresdey, J. L. (2013). Toxicity assessment of cerium oxide nanoparticles in cilantro (Coriandrum sativum L.) plants grown in organic soil *J.* Agric. Food Chemi., 61: 6224-6230.
- Rico, C. M., Lee, S. C., Rubenecia, R., Mukherjee, A., Hong, J., Peralta-Videa, J. R.;Gardea-Torresdey, J. L.(2014). Cerium oxide nanoparticles impact yield and modify nutritional parameters in wheat (*Triticum aestivum* L.). J. Agric. Food Chemi. 62: 9669-9675.
- 79. Wang, J., Koo, Y., Alexander, A., Yang, Y., Westerhof, S., Zhang, Q., Schnoor, J. L., Colvin, V. L.;Braam, J., Alvarez, P. J(2013). Phytostimulation of poplars and arabidopsis exposed to silver nanoparticles and ag⁺ at sublethal concentrations. Environ. Sci. Technol., 47: 5442-5449.
- Ma, C., Chhikara, S., Xing, B., Musante, C., White, J. C., Dhankher, O. P.(2013). Physiological and molecular response of *Arabidopsis thaliana* (L.) to nanoparticle cerium and indium oxide exposure. *ACS Sustain. Chemi. Eng.*, 1: 768-778.
- 81. Siddiqui, M. H and Al-Whaibi, M. H.(2014). Role of nano-sio2 in germination of tomato *(Lycopersicum esculentum seeds mill.)*. Saudi J. Biol. Sci., 21:13-17.
- 82. Sun, D., Hussain, H. I., Yi, Z, Rookes, J. E., Kong, L., Cahill, D. M.(2016). Mesoporous silica nanoparticles enhance seedling growth and photosynthesis in wheat and lupin. Chemosphere, 152: 81-91.
- López-Moreno, M. L., Avilés, L. L., Pérez, N. G., Irizarry, B. Á., Perales, O., Cedeno-Mattei, Y., Román, F.(2016). Effect of cobalt ferrite (CoFe₂O₄) nanoparticles on the growth and development of Lycopersicon lycopersicum (tomato plants). Sci. Total Environ. 550: 45-52.
- 84. Raliya, R., Singh Chadha, T., Haddad, K., Biswas, P.(2016). Perspective on nanoparticle technology for biomedical use. Curr. Pharmaceut. Des., 22: 2481-2490.
- 85. Ruffini Castiglione, M., Giorgetti, L., Geri, C.; Cremonini, R.(2011). The effects of nano-TiO2 on seed germination, development and mitosis of root tip cells of *Vicia narbonensis* L. And *Zea* mays L. J. Nanopart. Res., 13: 2443-2449.
- 86. Boonyanitipong, P., Kositsup, B., Kumar, P., Baruah, S., Dutta, J. (2011). Toxicity of ZnO and Tio₂ nanoparticles on germinating rice seed *Oryza sativa* L. Int. J. Biosci., Biochem. Bioinfo., 2011, 1, 282.
- 87. Kaul, R., Kumar, P., Burman, U., Joshi, P., Agrawal, A., Raliya, R., Tarafdar, J. (2012) . Magnesium and iron nanoparticles production using microorganisms and various salts. Mater. Sci.-Poland, 2012, 30, 254-258.
- 88. Raliya, R., Rathore, I., Tarafdar, J. (2013). Development of microbial nanofactory for zinc, magnesium, and titanium nanoparticles production using soil fungi. J. Bionanosci., 7: 590-596.
- 89. Raliya, R., and Tarafdar, J.(2013). Biosynthesis of gold nanoparticles using *Rhizoctonia bataticola* TFR-6. Adv.Sci., Eng. Med., 5: 1073-1076.
- 90. Li, S., Ren, Y., Biswas, P., Stephen, D. T.(2016). Flame aerosol synthesis of nanostructured materials and functional devices: Processing, modeling, and diagnostics. *Prog. Energy Combust. Sci.*, 55: 51-59.
- 91. Jiang, J., Chen, D.-R., Biswas, P.(2007). Synthesis of nanoparticles in a flame aerosol reactor with independent and strict control of their size, crystal phase and morphology. Nanotechnol, 18:285603.
- 92. Gajjar, P., Pettee, B., Britt, D. W., Huang, W., Johnson, W. P., Anderson, A. (2009). Antimicrobial activities of commercial nanoparticles against an environmental soil microbe, *Pseudomonas putida kt 2440*. J. Biologic. Eng., 3:9.
- 93. Collins, D., Luxton, T., Kumar, N., Shah, S., Walker, V. K., Shah, V.(2012). Assessing the impact of copper and zinc oxide nanoparticles on soil: A field study. PLoS One, *7*:42663.
- 94. Mehta, C., Srivastava, R., Arora, S., Sharma, A. Impact assessment of silver nanoparticles on plant growth and soil bacterial diversity. Biotech, *6*: 254.
- 95. Cao, J., Feng, Y., Lin, X., Wang, J(2016). Arbuscular mycorrhizal fungi alleviate the negative effects of iron oxide nanoparticles on bacterial community in rhizospheric soils. Frontiers in Env. Sci., 4:10.
- 96. Majumdar, S., Peralta-Videa, J. R., Trujillo-Reyes, J., Sun, Y.; Barrios, A. C., Niu, G.; Flores-Margez, J. P., Gardea-Torresdey, J.L.(2016). Soil organic matter influences cerium translocation and physiological processes in kidney bean plants exposed to cerium oxide nanoparticles. Sci. Total Environ., *569*:201-211.
- 97. Anderson, A., McLean, J., McManus, P., Britt, D.(2017). Soil chemistry influences the phytotoxicity of metal oxide nanoparticles. Int. J. Nanotechnol., *14*:15-21.
- 98. Fernández, V., Sotiropoulos, T., Brown, P. H(2013). Foliar fertilization: Scientific principles and field practices. Int. Fert. Ind. Assoc. 2013: 140.
- 99. Fageria, N. K., Filho, M. P. B., Moreira, A., Guimarães, C. M (2009). Foliar fertilization of crop plants. J.Plant Nutr.,

2009, 32, 1044-1064.

- 100. Fernandez, V. and Brown, P. H.(2013). From plant surface to plant metabolism: The uncertain fate of foliarapplied nutrients. Front. Plant Sci., 4:289.
- 101. Nel, A., Xia, T., Mädler, L., Li, N. (2006). Toxic potential of materials at the nanolevel. Science, 311: 622-627.
- 102. Maynard, A. D., and Kuempel, E. D. (2005). Airborne nanostructured particles and occupational health. J.Nanopart. Res., 7: 587-614.
- 103. Colvin, V. L.(2003). The potential environmental impact of engineered nanomaterials. Nat. Biotechnol., 21: 1166-1170.
- 104. Jiang, J., Oberdörster, G., Elder, A., Gelein, R., Mercer, P., Biswas, P.(2008). Does nanoparticle activity depend upon size and crystal phase? Nanotoxicol, 2: 33-42.
- 105. Maynard, A. D., Aitken, R. J., Butz, T., Colvin, V., Donaldson, K., Oberdörster, G., Philbert, M. A., Ryan, J., Seaton, A., Stone, V.(2006). Safe handling of nanotechnology. Nature, 444:267.
- 106. Huang, S., Wang, L., Liu, L., Hou, Y., Li, L. (2015). Nanotechnology in agriculture, livestock, and aquaculture in china. A review. Agronom. Sustain. Develop, 35: 369-400.
- 107. Eichert, T. and Goldbach, H. E. (2008). Equivalent pore radii of hydrophilic foliar uptake routes in stomatous and astomatous leaf surfaces–further evidence for a stomatal pathway. Physiologia Plantarum, 132: 491-502.
- 108. Wang, W.N., Tarafdar, J. C, Biswas, P. (2013). Nanoparticle synthesis and delivery by an aerosol route for watermelon plant foliar uptake. J. Nanopart. Res., *15*:1-13.
- 109. Raliya , R., Sanmathi Chavalmane,, C. F., Nair, R., Reed, R., Biswas, P.(2016). Quantitative understanding of nanoparticle uptake in watermelon plants. *Front. Plant Sci.*, 7:1288.
- 110. Huang, S., Wang, L., Liu, L., Hou, Y., Li, L. (2015). Nanotechnology in agriculture livestock and aquaculture in china. A review. Agronom. Sustain. Develop, 35: 369-400.
- 111. Eichert, T., and Goldbach, H. E. (2008). Equivalent pore radii of hydrophilic foliar uptake routes in stomatous and astomatous leaf surfaces–further evidence for a stomatal pathway. Physiologia Plantarum, 132: 491-502.
- 112. Delfani, M, Baradarn Firouzabadi, M, Farrokhi, N., Makarian, H.(2014) .Some physiological responses of blackeyed pea to iron and magnesium nanofertilizers. Comm. Soil Sci. Plant Anal., 45: 530-540.
- 113. Dan, Y,Zhang, W, Xue, R.,Ma, X., Stephan, C Shi, H.(2015). Characterization of gold nanoparticle uptake by tomato plants using enzymatic extraction followed by single-particle inductively coupled plasma-mass spectrometry analysis. *Environ. Sci. Technol.*, 49: 3007-3014.
- 114. Schwab, F, Zhai, G., Kern, M., Turner, A., Schnoor, J. L., Wiesner, M. R. Barriers, pathways and processes for uptake, translocation and accumulation of nanomaterials in plants–critical review. Nanotoxicol, 10:257-278.
- 115. Biju, V.(2014). Chemical modifications and bioconjugate reactions of nanomaterials for sensing, imaging, drug delivery and therapy. Chemi. Soc. Rev., 43:744-764.
- 116. Torney, F,Trewyn, B. G., Lin, V. S.-Y., Wang, K. (2007). Mesoporous silica nanoparticles deliver DNA and chemicals into plants. Nat. Nanotechnol., 2:295-300.
- 117. Mastronardi, E, Tsae, P. K., Zhang, X., Pach, A, Sultan, Y, DeRosa, M. C. (2016). Preparation and characterization of aptamer–polyelectrolyte films and microcapsules for biosensing and delivery applications. Methods, 97:75-87.
- 118. Monreal, C, DeRosa, M., Mallubhotla, S., Bindraban, P., Dimkpa, C (2016). Nanotechnologies for increasing the crop use efficiency of fertilizer-micronutrients. Biol. Fert. Soils, 2016, 52:423-437
- 119. Wang, P., Lombi, E., Zhao, F.-J., Kopittke, P. M. (2016). Nanotechnology: A new opportunity in plant sciences. Trends Plant Sci., 21:699-712.
- 120. Bowling, D. J. F. (1976) . Uptake of ions by plant root. Chapman and Hall, New York.
- 121. Venkatachalam, P., Priyanka, N., Manikandan, K, Ganeshbabu, I, Indiraarulselvi, P., Geetha, N., Muralikrishna, K., Bhattacharya, R. C., Tiwari, M., Sharma, N., Sahi, S. V. (2017). Enhanced plant growth promoting role of phycomolecules coated zinc oxide nanoparticles with supplementation in cotton (*Gossypium hirsutum L.*). *Plant Physiol. Biochem.*, *110*:118-127.

Copyright: © **2023 Author**. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.