

ORIGINAL ARTICLE

Design, Synthesis, Molecular Docking and Antitubercular Evaluation of 2-Substituted 1-Methyl-1h-Benzimidazoles bearing an N-(2-(4-Hydroxy-3-Carboxyphenyl)Ethyl) Moiety

Pratiksha Narayan Sonwane* and Manoj Ramesh Kumbhare

Department of Pharmaceutical Chemistry, S.M.B.T College of Pharmacy, Savitribai Phule Pune University, Dhamangaon, Igatpuri, Nashik 422403, India.

*Corresponding author's email: sonwane.pratiksha@gmail.com

ABSTRACT

In the present study, a series of 2-substituted 1-methyl-1H-benzimidazoles bearing an N-(2-(4-hydroxy-3-carboxyphenyl)ethyl) moiety was designed and synthesized to explore their potential as antitubercular agents. The synthesized compounds (P1–P6) were structurally confirmed through IR, ¹H NMR, and mass spectrometry. Their antimycobacterial activity was assessed using the Microplate Alamar Blue Assay (MABA), while molecular docking studies were performed with AutoDock Vina 1.5.6 against the 2Q1Y receptor. Among the series, compound P4 exhibited the most favorable docking score (–8.9), outperforming standard antitubercular drugs including isoniazid and para-aminosalicylic acid. Moreover, P4, P5, and P6 displayed strong binding affinity consistent with their in vitro activity, emphasizing the structural relevance of substitutions at the benzimidazole core. Collectively, these findings suggest that the synthesized benzimidazole derivatives, particularly P4, hold promise as lead candidates for the development of novel antitubercular therapeutics.

Keywords: Benzimidazole derivatives; Anti-tubercular activity; Molecular docking; Microplate Alamar Blue Assay; 2Q1Y receptor

Received 29.09.2025

Revised 02.10.2025

Accepted 12.11.2025

How to cite this article:

Pratiksha Narayan S and Manoj Ramesh K. Design, Synthesis, Molecular Docking and Antitubercular Evaluation of 2-Substituted 1-Methyl-1h-Benzimidazoles bearing an N-(2-(4-Hydroxy-3-Carboxyphenyl)Ethyl) Moiety. Adv. Biores., Vol 16 (6) November 2025: 06-16.

INTRODUCTION

Tuberculosis (TB), caused by *Mycobacterium tuberculosis* (Mtb), is a major infectious disease transmitted via respiratory droplets [1,2]. Once termed the “white plague,” TB was identified by Robert Koch in 1882, a discovery that earned him the Nobel Prize [3–5]. Despite medical progress, TB has resurged, particularly in immunocompromised individuals such as AIDS patients [6–11].

Globally, TB remains a leading cause of mortality, with over 3 million deaths each year. Nearly one-third of the world's population carries latent infection, with ~8.8 million new cases and ~7,000 deaths daily [12–15]. Drug resistance further intensifies the crisis, with over 0.5 million multidrug-resistant TB (MDR-TB) cases reported among the 9 million new infections in 2012 alone [16]. The disease burden is highest in Asia and Africa (>80% prevalence) compared to <10% in the U.S. [17]. India, China, Indonesia, South Africa, and Nigeria account for the majority of global cases, with India alone reporting ~0.23 million MDR-TB cases annually [18–20].

As shown in Figure 1, TB incidence is disproportionately concentrated in regions such as South-East Asia and sub-Saharan Africa, with countries like India, China, South Africa, and Nigeria representing significant hotspots of disease transmission.

Region	Estimated Incidence (per 100,000)	TB-Related Deaths	MDR-TB Cases (%)	HIV-TB Coinfection (%)
Africa	219	230,000	3.1%	30%
Southeast Asia	226	435,000	2.5%	10%
Western Pacific	96	150,000	1.8%	5%
Americas	28	25,000	1.2%	10%
Global Average	133	1.3 million	3.0%	8%

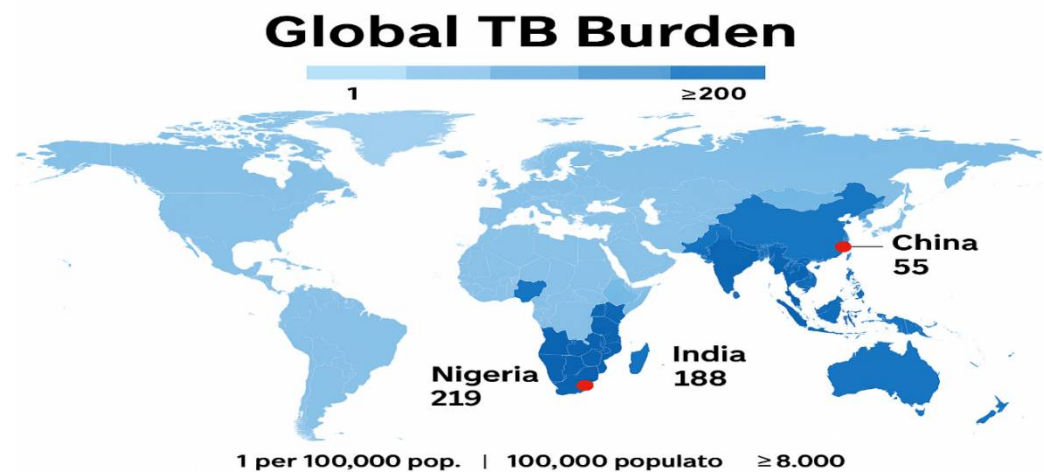


Figure 1: Global Burden of Tuberculosis
Figure 1 shows Global TB incidence rates by country (darker shades indicate higher incidence), highlighting high-burden nations such as India, China, South Africa, and Nigeria.

The urgent need for new chemotherapeutics has directed attention to heterocyclic scaffolds, with benzimidazole emerging as a key framework due to its broad pharmacological spectrum [25]. First synthesized by Debus in 1858 [26] via condensation of *o*-phenylenediamine with mono-/dibasic acids [27], benzimidazole exhibits diverse activities including antitubercular, anticancer, antifungal, antihypertensive, antimicrobial, antiviral, and anti-inflammatory effects [28–39] (Figure 2). Its NH group further imparts unique acidic–basic properties [40].

**Pharmacological Spectrum
BENZIMIDAZOLE**

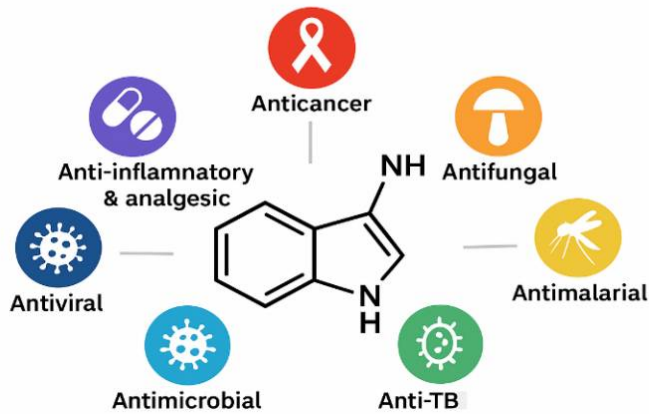


Figure 2: Pharmacological spectrum of benzimidazole derivatives

The infographic illustrates the broad range of biological activities associated with benzimidazole scaffolds, including antitubercular, anticancer, antifungal, antimicrobial, antimalarial, antiviral, antiparasitic, and anti-inflammatory/analgesic effects [41].

Structurally, benzimidazole (IUPAC: 1H-benzimidazole; synonyms: 1H-1,3-benzimidazole, 1H-benzo[d]imidazole) exhibits isosterism with indole and purine [42, 43] (Figure 3). This structural mimicry explains its significant role in biology, highlighted by the discovery of 5,6-dimethyl-1-(D-ribofuranosyl) benzimidazole as a key fragment of Vitamin B12 [82,83]. Furthermore, tautomerization at the N1 position renders the C5 and C6 carbons chemically equivalent, influencing substitution patterns and bioactivity [44].

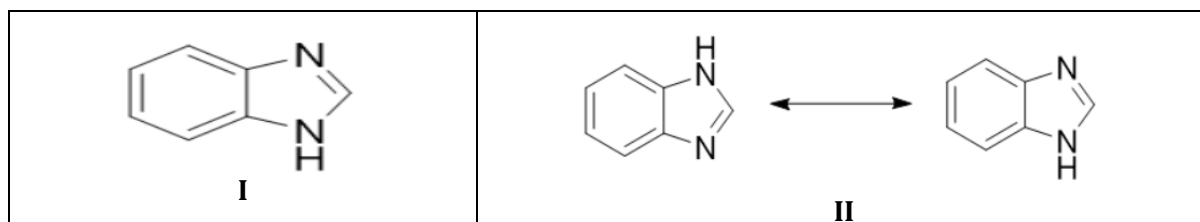
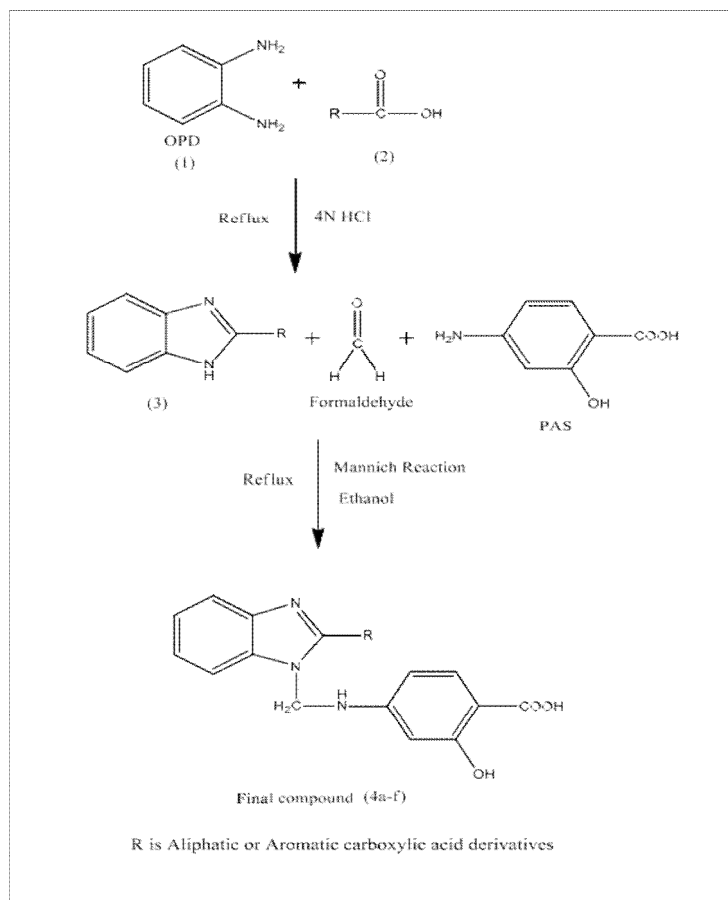


Figure 3: Structural representation.

Given these features, benzimidazole derivatives stand as promising scaffolds in anti-TB drug discovery, offering opportunities to combat drug resistance and improve therapeutic outcomes.

MATERIAL AND METHODS

All chemicals and solvents were of analytical grade (Sigma Aldrich, SD Fine Chemicals, Thomas Baker). Reactions were monitored by TLC on silica gel 60 F254 plates (Merck). Melting points were determined in Thiel's tube. IR spectra were recorded on a Shimadzu FTIR-1000 (KBr, 4000–400 cm^{-1}), ^1H NMR on a Bruker 400 MHz (DMSO- d_6 , TMS), and mass spectra by LC-MS.



Scheme-I: Synthetic route for preparation of benzimidazole derivatives

Table 2: Various Substitutions on Final compounds

Comp. Code	R/Ar
P1	-H
P2	-CH ₃
P3	-2-NH ₂ C ₆ H ₅
P4	-2,4-NO ₂ C ₆ H ₅
P5	-C ₂ H ₄ Cl
P6	-4-NO ₂ C ₆ H ₅

General procedure for the synthesis of Intermediates

o-Phenylenediamine and substituted carboxylic acids (0.01 mol) were refluxed in 4 N HCl at 100 °C (6–16 h). After TLC monitoring, the mixture was cooled, neutralized, and the precipitate recrystallized from ethanol to yield pure benzimidazole derivatives. [45]

General procedure for the synthesis of Final compounds (P1-P6)

Benzimidazole derivatives, para-aminosalicylic acid, and formaldehyde (0.02 mol each) were dissolved in ethanol (70 mL) and subjected to reflux (4 h). After solvent removal and trituration with petroleum ether (40–60 °C), PAS-conjugated benzimidazole derivatives were obtained in good yield.

Analytical data of synthesized benzimidazole derivatives**Table 3: Analytical data of synthesized benzimidazole derivatives**

Compound code	Chemical Name	M.P (°C)	% Yield	IR (cm ⁻¹)	¹ H NMR (δ, ppm)	Mass (M/Z)
P1	4-(((1H-benzo[d]imidazol-1-yl)methyl)amino)-2-hydroxybenzoic acid	145–150	85.25	N-H: 3502 OH: 3336 COOH: 3378 C=N: 1619 C=C: 1419	CH ₂ : 5.17 (2H, s) Ar-H: 6.37–7.96	401.15
P2	2-hydroxy-4-(((2-methyl-1H-benzo[d]imidazol-1-yl)methyl)amino)benzoic acid	139–140	80.90	N-H: 3595 OH: 3498 COOH: 1718 C=N: 1612 C-N: 1254	–	–
P3	4-(((2-(2-aminophenyl)-1H-benzo[d]imidazol-1-yl)methyl)amino)-2-hydroxybenzoic acid	273–276	65.80	N-H: 3352 OH: 3479 COOH: 3294 C=N: 1709 NH ₂ : 1277	CH ₂ : 5.36 (2H, s) Ar-H: 6.30–7.91	583.23
P4	4-(((2-(2,4-dinitrophenyl)-1H-benzo[d]imidazol-1-yl)methyl)amino)-2-hydroxybenzoic acid	215–220	78.20	N-H: 3471 OH: 3633 COOH: 3386 C=N/C=C: 1643 NO ₂ : 1516, 1342	–	–
P5	4-(((2-(2-chloroethyl)-1H-benzo[d]imidazol-1-yl)methyl)amino)-2-hydroxybenzoic acid	287–290	83.70	N-H: 3305 OH: 3629 COOH: 3379 C=N: 1631 CH ₂ Cl: 1234	–	–
P6	2-hydroxy-4-(((2-(4-nitrophenyl)-1H-benzo[d]imidazol-1-yl)methyl)amino)benzoic acid	215–219	62.50	N-H: 3236 OH: 3471 COOH: 3143 C=N: 1519 NO ₂ : 1560	CH ₂ : 5.43 (2H, s) Ar-H: 6.30–8.27	404.11

Molecular docking

Molecular docking was carried out using AutoDock Vina v1.5.6 [46], which enables rigid and flexible ligand docking within defined protein binding sites. The receptor structure was obtained from the Protein Data Bank or modeled through homology approaches. The primary goal was to predict optimal ligand–receptor binding conformations with minimum interaction energy, evaluated by scoring functions based on steric and electrostatic interactions [47].

Ligand structures of the synthesized benzimidazole derivatives were drawn in ChemDraw, converted into 3D models, and energy-minimized. Conformers were generated, and the lowest-energy conformer for

each compound was selected. Final structures were refined in PyMOL and saved in .pdb format for docking studies [47].

Preparation of ligands

Ligand structures were designed and optimized in ChemDraw Ultra 12.0, saved in .mol format, and converted to .pdb using PyMOL. Prior to docking in AutoDock Vina v1.5.6, water molecules and native ligands were removed from the target enzyme. Hydrogen atoms, torsions, and rotatable bonds were assigned, and the final ligand files were saved in .pdbqt format.

Preparation of protein

The crystal structure of FtsZ from *Mycobacterium tuberculosis* complexed with GTP- γ -S (PDB ID: 2Q1Y) was retrieved from the RCSB Protein Data Bank [48]. The structure was processed in Discovery Studio, where water molecules and the native ligand were removed, hydrogen atoms were added, and the binding site was defined before saving the receptor in .pdb format.

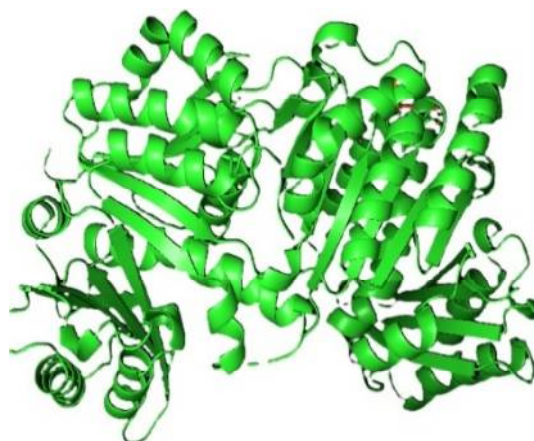


Figure 4: Crystal structure of the cell division protein FtsZ from *Mycobacterium TB* in association with GTP-gamma-S (PDB ID: 2Q1Y)

Anti- Myobacterial activity

Antitubercular Activity Evaluation

The antitubercular activity of synthesized compounds was evaluated by the Microplate Alamar Blue Assay (MABA) against *Mycobacterium tuberculosis* H37Rv [49–51]. Test compounds were dissolved, serially diluted in Middlebrook 7H9 broth (with OADC and Tween 80), and incubated with bacterial suspension in 96-well plates at 37 °C for 7 days. After addition of Alamar Blue and 24 h incubation, fluorescence was measured, and the minimum inhibitory concentration (MIC) was defined as the lowest concentration inhibiting $\geq 90\%$ of growth. Rifampicin and para-aminosalicylic acid were used as positive controls.

RESULT AND DISCUSSION

Chemistry

2-substituted benzimidazoles were synthesized by refluxing *o*-phenylenediamine with substituted carboxylic acids in 4N HCl at 70–80 °C, monitored by TLC. Final derivatives were obtained by coupling these intermediates with para-amino salicylic acid using K_2CO_3 in ethanol, followed by recrystallization. All compounds were characterized by IR, NMR, and mass spectrometry.

Molecular docking study

All synthesized derivatives (P1–P6) were evaluated for anti-mycobacterial activity. Docking results (Table 4) showed compound P4 with the highest binding affinity (–8.9) against the FtsZ receptor (PDB ID: 2Q1Y). Receptor–ligand interactions of P4, PAS, and isoniazid are illustrated in 3D (Figures 5, 7, 9) and 2D (Figures 6, 7, 10) representations.

Table 4: Molecular docking results of benzimidazole derivatives showing Interaction for their anti-tubercular activity

Sr. No.	Log P	Docking Score Kcal/mol	Interaction
P1	1.78	-7.5	ASP A:84, GLU B:136 & GLY B:19 (CHB), GLY B:104 (Amide-Pi stacked), GLU A:87 (Pi-Anion)
P2	2.45	-6.7	ARG B:139 (CHB), ALA B:144 (Pi-Sigma), GLY B:103 (Amide Pi-stacked), PRO B:132 (Pi-Alkyl)
P3	3.03	-7.5	ASN B:22 (CHB), ARG B:126 & GLU A:36 (Pi- Cation/ Pi-Anion), ARG B:181 (Pi-Alkyl), VAL A:10 (Pi-Sigma)
P4	4.4	-8.9	ARG B:304, SER B:260 & THR B:306 (CHB), ALA B:262 (Pi-Alkyl)
P5	3.36	-6.9	ARG B:304 & VAL B:305 (CHB), GLN B: 192 (Halogen), SER B:227 (Carbon-hydrogen bond), ALA B:262& ILE B:225 (Pi-Alkyl)
P6	4.6	-7.4	ILE A:11 & GLU A:36 (CHB), ASP A:53 & GLU B:29 (Pi-Anion), VAL A:10 (Pi-Alkyl & Carbon-hydrogen bond)
PAS	1.012	-4.7	ASP B:43 (pi-anion), ALA B:70 (pi-alkyl), GLY B:104, GLY B:19, GLY B:107, THR B:106 & ASP A:81 (CHB), GLY B:105 & GLY B:17 (C-H Bond)
Isoniazid	-0.64	-5.5	ALA B:70 (pi-alkyl), ASP B:43 (pi-anion), GLY B:18 (Van der waals), GLY B:107 & THR B:106 (CHB)

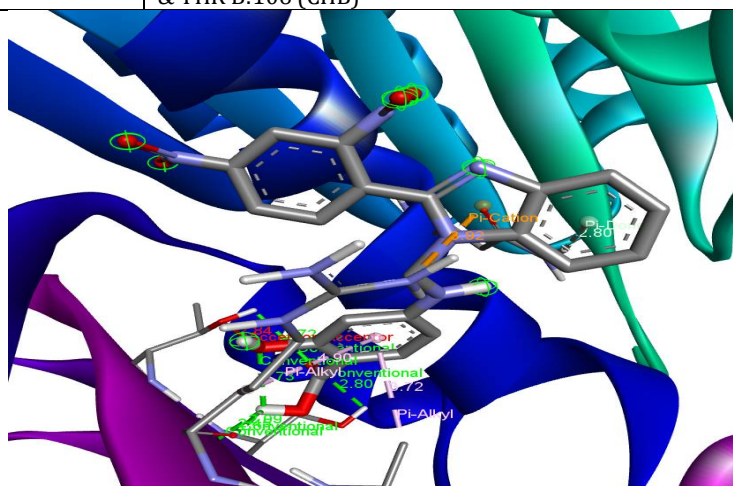


Figure 5: 3D receptor–ligand interaction of compound P4 with the FtsZ protein (PDB ID: 2Q1Y)

3D view of compound P4 bound to FtsZ protein (PDB ID: 2Q1Y). Key hydrogen bonds (dashed lines) and hydrophobic interactions are shown, illustrating the ligand's binding mode.

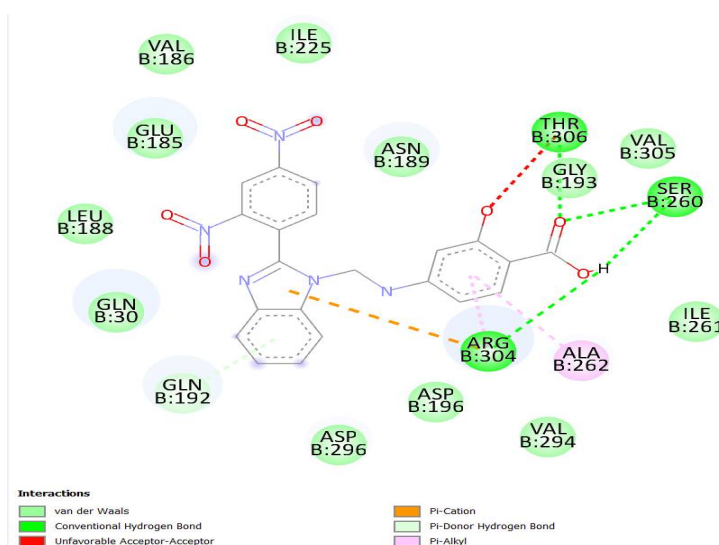


Figure 6: 2D receptor–ligand interaction of compound P4 with the FtsZ protein (PDB ID: 2Q1Y)

2D map showing key interactions of compound P4 with FtsZ protein (PDB ID: 2Q1Y), including hydrogen bonds and hydrophobic contacts.

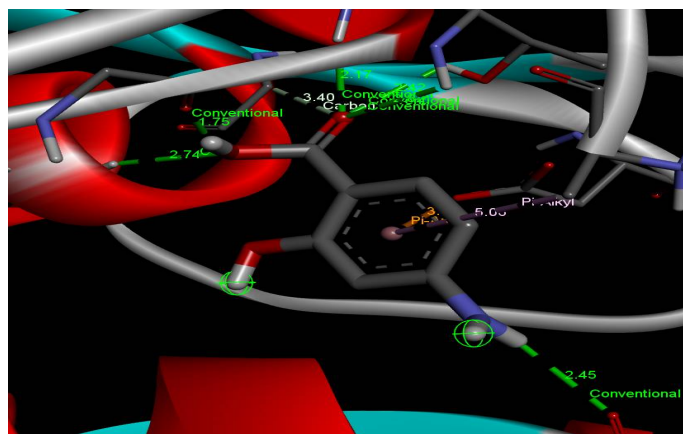


Figure 7: 3D receptor–ligand interaction of para-aminosalicylic acid (PAS) with the FtsZ protein (PDB ID: 2Q1Y)

3D view of PAS bound to FtsZ protein (PDB ID: 2Q1Y), showing key hydrogen bonds and hydrophobic interactions in the active site.

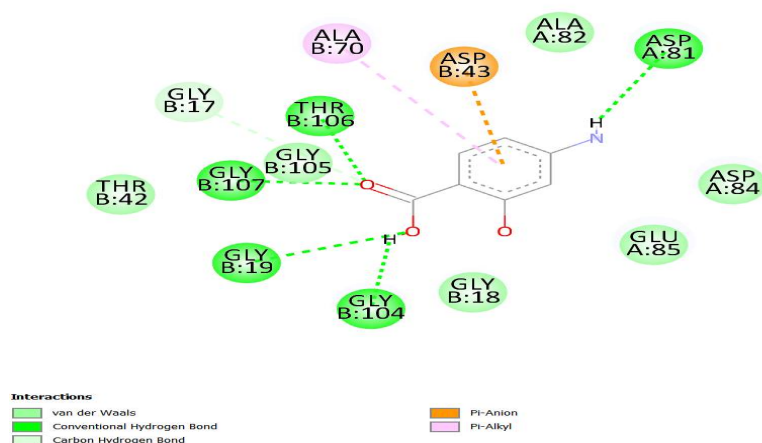


Figure 8: 2D receptor–ligand interaction of para-aminosalicylic acid (PAS) with the FtsZ protein (PDB ID: 2Q1Y)

Two-dimensional representation of para-aminosalicylic acid (PAS) interactions with FtsZ protein (PDB ID: 2Q1Y), highlighting hydrogen bonds, hydrophobic contacts, and key ligand–residue interactions in the binding site.

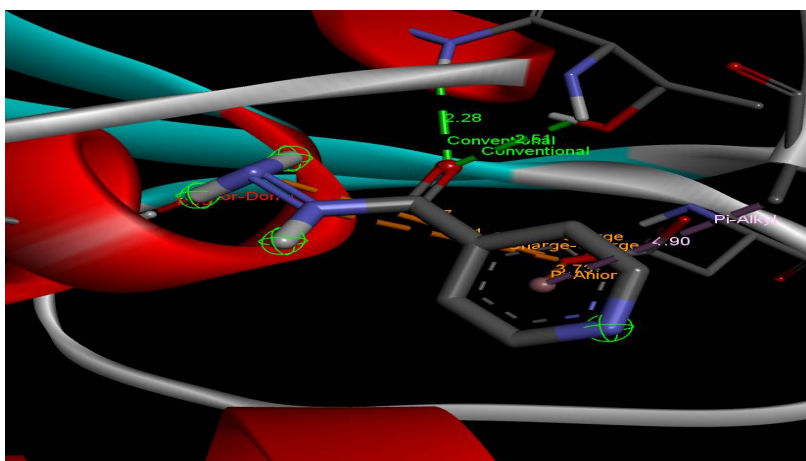


Figure 9: 3D receptor–ligand interaction of isoniazid with the FtsZ protein (PDB ID: 2Q1Y)

Three-dimensional representation of isoniazid bound to FtsZ protein (PDB ID: 2Q1Y). Hydrogen bonds are shown as dashed lines, and hydrophobic interactions are indicated, illustrating the binding mode of isoniazid within the active site.

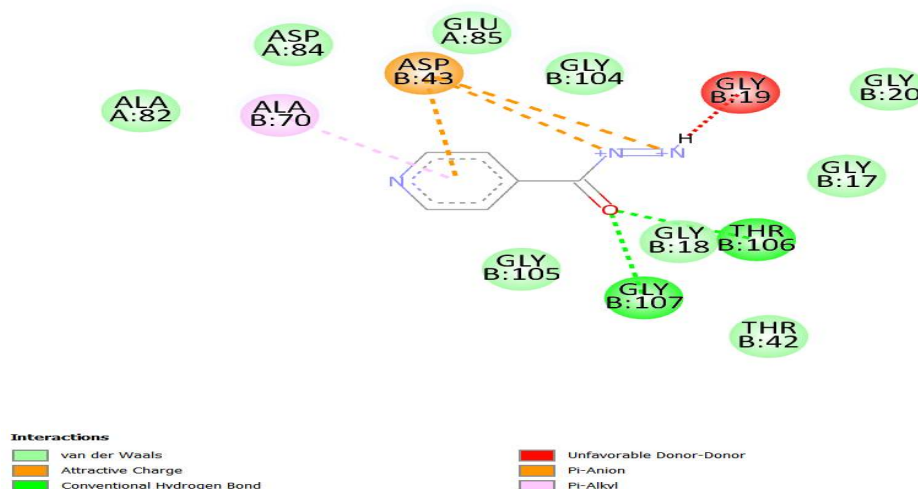


Figure 10: 2D receptor-ligand interaction of isoniazid with the FtsZ protein (PDB ID: 2Q1Y)

Two-dimensional representation of isoniazid interactions with FtsZ protein (PDB ID: 2Q1Y), showing hydrogen bonds, hydrophobic contacts, and key ligand-residue interactions in the binding site.

Pharmacological evaluation

The Microplate Alamar Blue Assay (MABA) was conducted at Andhra University, Visakhapatnam. Due to poor solubility, compounds were dissolved in DMSO for solution preparation. MIC values were determined and compared with standard drugs (rifampicin and PAS), using DMSO as a positive control.

- Each drug's test solution was added to a 96-well microplate containing Alamar Blue Reagent.
- Fluorescence was quantified with a microplate reader to determine the minimum inhibitory concentration (MIC).

The anti-tubercular activity of the synthesized derivatives was assessed using MABA (Figure 11), and the MIC values are summarized in Table 5. A comparative activity profile with PAS and Rifampicin is shown in Figure 12.

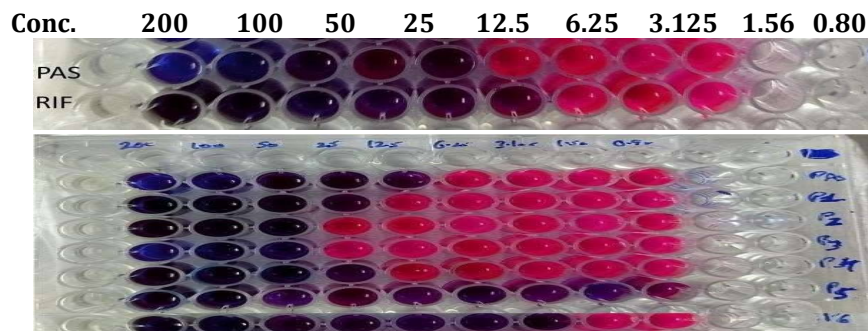


Figure 11: Microplate Alamar Blue Assay (Anti-tubercular activity results)

Table 5: Microplate Alamar Blue Assay

Sr. No.	Comp. Code	MIC (µg/ml) H ₃₇ R _v Strain
1.	P1	25
2.	P2	50
3.	P3	50
4.	P4	25
5.	P5	1.56
6.	P6	3.125
7.	PAS	12.5
8.	Rifampicin	6.25

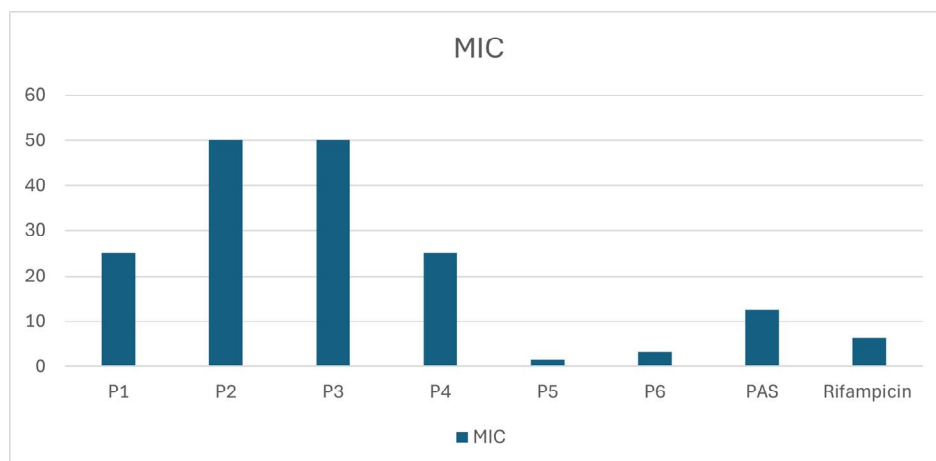


Figure 12: Graphical representation showing activity of derivatives (MIC values) with respect to PAS and Rifampicin

CONCLUSION

Tuberculosis remains a major global health challenge, demanding the discovery of new and effective therapeutics. In this study, benzimidazole derivatives were synthesized and evaluated, demonstrating significant anti-mycobacterial activity against the H37Rv strain. Molecular docking revealed strong affinity toward the FtsZ protein (PDB: 2Q1Y), with derivative P4 exhibiting superior binding compared to standard drugs such as PAS and isoniazid. Importantly, all derivatives displayed higher docking scores than conventional therapies, suggesting their potential as next-generation anti-TB agents. The Microplate Alamar Blue Assay further confirmed their bioactivity, establishing these compounds as promising scaffolds for rational drug design. Collectively, these findings not only highlight benzimidazole derivatives as valuable leads but also underscore their potential contribution to combating the escalating global TB burden.

ACKNOWLEDGEMENT

The authors are very thankful to the management of S.M.B.T. College of Pharmacy for their constant support and providing all facilities to complete this work.

Conflict Of Interest

The authors declare that they have no conflict of interest.

Ethical Approvals

This study does not involve experiments on animal or human subjects.

Data Availability Statement

All data which are generated and analyzed are included within this research article.

Author Contribution Statement

P. N. S. performed the synthesis, characterization, biological evaluation, computational studies, data analysis, and manuscript preparation. **M. R. K.** supervised the research, critically reviewed the manuscript, and approved the final version.

REFERENCES

1. Tuberculosis (online), <http://www.wrongdiagnosis.com/t/tuberculosis>, 26TH 2011.
2. Konstantinos A. (2010). Testing for tuberculosis. *Aust Prescr.* 33:12–8.
3. Grange JM. (2009). Tuberculosis: a comprehensive clinical reference. In: Schaaf S, Zumla AL, editors. Saunders; p. 44–59.
4. Dye C, Scheele S, Dolin P, Pathania V, (1999). Raviglione MC. World health organization (WHO). Global surveillance and monitoring project. *JAMA*; 282:677–86.
5. Blevins SM, Bronze MS. (2010). Robert Koch and the ‘golden age’ of bacteriology. *Int J Infect Dis* ;14:e 744–51.
6. Umesiri FE, Sanki AK, Boucau J, Ronning DR, Sucheck SJ. (2010). Recent advances toward the inhibition of mAG and LAM synthesis in *Mycobacterium tuberculosis*. *Med Res Rev*; 30:290–326.

7. Panteix G, Gutierrez MC, Boschirolu ML, Rouviere M, Plaidy A, Pressac D, et al. (2010). Pulmonary tuberculosis due to *Mycobacterium microti*: a study of six recent cases in France. *J Med Microbiol*; 59:984–9.
8. Sampaio EP, Elloumi HZ, Zelazny A, Ding L, Paulson ML, Sher A, et al. (2008). *Mycobacterium abscessus* and *M. avium* trigger toll-like receptor 2 and distinct cytokine response in human cells. *Am J Resp Cell Mol Biol*; 39:431–9.
9. Kaufmann SHE. (2003). A short history of Robert Koch's fight against tuberculosis: those who do not remember the past are condemned to repeat it. *Tuberculosis* ; 83:86–90.
10. Zumla A, Nahid P, Cole ST. (2013). Advances in the development of new tuberculosis drugs and treatment regimens. *Nat Rev Drug Discov* ; 12:388–404.
11. Dover LG, Coxon GD. (2011). Current status and research strategies in tuberculosis drug development. *J Med Chem*; 54:6157–65.
12. Bloom BR, Murray CJL. (1992). Tuberculosis: commentary on a reemerging killer. *Science*; 257:1055–64.
13. Okada M, Kobayashi K. (2007). Recent progress in mycobacteriology. *Kekkaku*; 82:783–99.
14. World Health Organization Report on TB epidemic, Global TB Programme, World Health Organization Geneva, (1997).
15. World health organisation, Tuberculosis (2007); Fact Sheet No.104.
16. World health organization. Global tuberculosis report. (2013).
17. Kaufmann SHE, Parida SK. (2008). Tuberculosis in Africa: learning from pathogenesis for biomarker identification. *Cell Host Microb*; 4:219–28.
18. Harper C. (2007). Tuberculosis, a neglected opportunity. *Nat Med*; 13:309–12.
19. Laughon BE. (2007). New tuberculosis drugs in development. *Curr Top Med Chem*; 7:463–73.
20. Becerra MC, Bayona J, Freeman J, Farmer PE, Kim JY. (2000). Redefining MDR-TB transmission 'hotspots. *Int J Tuberc Lung Dis*; 4:387–94.
21. Ramprasad J, Nayak N, Dalimba U, Yogeewari P, Sriram D, Peethambar SK, Achur R, Kumar HSS (2015). Synthesis and biological evaluation of new imidazo[2,1-b][1,3,4] thiadiazole-benzimidazole derivatives. *Eur J Med Chem* 95:49–63
22. Yoon YK, Ali MA, Wei AC, Choon TS, Ismail R (2015). Synthesis and evaluation of antimycobacterial activity of new benzimidazole aminoesters. *Eur J Med Chem* 93:614–624
23. Camacho J, Barazarte A, Gamboa N, Rodrigues J, Rojas R, Vaisberg A, Gilman R, Charris J, (2011). Synthesis and Evaluation of Antitubercular activity of N'-substituted benzo[d]imidazole-5- carbohydrazide Derivatives, *Bioorg Med Chem*, 19, 20-27.
24. Gong Y, Karakaya S S, Guo X, Zheng P, Gold B, Ma Y, Little D, Roberts J, Warriar T, Jiang X, Pingle M, Nathan C F, Liu G, (2014). Synthesis and Evaluation of Anti-tubercular activity of Substituted Benzimidazole, *Eur J Med Chem*, 75, 336.
25. Yoon YK, Ali MA, Wei AC, Shirazi AN, Parang K, Choon TS. (2014). Benzimidazoles as new scaffold of sirtuin inhibitors: Green synthesis, in vitro studies, molecular docking analysis and evaluation of their anti-cancer properties. *European journal of medicinal chemistry*. 18; 83:448-54.
26. Debus H, *Annals of Chemistry and Pharmacy*. 1858, 107(2), 199. <https://doi.org/10.1002/jlac.18581070209>
27. Ingle RG, Magar DD. (2011). Heterocyclic chemistry of benzimidazoles and potential activities of derivatives. *Int. J. Drug Res. Tech*. 1(1):26-32.
28. Tahlán S, Ramasamy K, Lim SM, Shah SA, Mani V, Narasimhan B. (2019). 4-(2-(1H-Benzo [d] imidazol-2-ylthio) acetamido)-N-(substituted phenyl) benzamides: design, synthesis and biological evaluation. *BMC chemistry*. ;13(1):1-6.
29. Wang Z., Deng X., Xiong S., Xiong R., Liu J., Zou L., Lei X., Cao X., Xie Z., Chen Y., Liu Y., Zheng X., Tang G., (2018). Design, synthesis and biological evaluation of chrysin benzimidazole derivatives as potential anticancer agents, *Nat. Prod. Res*. 32 (24). 2900-2909.
30. Morais G.R., Palma E., Marques F., Gano L., M. Oliveira C, Abrunhosa A., H. Miranda V., Outeiro T.F., Santos L., Paulo A., (2017). Synthesis and biological evaluation of novel 2-aryl benzimidazoles as chemotherapeutic agents, *J. Heterocyclic Chem*. 54 (1), 255-267
31. Shaker Y.M., Omar M.A., Mahmoud K., Elhallouty S.M., El-Senousy W.M., All M. M., Mahmoud A, E., Abdel-Halim A.H., Soliman S.M., El Diwani HL, Synthesis, *In vitro* and *In vivo* antitumor and antiviral activity of novel 1-substituted benzimidazole derivatives, *J. Enzyme Inhib. Med. Chem*. 30 (5) (2015) 826-845.
32. Onnis V., Demurtas M., Deplano A., Balboni G., Baldisserotto A., Manfredini S., Pacifico S., Lickens S., Balzarini J., Design, synthesis and evaluation of antiproliferative activity of new benzimidazolehydrazones, *J. Balzarini Mol*. 21 (5) (2016) 579-588.
33. Acar Çevik U., Sağlık B.N., Korkut B., Özkay Y., İlgin S, (2018). Antiproliferative cytotoxic, and apoptotic effects of new benzimidazole derivatives bearing hydrazone moiety, *J. Heterocyclic Chem*. 55 (1) : 138-148
34. Hanan M. Refaat et al., (2010). Synthesis and anticancer activity of some novel 2-substituted benzimidazole derivatives *European Journal of Medicinal Chemistry* 45; 2949e2956.
35. Özden Ö, Zengin G, et al., (2007). Synthesis and antimicrobial activity of some novel phenyl and benzimidazole substituted benzyl ethers *Bioorganic & Medicinal Chemistry Letters* 17; 2233– 2236
36. Weijie Si, Tao Z, Yaofa Li, Dongmei She, Wenliang Pan, Zhanlin Gao, Jun Ning and Xiangdong Mei, (2016). Synthesis and biological activity of novel benzimidazole derivatives as potential antifungal agents, *Journal of Pesticide Science*, 41(1), 15–19.

37. Alasmary F A. S., Snelling A M., Zain M E., Alafeefy A M., Awaad A S. and Karodia N, (2015). Synthesis and Evaluation of Selected Benzimidazole Derivatives as Potential Antimicrobial Agents, *Molecules*, 20, 15206-15223.
38. Alanazi A.H., Alam Md. T., Imran M., (2017). Design, molecular docking studies, in silico drug likeliness prediction and synthesis of some benzimidazole derivatives as antihypertensive agents, *Md. Imranl. Indo American J. of Pharmaceutical Sci.* 4 (04); 926–936.
39. Kankate R., Pangare A., Kakad R., Gide P., Nathe V., (2016). Synthesis and biological evaluation of benzimidazolyl biphenyl derivatives as antihypertensive agents, *Int. J. Chem. Concepts* 2 (2); 111–119.
40. Correia M, Rodrigues M, Paiga P, Delerue-Matos D, (2016). Fungicides. in *Encyclopedia of Food and Health*. Academic Press, Cambridge, MA, p. 169. <https://doi.org/10.1016/b978-0-12-384947-2.00342-1>
41. Tonelli M, Simone M, Tasso B, Novelli F, Boido V, Sparatore F, Paglietti G, Pricl S, Giliberti G, Blois S, Ibba C. (2010). Antiviral activity of benzimidazole derivatives. II. Antiviral activity of 2-phenylbenzimidazole derivatives. *Bioorganic & medicinal chemistry*.15;18(8):2937-53.
42. Wright, J.B., (1951), The chemistry of benzimidazoles, chemical review, 48(3), pp, 397-541.
43. Preston PN. (1981). Benzimidazoles, *Chemistry of Heterocyclic Compounds: Part 1*. 1; 40:1-285.
44. Chen AY, Yu C, Bodley A, Peng LF, Liu LF. (1993). A new mammalian DNA topoisomerase I poison Hoechst 33342: cytotoxicity and drug resistance in human cell cultures. *Cancer research*. 15;53(6):1332-7
45. Furniss B.S., Hannaford A.J., Smith P.W.G., Tatchell A.R. (1999). "Vogel's Textbook of Practical Organic Chemistry", 5th Edition, Pg.No.1162.
46. Fierz-David H.E. (2000). "The Fundamental Processes of dye Chemistry", 153-158.
47. Trott O., Olson J., (2010). AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading, *Journal of Computational Chemistry* 31, 455-461 DOI 10.1002/jcc.21334
48. RCSB. Protein data bank; Available from: <https://www.rcsb.org/>
49. Gadad A.K., Noolvi N.N., Rajshekhar V.K., (2004). *Bioorg. Med. Chem.* 12; 5651–5659
50. Yajko D.M., Madej J.J., Gee B., Babst A., Keith Hardley W., (1995). *J. Clin. Microbiol.* 33; 2324–2327.
51. Suling W.J., Seitz L.E., Pathak V., Westbrook L., Barrow E.W., Zywno-vanginkel S., Renolds R.C., Piper J.R., Barrow W.W., (2000). Antimicrobial. Agents. *Chemother.* 44 : 2784–2793.

Copyright: © 2025 Author. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.